



# The emerging roles of eosinophils: Implications for the targeted treatment of eosinophilic-associated inflammatory conditions

Carlo Lombardi<sup>a,\*</sup>, Alvise Berti<sup>b,c</sup>, Marcello Cottini<sup>d</sup>

<sup>a</sup> Departmental Unit of Allergology, Immunology & Pulmonary Diseases, Fondazione Poliambulanza, Brescia, Italy

<sup>b</sup> Ospedale Santa Chiara and Department of Cellular, Computational and Integrative Biology (CIBIO), University of Trento, Trento, Italy

<sup>c</sup> Thoracic Disease Research Unit, Mayo Clinic, Rochester, MN, USA

<sup>d</sup> Allergy and Pneumology Outpatient Clinic, Bergamo, Italy

## ARTICLE INFO

### Keywords:

Eosinophils  
Homeostasis  
Inflammation  
Asthma  
Type 2 phenotype  
Eosinophilic granulomatosis with polyangiitis  
Hypereosinophilic syndrome  
Mepolizumab  
Benralizumab  
Reslizumab  
Dupilumab

## ABSTRACT

Eosinophils have multiple relevant biological functions, including the maintenance of homeostasis, host defense against infectious agents, innate immunity activities, immune regulation through Th1/Th2 balance, anti-inflammatory, and anti-tumorigenic effects. Eosinophils also have a main role in tissue damage through eosinophil-derived cytotoxic mediators that are involved in eosinophilic inflammation, as documented in Th2-high asthma and other eosinophilic-associated inflammatory conditions.

Recent evidence shows that these multiple and apparently conflicting functions may be attributed to the existence of different eosinophil subtypes (i.e.: tissue resident and inducible eosinophils). Therapeutic intervention with biological agents that totally deplete tissues and circulating eosinophils or, *vice versa*, maintain a minimal proportion of eosinophils, particularly the tissue-resident ones, could therefore have a very different impact on patients, especially when considering the administration of these therapies for prolonged time. In addition, the characterization of the predominant pathway underlying eosinophilic inflammation by surrogate biomarkers (circulating eosinophils, organ-specific eosinophils levels such as eosinophil count in sputum, bronchoalveolar lavage, tissue biopsy; total circulating IgE levels, or the use of FeNO) in the single patient with an eosinophilic-associated inflammatory condition could help in choosing the treatment.

These observations are crucial in light of the increasing therapeutic armamentarium effective in modulating eosinophilic inflammation through the inhibition in different, yet complementary ways of eosinophil pathways, such as the interleukin-5 one (with mepolizumab, benralizumab, reslizumab) or the interleukin-4/13 one (with dupilumab and lebrikizumab), in severe T2-high asthma as well as in other systemic eosinophilic associated diseases, such as eosinophilic granulomatosis with polyangiitis and hypereosinophilic syndrome.

## 1. Background

In 1879 Paul Ehrlich identified blood eosinophils in several mammals (rabbits, dogs, and humans) by eosin staining (Kay, 2015; Lombardi, 1996). After this first observation, several studies have reported on the multiple biological functions of eosinophils. Indeed, the intrinsic roles of eosinophils are much more complex, including the maintenance of homeostasis, host defense against infectious agents, innate immunity activities, immune regulation through Th1/Th2 balance, anti-inflammatory, and anti-tumorigenic effects (Kanda et al., 2021). Moreover, eosinophils have a main role in tissue damage through eosinophil-derived cytotoxic mediators that are involved in eosinophilic

inflammation, as documented in Th2-high asthma and other eosinophilic-associated diseases (Choi et al., 2020; Januskevicius et al., 2020). Recently, it has been hypothesized that these multiple and apparently conflicting effects may be attributed to the existence of different eosinophil subtypes (i.e.: tissue resident and inducible eosinophils) (Kanda et al., 2021; Mesnil et al., 2016). These observations are crucial since we have now treatments very effective in modulating eosinophilic inflammation in different fashions through the inhibition of interleukin(IL)-5 activity (i.e. mepolizumab, benralizumab, reslizumab) or targeting IL4/13 activity (dupilumab, lebrikizumab) (Riggioni et al., 2020; Bagnasco et al., 2017). These eosinophil-targeted therapeutic strategies may impact on Th2-inflammation with different, yet

\* Corresponding author. Departmental Unit of Pneumology & Allergology, Fondazione Poliambulanza Istituto Ospedaliero, Via Bissolati, 57 Brescia, 25100, Italy.  
E-mail addresses: [carlo.lombardi@poliambulanza.it](mailto:carlo.lombardi@poliambulanza.it), [lombardicarlo@libero.it](mailto:lombardicarlo@libero.it) (C. Lombardi).

<https://doi.org/10.1016/j.crimmu.2022.03.002>

Received 30 November 2021; Received in revised form 6 February 2022; Accepted 7 March 2022

Available online 21 March 2022

2590-2555/© 2022 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

complementary mechanisms, potentially with different application within the therapeutic armamentarium of eosinophilic-associated disorders, as Th2-asthma, as well as in other systemic eosinophilic associated diseases, such as eosinophilic granulomatosis with polyangiitis and hypereosinophilic syndrome.

