

Available online at www.sciencedirect.com

# **ScienceDirect**

journal homepage: www.e-jmii.com



# Original Article



Rui-Xin Wu<sup>a</sup>, Ching-Mei Yu<sup>b</sup>, Sung-Teng Hsu<sup>c</sup>, Ching Hsun Wang<sup>d,\*</sup>

<sup>a</sup> Division of Infectious Diseases and Tropical Medicine, National Defense Medical Center, Tri-Service General Hospital Penghu Branch, Penghu, Taiwan

<sup>b</sup> Department of Clinical Pathology, National Defense Medical Center, Tri-Service General Hospital, Taiwan <sup>c</sup> Infection Control Office, Tri-Service General Hospital, National Defense Medical Center, Taipei, Taiwan

<sup>d</sup> Division of Infectious Diseases and Tropical Medicine, Department of Internal Medicine, Tri-Service General Hospital, National Defense Medical Center, Taipei, Taiwan

Received 27 September 2020; received in revised form 17 November 2020; accepted 15 December 2020 Available online 13 January 2021

#### **KEYWORDS**

Risk; Resistance; Stenotrophomonas maltophilia, levofloxacin; Trimethoprim/ sulfamethoxazole Abstract Background: The emergence of concurrent levofloxacin- and trimethoprim/sulfamethoxazole (TMP/SMX)-resistant Stenotrophomonas maltophilia (LTSRSM) in Taiwan is becoming a serious problem, but clinical data analysis on this has not been reported. *Methods:* A matched case-control-control study was conducted to investigate risk factors for LTSRSM occurrence in hospitalized patients. For patients with LTSRSM infection/colonization (the case group), two matched control groups were used: control group A with levofloxacinand TMP/SMX-susceptible S. maltophilia (LTSSSM) and control group B without S. maltophilia. Besides, tigecycline, ceftazidime, cefepime, ciprofloxacin, gentamicin, amikacin, and colistin susceptibilities in collected LTSRSM and levofloxacin- and TMP/SMX-susceptible S. maltophilia (LTSSSM) isolates were compared. *Results:* From January 2014 to June 2016, 129 LTSRSM from cultured 1213 S. maltophilia isolates (10.6%) were identified. A total of 107 LTSRSM infected patients paired with 107 LTSSSM-, and 107 non-S. maltophilia-infected ones were included. When compared with control group A, previous fluoroquinolone and TMP/SMX use was found to be independently

\* Corresponding author. Division of Infectious Diseases and Tropical Medicine, Department of Internal Medicine, Tri-Service General Hospital, National Defense Medical Center, No. 325, Section 2, Cheng-Kung Road, Neihu, 114, Taipei, Taiwan. Fax: +886 2 87927258. *E-mail address:* sasak0308@gmail.com (C.H. Wang).

#### https://doi.org/10.1016/j.jmii.2020.12.012

1684-1182/Copyright © 2021, Taiwan Society of Microbiology. Published by Elsevier Taiwan LLC. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

associated with LTSRSM occurrence. When compared with control group B, mechanical ventilation, cerebrovascular disease, and previous fluoroquinolone use were risk factors for LTSRSM occurrence. Eighty-five LTSRSM and 85 LTSSSM isolates were compared for antibiotic susceptibilities; the resistance rates and minimum inhibitory concentrations of tigecycline and ceftazidime were significantly higher for LTSRSM than for LTSSSM isolates.

*Conclusion:* The emergence of LTSRSM showing cross resistance to tigecycline and ceftazidime would further limit current therapeutic options. Cautious fluoroquinolone and TMP/SMX use may be helpful to limit such high-level resistant strains of *S. maltophilia* occurrence.

Copyright © 2021, Taiwan Society of Microbiology. Published by Elsevier Taiwan LLC. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

#### Introduction

Stenotrophomonas maltophilia, a non-fermenting gramnegative bacterium, is an important nosocomial pathogen in hospital settings.<sup>1</sup> In recent years, several worldwide surveillance studies have detected increasing infection rates of this bacterium in an expanding population of patients who were immunocompromised due to advances in medical technologies and treatments.<sup>2</sup> Although of low virulence, S. maltophilia can cause a wide range of infections including the respiratory system, bloodstream, skin and soft tissue, bone and joints, biliary tract, and urinary tract in immunocompromised patients.<sup>3</sup> Treatment of these infections is difficult because S. maltophilia exhibits extensive resistance to a variety of antibiotics.<sup>2</sup> Different resistance mechanisms, including inducible βlactamases L1 and L2, aminoglycoside-modifying enzymes, and overexpression of multidrug efflux pumps, render S. maltophilia resistant to multiple structurally unrelated antibiotics.<sup>2</sup> As a consequence, the World Health Organization has listed S. maltophilia as one of the multidrugresistant bacteria in hospital settings.<sup>4</sup> Trimethoprim/sulfamethoxazole (TMP/SMX) is considered the drug of choice for treating susceptible S. maltophilia infections and has been widely used for many years based upon reported in vitro activity and favourable clinical outcomes.<sup>5,6</sup> As alternatives to TMP/SMX, fluoroquinolones may be considered for patients infected with TMP/SMX-resistant S. maltophilia or those intolerant to TMP/SMX due to adverse drug effects. Recent studies comparing treatments with fluoroquinolones and TMP/SMX have suggested that fluoroquinolones have similar efficacy but fewer adverse drug effects than TMP/SMX.<sup>7,8</sup> Nevertheless, the clinical efficacy of fluoroquinolones other than levofloxacin still needs to be validated due to the limited number of clinical studies reported and the current lack of clinical breakpoints.