In this review, we discuss the recent literature evidence on the role of eosinophil in health and diseases. We also discussed about the potential impact of recent biological anti-IL-5 and IL4/13 therapies on these different eosinophil subtypes.

## 2. Biology of eosinophils

Eosinophils are terminally differentiated leukocytes approximately accounting for 1–5% of peripheral blood leukocytes in healthy individuals (Kim and Jung, 2020; Jung and Rothenberg, 2014; Jung, 2015). Eosinophils are produced in the bone marrow and thus they migrate into the peripheral circulation as mature cells, identified as Siglec-F<sup>+</sup> CCR3<sup>+</sup> F4/80<sup>+</sup> CD62L<sup>+</sup>. Here they have a relatively short half-life, approximately 18 h (Fig. 1). They migrate to the peripheral tissues under homeostatic conditions, or to inflammatory sites in response to recruitment signals, primarily IL-5 and eotaxin-1 (CCL11) (Rothenberg and Hogan, 2006; Rosenberg et al., 2013). GATA-1 is the most important transcription factor for eosinophil maturation, as observed in mice with targeted deletion of the high-affinity GATA-1 binding site in the Gata1 promoter (Yu et al., 2002). Further eosinophils evolutionary states (differentiation and proliferation) are also regulated by IL-5, IL-3 and GM-CSF (Hara and Miyajima, 1996).

IL-5 is the most specific cytokine released for inducing the selective differentiation and mobilization of eosinophils from the bone marrow during inflammation, as well as for the eosinophil homing into various tissues in the steady state (Mould et al., 1997). During allergic inflammation, in addition to Th2 cells group 2, innate lymphoid cells (ILC2s) are a major non-T cell source of IL-5 (Klose and Artis, 2016). Proliferation and tissue accumulation of eosinophils is modulated by a complex interaction between pro-survival signaling factors (GM-CSF, IL-3, IL-5) and inhibitory receptor signaling (Kim and Jung, 2020). Notably, the tissue distribution of eosinophils may be IL-5 independent, as demonstrated by the observation of residual tissue eosinophils in IL-5-deficient

mice (Mould et al., 1997). Furthermore, eotaxin promotes selective recruitment of eosinophils into eotaxin-expressing tissues cooperatively with IL-5 via an IL-5-independent manner (Weller and Spencer, 2017). Beside IL-5 and eotaxin, eosinophil trafficking into inflamed tissues also involves IL-4 and IL-13 (with IL-5, Th2 cytokine pattern), and a number of cytokines, chemokines, and adhesion molecules (i.e.,  $\alpha4\beta7$  and  $\alpha4\beta1$  integrin) that contribute in different ways to eosinophil homing (Fig. 1).

### 2.1. Eosinophils as contributors to tissue homeostasis and regulators of host defence

Eosinophils have a wide range of pleiotropic activities including protective immunity and anti-microbial activity, but are also responsible of several physiological responses, such as organ development and metabolism. Although eosinophils are normally considered circulating blood cells, a fraction of eosinophils reside stably in various tissues (Fig. 2). In particular, eosinophils have been shown to be present in the thymus, adipose tissue, gastrointestinal (GI) tract, lungs and female reproductive system during homeostatic conditions (Kim and Jung, 2020). The GI tract harbors the largest number of tissue-resident eosinophils in the body (about 20%–30% of the total number of resident leukocytes), whereas eosinophils in the adipose tissue and the lung constitute only  $\leq 4\%$  of the stromal/vascular fraction and  $\leq 1\%$  of total leukocytes, respectively (Wu et al., 2011; Mesnil et al., 2016). Tissue-resident eosinophils also can be found in the mammary gland and uterus tissues in homeostatic and various inflamed tissues. Tissue-specific microenvironmental signals have significant roles in the phenotypic and functional properties of eosinophils.

In some of these organs, a reference value to establish an excess of eosinophils have been established in humans. For instance, eosinophils  $\geq 2\%$  of cells in sputum as well as  $\geq 3\%$  in bronchoalveolar lavage are usually considered pathologic. For gastrointestinal tract, for eosinophilic esophagitis  $\geq 15$  eosinophils per high-power microscopy field (HPF) are required, while the cut off may change for colitis from  $>50$ /HPF in the right colon,  $>35$ /HPF in the transverse colon and  $>25$  per HPF in the left colon (Impellizzeri et al., 2019).