Although the resistance rates to levofloxacin or TMP/SMX vary geographically, longitudinal global surveillance reports have revealed that this resistance has generally remained less than 10%.<sup>9</sup> In recent years, increased resistance rates of *S. maltophilia* to TMP/SMX or levofloxacin have been observed in several countries from local surveillance reports.<sup>10–15</sup> In Taiwan, a more worrisome phenomenon was noted in which resistance rates of *S. maltophilia* to both TMP/SMX and levofloxacin were elevated in nation-wide surveillance reports.<sup>16</sup> Furthermore, analysing

antibiotic susceptibilities of such resistant strains revealed that TMP/SMX-resistant S. maltophilia isolates often exhibited concurrent levofloxacin resistance and vice versa.<sup>16–19</sup> This feature of S. maltophilia, where mutationdriven resistance to one antibiotic results in crossresistance to others, has also been observed in experimental evolution studies under laboratory settings.<sup>20</sup> The emergence of such strains showing both levofloxacin and TMP/SMX resistance would further limit therapeutic options in treating S. maltophilia infections, and this may become a public health concern in the future. This alarming phenomenon underscores an urgent need for effective control and prevention measures that could combat further dissemination. We therefore initiated a matched casecontrol-control study to identify risk factors for concurrent levofloxacin- and TMP/SMX-resistant S. maltophilia (LTSRSM) infections among hospitalized patients. Moreover, the antibiotic susceptibilities of resistant and susceptible strains were analysed and compared.

#### Methods

#### Study setting, design, and patient identification

This study was conducted at the Tri-Service General Hospital, which is a medical center with 1800 beds located in northern Taiwan. The study period was from January 2014 to June 2016. The approval of Institutional Review Board of the hospital (TSGHIRB approval number: 2-101-05-074). To assess risk factors for LTSRSM occurrence, a 1:1:1 matched case-control-control design was used. Adult inpatients (>18 years old) infected/colonized with LTSRSM and those infected/colonized with levofloxacin- and TMP/SMXsusceptible S. maltophilia (LTSSSM) were identified via a computerized medical records system and classified as the case group and control group A, respectively. Patients infected/colonized with S. maltophilia that showed intermediate resistance to levofloxacin were categorized in the case group for analysis. For patients with multiple episodes of S. maltophilia infection/colonization, only the first was included. The control group B patients were selected from the inpatient population without S. maltophilia during their hospital stay. Patients with S. maltophilia isolated less than 48 h after admission or those younger than 18 years of age were excluded. The case group was matched to control group A by age (within 5 years), sex, and the site of isolation. The criteria for matching the case group to control group B included age (within 5 years), sex, and time at risk. Once several eligible controls were identified, they were randomly chosen using Microsoft Excel 2013® software (Microsoft Corp., Redmond, WA, USA).

#### Relevant clinical data and definitions

The clinical information from identified patients was retrieved from a computerized medical records system. Possible risk factors were recorded including the following: age, sex, time at risk, recent admission and intensive care unit (ICU) records, recent chemotherapy and surgery, comorbidities, previous antimicrobial exposure, and indwelling medical devices. In the case group and control group A, the time at risk was defined as the number of days elapsed from patient admission to the date of the first S. maltophilia isolation. In control group B, the time at risk was defined as the number of days from patient admission to discharge. Previous antibiotic exposure in the case group and control group A was defined as at least 24 h of therapy within two weeks before S. maltophilia isolation. In control group B, previous antibiotic exposure was defined as 24 h of therapy within two weeks prior to discharge. Recent therapeutic measures were defined as having occurred 30 days before S. maltophilia isolation in patients in the case group and control group A; in control group B, recent therapeutic measures were defined as having occurred 30 days before index discharge. Indwelling invasive medical devices used before S. maltophilia isolation in the case group and control group A were recorded. For control group B, indwelling invasive medical devices used before discharge were recorded.