Tissue-resident eosinophils are primarily found in several mucosal sites where they contribute to various homeostatic and tissue-protective

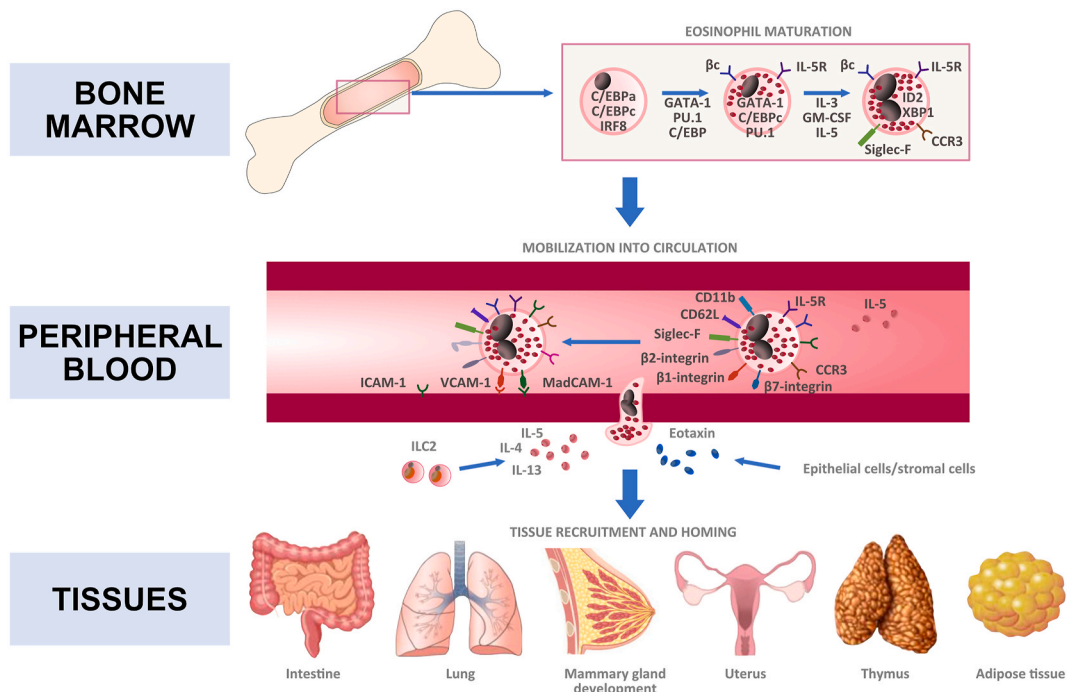


Fig. 1. Development of eosinophils expressing various types of functional cell surface molecules: from bone marrow to tissues.

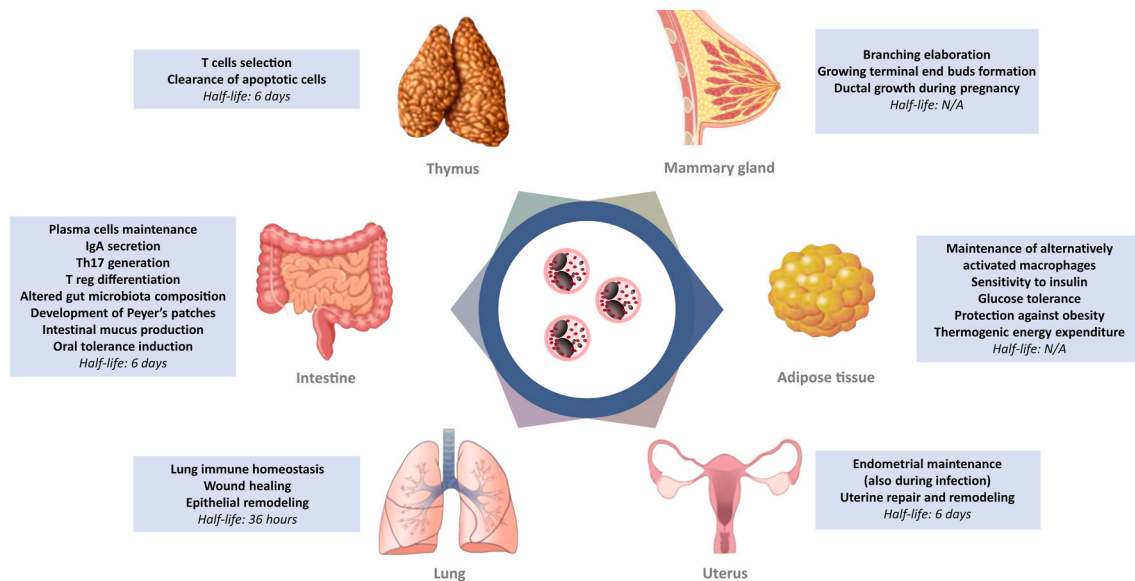


Fig. 2. Schematic overview of the homeostatic function of eosinophils in the individual organ tissues.

functions (Shah et al., 2020). Eosinophils play a beneficial role in regulating and modulating immune responses. Indeed, it has become clear that eosinophils also play important non-inflammatory activities in the generation and maintenance of adaptive immune responses. Eosinophils, being a major source of the plasma cell survival factor APRIL (activation and proliferation-induced ligand), are essential not only for the long-term survival of plasma cells in the bone marrow, but also for the maintenance of these cells in the lamina propria underlying the gut epithelium (Berek, 2015). Furthermore, data suggest that eosinophils are required for tissue integrity and are necessary for tissue remodeling (Lee et al., 2010). It has been demonstrated also the role of these cells in the generation of IgA expressing B cells, and thus in the development of the IgA plasma cells which reside in the lamina propria and contribute to immune defense at mucosal surfaces (Chu et al., 2014). Innate immune activities of eosinophils include expression of pattern recognition receptors, such as Toll-like receptors 1–5, 7, and 9, nucleotide oligomerization domains 1 and 2, Dectin-1, and receptor for advanced glycation end products, which recognize pathogen-associated molecular patterns or danger-associated molecular patterns (Jacobsen et al., 2012). In recent years, eosinophils have also been recognized as having numerous immuno-modulatory functions toward B cell lineage maturation or maintenance: a) in T cell-dependent immune responses, eosinophils enhance early B cell activation; b) eosinophils and megakaryocytes are required for efficient homing of plasma cells to bone marrow; c) eosinophils are the main source of plasma cell survival factors; and d) eosinophils are required for the maintenance of plasma cells in the lamina propria (Berek, 2015).