#### Microbiologic methods

We analysed the antibiotic susceptibilities of 85 consecutive, non-duplicated preserved isolates of LTSRSM from the respiratory tract and preserved 85 LTSSSM isolates randomly chosen from the same source as LTSRSM in hospitalized patients during the study period for comparison. The identification of S. maltophilia isolates was performed by matrix-assisted laser desorption/ionization time-offlight mass spectrometry (bioMérieux Inc., Marcy-l'Etoile, Rhône, France). The minimum inhibitory concentrations (MICs) of identified S. maltophilia isolates were determined using the VITEK 2 automated system (bioMérieux Inc.). The breakpoints for TMP/SMX, levofloxacin, and ceftazidime for S. maltophilia were established according to 2020 Clinical and Laboratory Standards Institute (CLSI) criteria. The clinical breakpoints for tigecycline were established according to the 2020 European Committee on Antimicrobial Susceptibility Testing for Enterobacterales. The breakpoints for other tested antibiotics including ciprofloxacin, cefepime, colistin, gentamicin, and amikacin were established according to the 2020 CLSI criteria for Pseudomonas aeruginosa.

#### Statistical analysis

Continuous variables are presented as means  $\pm$  standard deviations (SDs), and we used t-tests or Mann-Whitney U tests for comparisons as appropriate. Categorical variables are presented as numbers and percentages and compared using the chi-square test or Fisher's exact test. Variables with p values < 0.05 on bivariate analysis were included in a forward stepwise conditional logistic regression model for multivariate analysis. Two simultaneous multivariate models were produced from the data obtained. The first model used the pairs of the case group and matched control group A, while the second model used the pairs of the case group and matched control group B. All tests were 2-tailed, and a p value of <0.05 was considered statistically significant. All results were analysed using a commercially available software package (SPSS, version 16.0; SPSS Inc., Chicago, IL, USA).

## Results

During the study period from January 2014 to June 2016, 1213 patients were identified as having cultures positive for S. maltophilia. Of these patients, 593 were hospitalized in the ICU and the others were admitted to the general ward. The most common isolation source was from the respiratory tract (1038/1213, 85.5%), followed by the blood (62/1213, 5.1%). One hundred and twenty-nine of the 1213 S. maltophilia isolates exhibited concurrent levofloxacin- and TMP/SMX resistance (10.6%). Among the 129 patients with LTSRSM, we excluded 15 patients with LTSRSM isolated within 48 h after admission, and seven unsuccessfully matched cases. Consequently, the case group consisted of 107 patients with LTSRSM. Among 107 cases in the case group with LTSRSM, the isolation source were from the respiratory tract (98/107, 91.5%), followed by the wound tissue (6/107, 5.6%), blood (2/107, 1.8%) and the urinary tract (1/107, 0.9%). After matching, control group A consisted of 107 patients with LTSSSM and control group B consisted of 107 patients randomly identified from the inpatient population without S. maltophilia infections were included for comparisons to determine independent risk factors for LTSRSM occurrence.

# Risk factors for LTSRSM occurrence among hospitalized patients

As shown in Table 1, when the case group was compared to the control group A, there were no statistically significant between-group differences in time at risk, recent admission records, recent ICU admission, recent chemotherapy and surgery, comorbidities, and indwelling medical devices (p > 0.05). Previous antibiotic exposure revealed that the case group was more likely than the controls to have been exposed to fluoroquinolone and TMP/SMX (43.0% vs. 7.5% and 12.1% vs. 1.9%, p < 0.001 and 0.003, respectively). When the case group was compared to control group B, the