It is also well known that eosinophils have anti-microbial activity. In particular, eosinophil numbers are often elevated in the course of parasitic infections and represent important effector cells in helminth infections (Simon et al., 2020). Furthermore, recent studies demonstrating anti-bacterial activities for eosinophils in the context of eosinophil extracellular traps (EET) formation, and less recently it has already been shown that MBP and ECP have bactericidal properties of MBP and ECP *in vitro*. Data *in vivo* seem to support a direct anti-bacterial role of eosinophils, as the adoptive transfer of eosinophils has been found to be sufficient to protect against bacterial septic shock (Lee et al., 2010; Yousefi et al., 2008). Eosinophils may also exhibit anti-viral properties. Granule protein eosinophil-derived neurotoxin (EDN) has been shown to have anti-viral activity *in vitro* (Rugeles et al., 2003). In experimental models, hyper-eosinophilic mice clear respiratory syncytial virus more effectively than wild-type mice (Phipps et al., 2007) and can prevent

infection with the natural rodent pathogen pneumonia virus of mice (Percopo et al., 2014). In guinea pigs, allergen-induced eosinophilia has been associated with a decreased viral load during parainfluenza virus infection (Adamko et al., 1999). Both human and murine eosinophils produce NO via inducible NO synthase, which can have direct antiviral effects on parainfluenza virus and RSV (Drake et al., 2016). Although the role of eosinophils in COVID-19 has not been elucidated yet, it has been shown that SARS-CoV2 infection can be associated with profound eosinopenia and that persistent eosinopenia may be associated with clinical deterioration and increased risk of mortality (Roca et al., 2020).

## 2.2. Eosinophils in inflammatory processes

Eosinophils can regulate local immune and inflammatory responses, and their accumulation in the blood and tissue is associated with several allergic, rheumatologic, infectious, neoplastic, and rare idiopathic disorders. Although eosinophils can contribute to tissue homeostasis in steady-state conditions, many studies have trended toward focusing on the contribution of eosinophils in the pathogenesis of eosinophil-associated diseases. Indeed, eosinophils may exert their biological effects via cytotoxic mediators such as type 2 cytokines (IL-4, IL-5, IL-9, IL-13, and IL-25), type 1 cytokines (IL-12, IFN- $\gamma$ ), acute proinflammatory cytokines (TNF- $\alpha$ , IL-1b, IL-6, and IL-8), chemokines, and lipid mediators (PAF and LTC4) (Table 1) (Kanda et al., 2021). A significant association was established between eosinophils and a number of disease conditions characterized by an inflammatory state (Lombardi and Pas-salacqua, 2003). Activation of eosinophils and release of proinflammatory lipid mediators, cytokines, free oxygen radicals, highly charged cationic proteins contribute to the onset and maintenance of tissue inflammation. Furthermore, eosinophil accumulation in blood and tissues has been related to a defect in their apoptotic death (Shen and Malter, 2015).

Nowadays the scientific community is changing perspective, recognizing the increasingly paramount role of eosinophils in the pathogenesis of non-allergic inflammatory diseases, beside the well-known importance in the pathogenesis of the allergic ones. Indeed, eosinophilic inflammation can involve locomotor, urinary, cardiovascular, nervous, gastrointestinal systems and other mucosal surfaces, such inflammation also can accompany tissue trauma, foreign-body reactions, and necrotic or granulomatous processes (Gonlugur and Gonlugur, 2006).

Eosinophils may also play a role in tissue repair and regeneration; for example, muscle damage promotes rapid recruitment of eosinophils in