Risk factor, n (%)	Case group	Control group A	P <sup>a</sup>	Control group B	P <sup>b</sup>
, , , , , , , , , , , , , , , , , , ,	(n = 107)	(n = 107)	value	(n = 107)	value
Age in years <sup>c</sup>	77.0 (22.0)	77.0 (22.0)	0.964	75.0 (25.0)	0.999
Male.	73 (68.2)	73 (68.2)	-	73 (68.2)	1.000
Time at risk (days) <sup>c,d</sup>	23 (23)	18 (19)	0.085	23 (23)	1.000
Recent admission records in the last 3 months	28 (26.2)	35 (32.7)	0.294	38 (35.5)	0.139
Prior ICU admission	89 (83.2)	87 (81.3)	0.721	32 (29.9)	<0.001
Chemotherapy in the last month	5 (4.7)	2 (1.9)	0.445	14 (13.1)	0.031
Surgery in the last month	59 (55.1)	53 (49.5)	0.412	40 (37.4)	0.009
Comorbidities	<b>、</b>	· · · ·			
Cerebrovascular disease	43 (40.2)	34 (31.8)	0.200	24 (22.4)	0.005
Dementia	15 (14.0)	20 (18.7)	0.355	10 (9.3)	0.287
Heart failure	19 (17.8)	17 (15.9)	0.715	11 (10.3)	0.115
Chronic renal insufficiency	40 (37.4)	29 (27.1)	0.108	28 (26.2))	0.078
Chronic lung disease	13 (12.1)	12 (11.2)	0.831	9 (8.4)	0.368
Liver cirrhosis	4 (3.7)	4 (3.7)	1.000	9 (8.4)	0.152
Diabetes mellitus	38 (35.5)	41 (38.3)	0.671	39 (36.4)	0.887
Autoimmune disease	8 (7.5)	6 (5.6)	0.580	3 (2.8)	0.122
Cancer	22 (20.6)	29 (27.1)	0.261	34 (31.8)	0.062
Peripheral vascular disease	6 (5.6)	3 (2.8)	0.498	3 (2.8)	0.498
Peptic ulcer disease	24 (22.4)	22 (20.6)	0.739	21 (19.6)	0.615
Previous antibiotics exposure					
Macrolide	11 (10.3)	12 (11.2)	0.825	4 (3.7)	0.061
Aminoglycoside	9 (8.4)	7 (6.5)	0.603	2 (1.9)	0.030
penicillin/β-lactamase inhibitor	44 (41.1)	46 (43.0)	0.782	37 (34.6)	0.324
3rd generation cephalosporin	23 (21.5)	31 (29.0)	0.208	17 (15.9)	0.293
4th generation cephalosporin	37 (34.6)	37 (34.6)	1.000	11 (10.3)	<0.001
Carbapenem	34 (31.8)	41 (38.3)	0.316	15 (14.0)	0.002
Glycopeptide	28 (26.2)	31 (29.0)	0.646	6 (5.6)	<0.001
Fluoroquinolone	46 (43.0)	8 (7.5)	<0.001	23 (21.5)	0.001
TMP/SMX	13 (12.1)	2 (1.9)	0.003	3 (2.8)	0.009
Invasive medical devices					
Central venous catheter insertion	61 (57.0)	62 (57.9)	0.890	35 (32.7)	<0.001
Nasogastric tube insertion	99 (92.5)	98 (91.6)	0.800	56 (52.3)	<0.001
Foley catheter insertion	86 (80.4)	80 (74.8)	0.325	54 (50.5)	<0.001
Percutaneous surgical wound drainage use	20 (18.7)	26 (24.3)	0.318	24 (22.4)	0.499
Mechanical ventilation	87 (81.3)	77 (72.0)	0.106	20 (18.7)	<0.001

**Table 1** Risk factors associated with levofloxacin- and trimethoprim/sulfamethoxazole (TMP/SMX)-resistant *Steno-trophomonas maltophilia* isolates determined by bivariate analysis.

<sup>a</sup> Comparison of patients in the case group and control group A.

<sup>b</sup> Comparison of patients in the case group and control group B.

<sup>c</sup> Data are presented as means (standard deviations).

<sup>d</sup> Days of stay prior to isolation of S. *maltophilia*.

ICU, intensive care unit.

patients in the case group had higher rates for recent ICU admission (83.2% vs. 29.9%, p < 0.001), recent surgery (55.1% vs. 37.4%, p = 0.009), cerebrovascular disease (40.2% vs. 22.4%, p = 0.005), and exposure to multiple antibiotics including aminoglycoside (8.4% vs. 1.9%, P = 0.030), fourth-generation cephalosporin (34.6% vs. 10.3%, p < 0.001), carbapenem (31.8% vs. 14.0%, p = 0.002), glycopeptide (26.2% vs. 5.6%, p < 0.001), fluoroquinolone (43.0% vs. 21.5%, p < 0.001), and TMP/SMX (12.1% vs. 2.8%, p = 0.009). Moreover, patients in the case group were more likely to have invasive medical devices, including a central venous catheter, nasogastric tube, Foley

catheter, and ventilator, than those in control group B (57.0% vs. 32.7%, 92.5% vs. 52.3%, 80.4% vs. 50.5% and 81.3% vs. 18.7%, all p < 0.001).

The results of multivariate logistic regression are summarized in Table 2. In this analysis comparing the case group to control group A, previous usage of fluoroquinolone (odds ratio [OR] 22.824; 95% confidence interval [CI] 4.984–104.524) and previous usage of TMP/SMX (OR 17.724; 95% CI 1.495–210.097) were independent risk factors for LTSRSM isolation. When analysed using control group B, mechanical ventilation (OR 59.471; 95% CI 2.698–1311.079), cerebrovascular disease (OR 12.371; 95%

Table 2	Multivariate analysis (logistic	regression) of risk factors for	levofloxacin- a	and trimethoprim/sulfamethoxazole	(TMP/
SMX)-resis	stant Stenotrophomonas malte	ophilia isolates.			