**Table 1**  
Mediators of eosinophils.

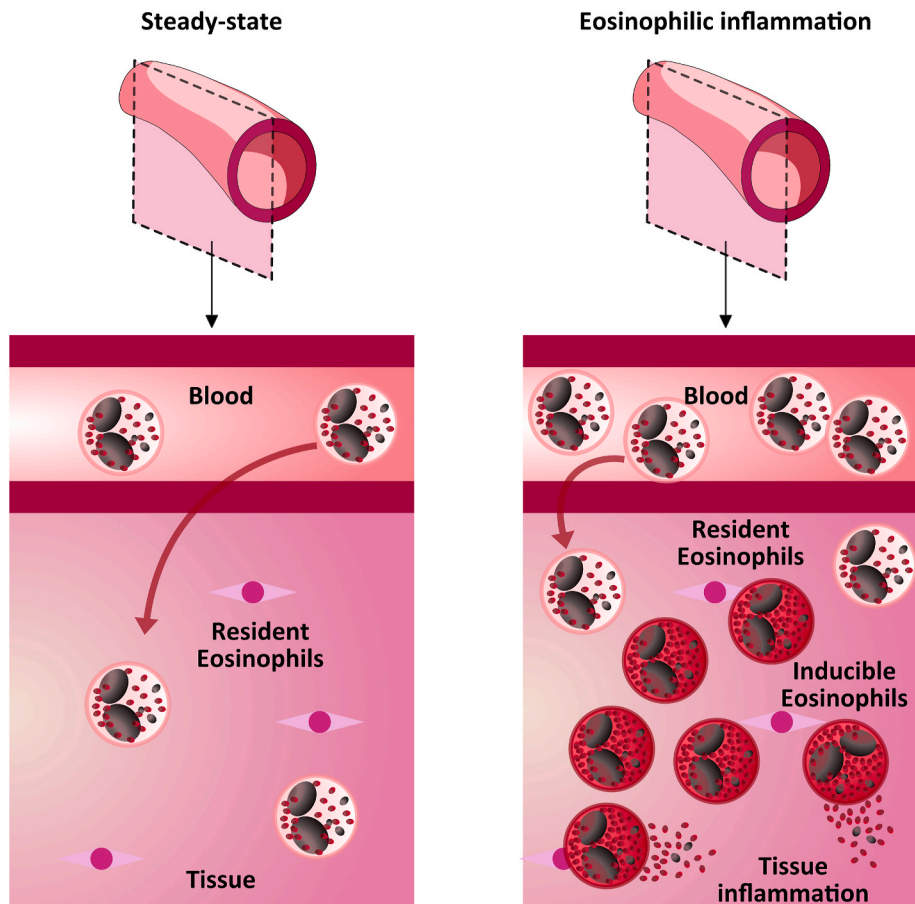
CLASS OF MEDIATORS	EXAMPLES
Granule-associated proteins	Major Basic Protein (MBP), Eosinophil Peroxidase (EPX), Eosinophil Cationic Protein (ECP), Eosinophil-Derived Neurotoxin (EDN), Charcot-Leyden Crystal (CLC) protein
Cytokines	IL-1 $\beta$ , IL-1R $\alpha$ , IL-2, IL-3, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, IL-13, IL-16, IL-17, IFN- $\gamma$ , GM-CSF, TGF- $\beta$ , TNF- $\alpha$
Chemokines	CCL-1/eotaxin-1, CCL-1/MCP-4, CCL2, CCL3, CCL5, CCL7, CCL8, CCL11, CCL13, CXCL1, CXCL10, CXCL12
Growth factors	Vascular Endothelial Growth Factor (VEGF), Platelet-Derived-Growth Factor (PDGF), Proliferation-Inducing Ligand (APRIL), Epidermal Growth Factor (EGF), Stem Cell Factor (SCF)
Neuromediators	Substance P, Vasoactive Intestinal Peptide (VIP), Nerve Growth Factor (NGF)
Lipid mediators	Leukotrienes (LTD4-LTE4), Prostaglandines (PGE1, PGE2)
Enzymes	Matrix metalloproteinase-9 (MMP-9), acid phosphatase, collagenase, histaminase, phospholipase D, catalase, arylsulphatase B
Adhesion molecules	$\beta$ 1-integrin, $\beta$ 2-integrin, CD62L, CD49f, CD49d, CD11a, CD11b, CD11c

the inflammatory foci and eosinophils play a key role in muscle regeneration during muscle injury as a major source of IL-4. IL-4 produced by eosinophils activates muscle resident fibrocyte-adipocyte progenitors (FAPs), which induce regeneration of injured muscles (Aoki et al., 2021). Eosinophils can induce angiogenesis by the production of pre-formed pro-angiogenic mediators, among others the vascular endothelial growth factor (VEGF). Eosinophil-derived IL-4 is also required for liver regeneration. Eosinophil-derived IL-4 induces the

proliferation of quiescent hepatocytes and regulates the regeneration of the liver (Li and Hua, 2017). Furthermore, eosinophils confer protection following myocardial infarction (Lavigne, 2020). Thus, eosinophils play a profound role in tissue repair and regeneration in various organs. The study of various types of eosinophilic inflammation may increase our understanding of the biological responses of eosinophil leukocytes to different inflammatory stimuli and recent research suggests that eosinophils may have additional roles in these settings that are related to control and resolution of inflammation (Strandmark et al., 2016).