Risk factor, n (%)	OR (95% CI)	P value
Case group <sup>a</sup> vs. control group A <sup>b</sup>		
Fluoroquinolone	22.824 (4.984–104.524)	<0.001
Trimethoprim/sulfamethoxazole	17.724 (1.495–210.097)	0.02
Case group vs. control group B <sup>c</sup>		
Mechanical ventilation	59.471 (2.698–1311.079)	0.010
Cerebrovascular disease	12.371 (1.943-78.748)	0.008
Fluoroquinolone	13.075 (1.502–113.819)	0.020

CI, confidence interval; OR, odds ratio.

<sup>a</sup> Patients with levofloxacin- and TMP/SMX-resistant S. *maltophilia* (case group).

<sup>b</sup> Patients with levofloxacin- and TMP/SMX-susceptible S. maltophilia (control group A).

<sup>c</sup> Patients without S. *maltophilia* infection (control group B).

CI 1.943–78.748), and previous usage of fluoroquinolone (OR 13.075; 95% CI 1.502–113.819) were independently associated with LTSRSM isolation. Only one factor, previous antibiotic exposure to fluoroquinolone, was found to be independently associated with occurrence of LTSRSM when the case group was compared to both control groups A and B, respectively.

# Antimicrobial susceptibility comparisons between LTSRSM and LTSSSM

The MIC ranges,  $MIC_{50}$ ,  $MIC_{90}$  values, and the resistant percentages to the tested antibiotics of the 85 LTSRSM and 85 LTSSSM isolates are shown in Table 3. All LTSRSM isolates consistently showed resistance to ciprofloxacin, another type of fluoroquinolone, with  $MIC_{50/90}$  values > 4  $\mu$ g/mL. Among the 170 S. maltophilia isolates tested, including the 85 LTSRSM and 85 LTSSSM isolates, 31% were resistant to ceftazidime, and the LTSRSM isolates were significantly more likely to exhibit resistance to ceftazidime than the LTSSSM isolates (43.5% vs. 25.9%, p = 0.016). With regard to tigecycline susceptibility, the resistance rates were significantly higher among the LTSRSM isolates than among the LTSSSM isolates (90.6% vs. 11.8%, p < 0.001). Moreover, the values of MIC<sub>50</sub> and MIC<sub>90</sub> were 4  $\mu$ g/mL and >8  $\mu$ g/mL for the LTSRSM isolates, respectively, which were at least 8fold higher than those for the LTSSSM isolates (<0.5  $\mu$ g/

mL and 1  $\mu$ g/mL, respectively). The other antibiotics tested (cefepime, gentamicin, amikacin, and colistin) revealed limited *in vitro* activity and resistant rates were not significantly different between the LTSRSM and LTSSSM isolates according to the CLSI MIC breakpoints for *P. aeru-ginosa* (p > 0.05).

#### Discussion

To our knowledge, the current study was the first to investigate risk factors and antibiotic susceptibilities for LTSRSM from hospitalized patients. A matched casecontrol-control design was used and compared the case group of patients with LTSRSM to two control groups of patients, one with LTSSSM and one comprised of randomly selected inpatients without *S. maltophilia* infection to eliminate the potential biased risk estimates from traditional case—control studies.<sup>21</sup> The main findings of our study were that previous antibiotic use of fluoroquinolone and TMP/SMX, patients with recent mechanical ventilation and cerebrovascular diseases were independent risk factors associated with subsequent LTSRSM occurrence. In addition, the MICs and resistance rates of tigecycline and ceftazidime were higher in LTSRSM than LTSSSM isolates.

Inpatients with multiple comorbidities frequently subjected to indwelling medical devices such as ventilators are

**Table 3** Minimal inhibitory concentration and susceptibility comparisons of antibiotics between levofloxacin- and trimethoprim/sulfamethoxazole (TMP/SMX)-resistant *Stenotrophomonas maltophilia* (LTSRSM) and levofloxacin- and TMP/SMXsusceptible S. *maltophilia* (LTSSSM).

Antibiotic MIC range (µg/mL)		MIC50 (µg/mL)		MIC90	MIC90 (µg/mL)		No. (%) resistant isolates		
	LTSRSM (n = 85)	LTSSSM (n = 85)	LTSRSM (n = 85)	LTSSSM (n = 85)	$\frac{1}{(n = 85)}$	LTSSSM (n = 85)	LTSRSM (n = 85)	LTSSSM (n = 85)	P value
Ceftazidime	<1->64	<1–>64	16	4	>64	>64	37 (43.5)	22 (25.9)	0.016
Cefepime	<1->64	<1–>64	16	32	>64	>64	63 (74.1)	64 (75.2)	1.000
Tigecycline	<0.5->8	<0.5–4	4	<0.5	>8	1	77 (90.6)	10 (11.8)	<0.001
Colistin	<0.5->16	<0.5->16	<0.5	<0.5	>16	>16	34 (40.0)	24 (28.2)	0.106
Ciprofloxacin	i >4	<0.12-4	>4	0.5	>4	2	85 (100)	12 (14.1)	<0.001
Gentamicin	2->16	<1–>16	>16	8	>16	>16	43 (50.6)	41 (48.2)	0.878
Amikacin	4->64	<2–>64	>64	>64	>64	>64	43 (50.6)	44 (51.8)	1.000