### 2.3. Eosinophils subpopulations in health and diseases

Eosinophils are a heterogeneous cell population with different functional characteristics depending on the site of residence. The function exerted is likely controlled by the local *milieu*, that can induce the upregulation of pro- or anti-inflammatory eosinophils or the conversion of inflammatory eosinophils into anti-inflammatory ones or *vice versa* (Yang et al., 2017). It would be worthwhile to phenotype different eosinophil subpopulations by membrane surface markers, in order to distinguish homeostatic versus inflammatory eosinophils. A functional distinction has been drawn between eosinophils recruited from circulation in response to tissue pathology (e.g. allergies or parasitic infection) and those that maintain residence and are present in tissues at homeostasis (Fig. 3). Like T and B cells, ILC, and dendritic cells, a new classification system that defines eosinophil subpopulations on the basis of surface antigen expression, tissue localization, content, and function, has been presented for mouse eosinophils: eosinophilic precursors, steady state eosinophils, type 1 and type 2 eosinophils (Simon et al., 2020; Abdala-Valencia et al., 2018). Eosinophil Progenitors are



**Fig. 3.** Eosinophilic subpopulations. Left panel: steady state with resident eosinophils (rEOSs); Right panel: eosinophilic pathological condition with inducible eosinophils (iEOSs) tissue accumulation" (modified from Kanda et al. (Kanda et al., 2021)).

immature eosinophils or committed precursors undergoing *in situ* hematopoiesis; steady state eosinophils are true resident eosinophils in quiescent tissues, with nonsegmented “donut-shape” nuclear morphology and eosin staining. Type 1 eosinophils are found in interstitium or stroma in “transient” morphogenetic contexts and during Type 1 immune activation, featuring segmented nuclear morphology but lacking vacuolarization. Type 2 eosinophils are found within the epithelium and during Type 2 immune inflammation, characterized by highly segmented nuclei and the presence of vacuoles (Abdala-Valencia et al., 2018). Phenotypically, type 1 eosinophils are CD101<sup>low</sup>, CD62<sup>L</sup>, Siglec-F<sup>med</sup>, while type 2 eosinophils have higher CD101 and Siglec-F, while lacking expression of CD62. In addition, mouse eosinophils exhibited a proinflammatory gene expression signature that was lost upon initiation of the repair phase (Reichman et al., 2017).

In humans, the intestinal mucosa of subjects with eosinophil esophagitis included both CD25 positive and negative eosinophils (Straumann et al., 2005). In eosinophilic skin diseases, populations of eosinophils expressing unique subsets of cytokines were evaluated (Roth et al., 2011). Classically, they have been divided into 2 different types: normodense and hypodense eosinophils, which are indicated by normal density and lower density, respectively (Prin et al., 1983). In asthmatic patients, an increased number of hypodense eosinophils in the peripheral blood was correlated with clinical severity and airway hyper-responsiveness, and inhaled corticosteroids significantly decreased hypodense eosinophils (Kuo et al., 1994). Furthermore, increased number of hypodense eosinophils was observed in bronchoalveolar lavage eosinophils following an antigen-challenge (Kroegel et al., 1994). In patients with atopic dermatitis and peripheral eosinophilia, a correlation between hypodense eosinophil levels and disease severity has also been demonstrated (Miyasato et al., 1996).

More recently, it has also been shown that eosinophils can be phenotypically distinguished into subtypes in mice lungs. Indeed, Mesnil et al. found 2 phenotypically distinct subtypes of eosinophils in the lung tissue under steady-state and house dust mites-induced allergic inflammation: resident eosinophils (rEos; Siglec-Fint CD62L + CD101low), which are IL-5 independent parenchymal cells with a ring-shaped nucleus, and inducible eosinophils (iEos; Siglec-Fhigh CD62L- CD101high), which are IL-5 dependent peribronchial cells with a segmented nucleus (Kanda et al., 2021; Mesnil et al., 2016). Lung rEos were located in the parenchyma, which contrasts with the localization of eosinophils in asthma, in which eosinophils are classically within the peribronchial area (Barnes, 2008). In particular, it was observed that rEos expressed genes implicated in the negative regulation of immune responses (e.g., Anxa1, Nedd4, Runx3, Serpinb1a, and Ldlr) in contrast to iEos that highly express several proinflammatory genes (e.g., Slc3a2, Tlr4, C3ar1, Il13ra1, and Il6) (Mesnil et al., 2016). Kanda et al. have also shown the presence of phenotypically distinct rEos and iEos in the lungs from naïve and OVA-induced allergic asthma mice, and these rEos and iEos were characterized by normodense and hypodense eosinophils, respectively (Kanda et al., 2021). However, a recent study analyzing gene expression of human or murine eosinophils at steady-state conditions *in vivo* and after stimulation *in vitro*, showed that restriction of IL5 availability did not elicit any detectable transcriptional response in steady-state residual eosinophils, and influenced only a few genes in their response to *in vitro* stimulation. Ultimately, this seems to suggest that targeting the IL5 pathways spares a pool of circulating residual eosinophils largely resembling those of healthy individuals (Von Hulst, 2021).

In contrast with mice, rEos and iEos has not been fully characterized in human tissue. In humans, the modulation of surface expression levels in a large variety of receptors on eosinophils depends on environmental factors. The upregulation of several surface receptors reflecting activation markers (i.e., CD11b, CD11c, CD13, CD18, CD25, CD29, CD58, CD63, CD66e, CD67, CD108, CD123, HLA-DR, and TSLPR) was observed on eosinophils in the BAL and/or induced sputum of patients with allergic disease, which leads to increased inflammation (Johansson, 2014; Metcalfe et al., 2016). Furthermore, activated eosinophils from

the nasal polyps of patients with eosinophilic chronic rhinosinusitis with polyposis expressed CD69 whereas eosinophils from the peripheral blood of the same patients did not (Yun et al., 2020). There is experimental and clinical evidence that only activated eosinophils can express MHC class II (Hansel et al., 1991; Duez et al., 2004). Moreover, Januskevicius et al. found out that eosinophil subtypes differ in their adhesive properties and survivability (Januskevicius et al., 2020). Finally, in contrast with iEos, human rEos do not evoke inflammatory reactions or tissue damage (Straumann et al., 2005).