at risk of acquiring multidrug-resistant strains.<sup>22–24</sup> It may be that such patients with medical advices who need more nursing care during hospitalization are more likely to be infected from cross transmission and that infected resistant organisms are more difficult to be eradicated with antibiotic treatment than susceptible ones. Consistently, our study revealed that ventilator use was associated with subsequent LTSRSM occurrence. For the same reason, patients with poor functional status showing increased activities of daily living (ADL) score may need increased bedside care and are more likely to be infected with resistant bacteria.<sup>25–27</sup> This may explain why being a patient with cerebrovascular disease who may have a high ADL score was identified as another risk factor for LTSRSM occurrence in our study. The risk factors identified for LTSRSM occurrence mentioned above indicate that implementation of strict hand hygiene protocols in caring for inpatients may be helpful for preventing LTSRSM from spreading in the hospital. In our study, we also found that previous fluoroquinolone and TMP/SMX use were associated with subsequent LTSRSM occurrence, which was consistent with a previous study found that both antibiotics increased the likelihood of subsequent multidrug-resistant S. maltophilia isolation in cancer patients.<sup>28</sup> Besides past and our present clinical studies, we were able to observe similar results from basic studies of S. maltophilia in the laboratory. In experimental evolution studies reported, S. maltophilia exhibited simultaneous resistance to quinolone and TMP/ SMX after either TMP/SMX or quinolone exposure. The resistance mechanism was resulted from overexpression of resistance-nodulation-cell division (RND) efflux pumps, mainly SmeDEF.<sup>20,29</sup> Since overexpression of the RND efflux pump was also reported the major resistance mechanism for tigecycline in S. maltophilia, cross resistance to tigecycline may be anticipated.<sup>30</sup> Agree with our inference, we also noted elevated MICs and higher resistance rates of tigecycline in LTSRSM than LTSSSM in our study. Tigecycline has broad activity against gram-positive and -negative organisms, including multidrug-resistant organisms. From recent global surveillance studies, tigecycline also demonstrated good in vitro activity against S. maltophilia.<sup>31,32</sup> Although the clinical breakpoints of tigecycline for S. maltophilia have not been determined, clinical studies have suggested that tigecycline has efficacy equivalent to that of TMP/SMX in treating S. maltophilia infections.<sup>33</sup> Consequently, tigecycline has been considered an alternative therapeutic option. The cross-resistance of LTSRSM to tigecycline we presented here may further restrict already limited therapeutic choices in treating S. maltophilia infections. The MIC<sub>50</sub> values and resistance rates of ceftazidime were also higher in LTSRSM than in LTSSSM. The resistance mechanism of S. maltophilia for ceftazidime may generally be considered to correlate with intrinsic beta-lactamases L1 and L2 but overexpression of efflux pump transporters was also reported.<sup>34</sup> Whether there was difference between LTSSSM and LTSRSM on resistance mechanisms to ceftazidime warranted future investigations for further exploration. Study results on mechanisms of concurrent TMP/SMX and fluoroquinolone resistance reported before were from basic studies based on laboratory S. maltophilia strains under the laboratory environment. which may not be applicable to real clinical settings.

Results of current study from clinical data analysis revealing consistent results strengthened findings from past basic studies but molecular mechanism characterisation using clinical *S. maltophilia* isolates to correlate clinical data results was warranted for further validation. Cautious use of TMP/SMX and fluoroquinolone to reduced selective pressure may be important to limit LTSRSM emergence in the hospital. Other antibiotics tested in our study, including colistin, all revealed limited *in vitro* activity against LTSRSM isolates. The emergence of multidrug-resistant *S. maltophilia*, as presented here, would be a great concern for current and future medical practice. New drug development would be important to provide clinicians more therapeutic options when faced with infections caused by such resistant strains.

There were some limitations to this study. First, this study was retrospective in nature; hence, our results were susceptible to potential selection biases in the study. Second, molecular epidemiology of the collected isolates was not carried out. We therefore could not assess if patient-to-patient transmission occurred during the study period, which may have influenced our results in the clinical data analysis. Third, this was a single medical center in northern Taiwan and caution was necessary when extrapolating these results to other regions in Taiwan or other countries.

### Conclusion

The current study identified that mechanical ventilation use, patients with cerebrovascular disease, and previous fluoroquinolone and TMP/SMX exposure were independent risk factors for subsequent LTSRSM occurrence. LTSRSM isolates exhibited higher rates of resistance against tigecycline and ceftazidime than LTSSSM. Future multicenter studies with prospective evaluations and molecular characterizations of clinical LTSRSM isolates to corroborate our study results were necessary.