Altogether, rEos and iEos appear to be functionally different for multiple reasons: 1) lung rEos are parenchymal Siglec-Fint CD125int cells with a ring-shaped nucleus; 2) lung rEos are not affected by allergic inflammation and differ from iEOS; 3) lung rEos and iEos have distinct gene expression profiles; 4) expression of CD62L and CD101 discriminates between mouse rEos and iEos; 5) rEos are independent of IL-5 for their presence in the lung; 6) rEos, but not iEos, can inhibit the development of Th2 responses to house dust mites and the pro-Th2 potential of dendritic cells; and 6) lung parenchymal eosinophils exist in humans and are phenotypically distinct from iEos found in asthmatic airways.

### 3. Eosinophilic-associated inflammatory conditions

The causes of eosinophilia are multiple. It is essential to systematically approach patients who present with unexplained eosinophilia, because the only a correct and timely diagnosis lead to a potentially resolute treatment. Regarding the common causes of blood eosinophilia, there are significant geographical differences, with reported parasitic infections in tropical settings and allergic diseases or drug hypersensitivity reactions in more developed countries (Lombardi, 1996; Lombardi and Passalacqua, 2003). Many other hematologic (primary) and non-hematologic (reactive or secondary) conditions associated with eosinophilia have been described (Table 2). Only after an appropriate diagnostic work up to rule out of the more common causes, the diagnosis of an eosinophilic-associated inflammatory condition should be made.

Eosinophilic-associated inflammatory conditions are a heterogeneous and often overlapping group of different diseases accumulated by high tissue infiltrating and/or circulating eosinophils, potentially affecting different organs, such as upper and lower respiratory tract or digestive system, without a known cause (Table 3).

**Table 2**  
Differential diagnosis of peripheral eosinophilia.

CLASS	CONDITIONS
INFECTIONS	<ul style="list-style-type: none"> <li>- Parasitic (helminths, ectoparasites, Strongyloides, Toxocara Canis, etc)</li> <li>- Virus (HIV, HTLV)</li> <li>- Fungal (coccidiomycosis)</li> <li>- Bacterial (Tuberculosis)</li> </ul>
DRUG HYPERSENSITIVITY	<ul style="list-style-type: none"> <li>- Antibiotics (penicillins, cephalosporins, quinolones, sulfonamides)</li> <li>- NSAIDs</li> <li>- Antiepileptics (valproate, phenytoin)</li> <li>- Antidepressants (fluoxetine, amitriptyline)</li> <li>- Antihypertensives (ACE inhibitors, B-blockers)</li> </ul>
HEMATOLOGIC/NEOPLASTIC DISORDERS	<ul style="list-style-type: none"> <li>- Systemic mastocytosis</li> <li>- Solid tumors (adenocarcinoma, squamous cells carcinomas, etc)</li> <li>- Hematologic malignancy (chronic myeloid leukemia, Hodgkin lymphoma)</li> </ul>
IMMUNE DYSREGULATIONS	<ul style="list-style-type: none"> <li>- Hyper-IgE-syndrome</li> <li>- Kimura disease</li> <li>- Sarcoidosis</li> <li>- Inflammatory bowel diseases</li> <li>- IgG4-related disease</li> <li>- Autoimmune lymphoproliferative syndrome</li> </ul>
COMMON ALLERGIC DISORDERS	<ul style="list-style-type: none"> <li>- Allergic rhinitis, asthma, atopic dermatitis, etc.</li> <li>- Allergic Bronchopulmonary Aspergillosis</li> </ul>
OTHERS	

**Table 3**  
Eosinophilic-associated inflammatory conditions.

Systemic eosinophilic-associated conditions	Organ-limited eosinophilic-associated conditions
<b>Eosinophilic granulomatosis with polyangiitis (EGPA)</b>	<i>Eosinophilic Asthma</i>
<b>Hypereosinophilic syndrome (HES)</b>	<i>Chronic Rhinosinusitis (CRS)±Nasal Polyps (CRSwNP)</i>
<b>Other systemic eosinophilic-associated conditions</b>	<i>Eosinophilic Esophagitis/Gastroenteritis/colitis</i>
<b>Gleich syndrome</b>	<i>Eosinophilic Pneumonia</i>
<b>Kimura Disease</b>	<i>Eosinophilic Cystitis</i>
<b>Others</b>	<i>Eosinophilic Fasciitis</i>

### 3.1. Eosinophilic type 2 asthma

At least three major determinants have been identified in asthma: allergy (allergic vs non-allergic asthma), age of onset (early versus late onset) and prevalent type of airway inflammation (T2-high or T2-low, eosinophilic/neutrophilic/paucigranulocytic/mixed).