## Declaration of competing interest

The authors declare no conflict of interest.

### Acknowledgements

This work was supported by a grant from the Tri-Service General Hospital Penghu Branch (TSGH-PH-107-02 and TSGH-PH-109-5) and the Tri-Service General Hospital, National Defense Medical Center (TSGH-C103-187, TSGH-C104-195).

#### References

- Senol E. Stenotrophomonas maltophilia: the significance and role as a nosocomial pathogen. J Hosp Infect 2004;57: 1–7.
- 2. Chang YT, Lin CY, Chen YH, Hsueh PR. Update on infections caused by *Stenotrophomonas maltophilia* with particular attention to resistance mechanisms and therapeutic options. *Front Microbiol* 2015;6:893.

- Looney WJ, Narita M, Muhlemann K. Stenotrophomonas maltophilia: an emerging opportunist human pathogen. Lancet Infect Dis 2009;9:312–23.
- **4.** Brooke JS. New strategies against *Stenotrophomonas maltophilia*: a serious worldwide intrinsically drug-resistant opportunistic pathogen. *Expert Rev Anti Infect Ther* 2014;**12**:1–4.
- Andelkovic MV, Jankovic SM, Kostic MJ, Zivkovic Zaric RS, Opancina VD, Zivic MZ, et al. Antimicrobial treatment of *Stenotrophomonas maltophilia* invasive infections: systematic review. J Chemother 2019;31:297–306.
- 6. Muder RR. Optimizing therapy for *Stenotrophomonas maltophilia*. Semin Respir Crit Care Med 2007;**28**:672–7.
- 7. Cho SY, Kang CI, Kim J, Ha YE, Chung DR, Lee NY, et al. Can levofloxacin be a useful alternative to trimethoprimsulfamethoxazole for treating *Stenotrophomonas maltophilia* bacteremia? *Antimicrob Agents Chemother* 2014;**58**:581–3.
- 8. Wang YL, Scipione MR, Dubrovskaya Y, Papadopoulos J. Monotherapy with fluoroquinolone or trimethoprim-sulfamethoxazole for treatment of *Stenotrophomonas maltophilia* infections. *Antimicrob Agents Chemother* 2014;58:176–82.
- 9. Gales AC, Seifert H, Gur D, Castanheira M, Jones RN, Sader HS. Antimicrobial susceptibility of Acinetobacter calcoaceticus-Acinetobacter baumannii complex and Stenotrophomonas maltophilia clinical isolates: results from the SENTRY antimicrobial surveillance program (1997-2016). Open Forum Infect Dis 2019;6:S34-46.
- Herrera-Heredia SA, Pezina-Cantu C, Garza-Gonzalez E, Bocanegra-Ibarias P, Mendoza-Olazaran S, Morfin-Otero R, et al. Risk factors and molecular mechanisms associated with trimethoprimsulfamethoxazole resistance in *Stenotrophomonas maltophilia* in Mexico. J Med Microbiol 2017;66:1102–9.
- 11. Hu LF, Chen GS, Kong QX, Gao LP, Chen X, Ye Y, et al. Increase in the prevalence of resistance determinants to trimethoprim/sulfamethoxazole in clinical *Stenotrophomonas maltophilia* isolates in China. *PloS One* 2016;11:e0157693.
- Neela V, Rankouhi SZ, van Belkum A, Goering RV, Awang R. Stenotrophomonas maltophilia in Malaysia: molecular epidemiology and trimethoprim-sulfamethoxazole resistance. Int J Infect Dis 2012;16:e603–7.
- Lai CC, Chen YS, Lee NY, Tang HJ, Lee SS, Lin CF, et al. Susceptibility rates of clinically important bacteria collected from intensive care units against colistin, carbapenems, and other comparative agents: results from Surveillance of Multicenter Antimicrobial Resistance in Taiwan (SMART). *Infect Drug Resist* 2019;12:627–40.
- 14. Zhanel GG, Adam HJ, Baxter MR, Fuller J, Nichol KA, Denisuik AJ, et al. Antimicrobial susceptibility of 22746 pathogens from Canadian hospitals: results of the CANWARD 2007-11 study. J Antimicrob Chemother 2013;68:7–22.
- 15. Chung HS, Hong SG, Kim YR, Shin KS, Whang DH, Ahn JY, et al. Antimicrobial susceptibility of *Stenotrophomonas maltophilia* isolates from Korea, and the activity of antimicrobial combinations against the isolates. *J Kor Med Sci* 2013;28:62–6.
- Wu H, Wang JT, Shiau YR, Wang HY, Lauderdale TL, Chang SC, et al. A multicenter surveillance of antimicrobial resistance on Stenotrophomonas maltophilia in Taiwan. J Microbiol Immunol Infect 2012;45:120–6.
- Wang CH, Yu CM, Hsu ST, Wu RX. Levofloxacin-resistant Stenotrophomonas maltophilia: risk factors and antibiotic susceptibility patterns in hospitalized patients. J Hosp Infect 2020; 104:46–52.
- Wang CH, Lin JC, Chang FY, Yu CM, Lin WS, Yeh KM. Risk factors for hospital acquisition of trimethoprim-sulfamethoxazole resistant *Stenotrophomonas maltophilia* in adults: a matched case-control study. *J Microbiol Immunol Infect* 2017;50: 646–52.

- Pien CJ, Kuo HY, Chang SW, Chen PR, Yeh HW, Liu CC, et al. Risk factors for levofloxacin resistance in *Stenotrophomonas* maltophilia from respiratory tract in a regional hospital. J Microbiol Immunol Infect 2015;48:291-5.
- 20. Sanchez MB, Martinez JL. Overexpression of the efflux pumps SmeVWX and SmeDEF is a major cause of resistance to Cotrimoxazole in *Stenotrophomonas maltophilia*. *Antimicrob Agents Chemother* 2018;62:e00301–18.
- 21. Rafailidis PI, Bliziotis IA, Falagas ME. Case-control studies reporting on risk factors for emergence of antimicrobial resistance: bias associated with the selection of the control group. *Microb Drug Resist* 2010;16:303–8.
- 22. van Loon K, Voor In 't. A systematic review and meta-analyses of the clinical epidemiology of carbapenem-resistant enterobacteriaceaeHolt AF, Vos MC, editors. *Antimicrob Agents Chemother* 2018;62:e01730. 17.
- 23. Mittal G, Gaind R, Kumar D, Kaushik G, Gupta KB, Verma PK, et al. Risk factors for fecal carriage of carbapenemase producing Enterobacteriaceae among intensive care unit patients from a tertiary care center in India. *BMC Microbiol* 2016;16: 138.
- 24. Drinka P, Niederman MS, El-Solh AA, Crnich CJ. Assessment of risk factors for multi-drug resistant organisms to guide empiric antibiotic selection in long term care: a dilemma. *J Am Med Dir Assoc* 2011;12:321–5.
- **25.** Brito V, Niederman MS. Healthcare-associated pneumonia is a heterogeneous disease, and all patients do not need the same broad-spectrum antibiotic therapy as complex nosocomial pneumonia. *Curr Opin Infect Dis* 2009;**22**:316–25.
- 26. El Solh AA, Pietrantoni C, Bhat A, Bhora M, Berbary E. Indicators of potentially drug-resistant bacteria in severe nursing home-acquired pneumonia. *Clin Infect Dis* 2004;39:474–80.
- 27. Trick WE, Weinstein RA, DeMarais PL, Kuehnert MJ, Tomaska W, Nathan C, et al. Colonization of skilled-care facility residents with antimicrobial-resistant pathogens. J Am Geriatr Soc 2001;49:270–6.
- Ansari SR, Hanna H, Hachem R, Jiang Y, Rolston K, Raad I. Risk factors for infections with multidrug-resistant *Stenotrophomonas maltophilia* in patients with cancer. *Cancer* 2007; 109:2615–22.
- 29. Pak TR, Altman DR, Attie O, Sebra R, Hamula CL, Lewis M, et al. Whole-genome sequencing identifies emergence of a quinolone resistance mutation in a case of *Stenotrophomonas* maltophilia bacteremia. Antimicrob Agents Chemother 2015; 59:7117–20.
- Blanco P, Corona F, Martinez JL. Mechanisms and phenotypic consequences of acquisition of tigecycline resistance by *Stenotrophomonas maltophilia*. J Antimicrob Chemother 2019;74: 3221–30.
- Farrell DJ, Sader HS, Jones RN. Antimicrobial susceptibilities of a worldwide collection of *Stenotrophomonas maltophilia* isolates tested against tigecycline and agents commonly used for *S. maltophilia* infections. *Antimicrob Agents Chemother* 2010; 54:2735–7.
- 32. Pfaller MA, Flamm RK, Duncan LR, Mendes RE, Jones RN, Sader HS. Antimicrobial activity of tigecycline and cefoperazone/sulbactam tested against 18,386 Gram-negative organisms from Europe and the Asia-Pacific region (2013-2014). *Diagn Microbiol Infect Dis* 2017;88:177–83.
- **33.** Tekce YT, Erbay A, Cabadak H, Sen S. Tigecycline as a therapeutic option in *Stenotrophomonas maltophilia* infections. *J Chemother* 2012;**24**:150–4.
- **34.** Blanco P, Corona F, Martinez JL. Involvement of the RND efflux pump transporter SmeH in the acquisition of resistance to ceftazidime in *Stenotrophomonas maltophilia*. *Sci Rep* 2019;**9**: 4917.