These features combined themselves to form clinically overlapping phenotypes. The phenotype of severe type 2 asthma is characterized by Th2 inflammation and activation of innate lymphoid cells-2 (ILC-2). The identification of T2-high patients is facilitated by the use of several non-invasive biomarkers, as elevated peripheral blood eosinophil count and elevated circulating IgE, which are used in clinical practice because of their diffuse availability, easy interpretation and reproducibility (Lim and Nair, 2018). The measurement of eosinophils in induced sputum is an extensively studied validated biomarker, but it is investigative, not available to all centers and it is more expensive. However, it is important to remember that blood eosinophil count is subject to wide variability over time and may not always accurately reflect the cellular pattern of the airways in asthmatics (Petsky et al., 2018). Measurement of fractional exhaled nitric oxide (FENO) is a noninvasive, safe, and simple method of quantifying airway inflammation, that reflects eosinophilic inflammation of the airways. Of note, inducible nitric oxide synthase (iNOS) is induced by a variable range of cytokines and mediators, but more specifically by IL-13 (Locksley, 2010; Amelink et al., 2013; de Groot et al., 2016).

The pathobiology of T2-high early-onset allergic asthma has been well established in animal and human studies. In patients who are genetically predisposed to an allergic immune response, the inhalation of an environmental stimuli such as the aeroallergen triggers the epithelial cells to release cytokines (IL-25, IL-33, thymic stromal lymphopoietin) and initiate a series of downstream events that differentiate naïve T cells into mature Th2 lymphocytes, that together with ILC-2, are capable of producing the classic Th2 cytokine pattern: IL-4, IL-5 and IL-13. The release of the Th2 cytokine IL-4 stimulates the cellular B isotype switch leading to IgE synthesis, a hallmark of allergic inflammation (Yu et al., 2014). In the event of re-exposure to allergens to which the patient is sensitized, IgE attached to mast cells and basophils leads to the release of inflammatory mediators (histamines, prostaglandins and leukotrienes). Through their effects on airway smooth muscle, these mediators are responsible for the clinical asthma syndrome that characterizes the initial response to allergen exposure. In addition, through their systemic effects on other organs and systems, inflammatory mediators can lead to the development of comorbidities, such as rhinitis and polyposis.

The late-onset variant of severe asthma is characterized by pathophysiological mechanisms different from childhood allergic asthma, typically presenting in the fourth or fifth decade of life. In this phenotype, T2-high inflammation is demonstrable by increased airway eosinophils, which in a remarkable fraction of patients tends to persist despite corticosteroid therapy. Frequent asthmatic exacerbations occur with significant dependence on oral corticosteroids. Chronic rhinosinusitis and nasal polyposis with or without sensitivity to acetylsalicylic acid (ASA) may also be present in these patients. Although late-onset

eosinophilic asthma is characterized by T2-high inflammation, as in early-onset, experimental evidence suggests that “allergen-independent” signals involving ILCs producing IL-5 and IL-13 are activated (Smith et al., 2016). These observations have therefore led to the distinction in adult eosinophilic asthma between an allergic form from a non-allergic form. Some evidence has demonstrated that persistent eosinophilia in upper and lower airway mucosa contributes to asthma severity by producing various mediators including cytokines, chemokines and granule proteins. Moreover, extracellular traps released from eosinophils have been revealed to enhance type 2 inflammation in patients with severe asthma (Choi et al., 2020). Alarmin-like cytokines (IL-33 and thymic stromal lymphopoietin (TSLP), released from airway epithelium, are involved in the development of severe asthma (Li et al., 2018; Mitchell and O’Byrne, 2017). IL-33 activates the ILC to exacerbate airway inflammation but stimulates eosinophils as well (Stolarski et al., 2010). TSLP is known to be important for inducing type 2 cytokine production, leading to the activation of eosinophils (Wong et al., 2010). Persistent airway inflammation induced by eosinophils leads to constant tissue damage, resulting in smooth muscle thickening, goblet cell hyperplasia and extracellular matrix protein deposition called airway remodeling (Tagaya and Tamaoki, 2007).

### 3.2. Eosinophilic granulomatosis with polyangiitis

Eosinophilic granulomatosis with polyangiitis (EGPA) is a rare primary systemic necrotizing vasculitis affecting the small vessels, often occurring in patients with late-onset asthma and sustained peripheral blood eosinophilia (Berti et al., 2020a). EGPA presents with upper airway tract and lung involvement in virtually all cases, including rhinosinusitis, asthma and migratory pulmonary infiltrates; peripheral neuropathy, cardiac involvement and skin lesions (Churg and Strauss, 1951; Groh et al., 2015). Anti-neutrophil cytoplasmic antibodies (ANCA) are usually directed against myeloperoxidase (MPO) are present in up to 40% of patients affected (Sinico et al., 2005).

Genetic background supports the distinction between MPO-ANCA positive and negative patients. MPO-ANCA positive EGPA is an eosinophilic autoimmune disease sharing HLA-DQ association with MPO-ANCA positive ANCA associated vasculitis (e.g. certain forms of microscopic polyangiitis and granulomatosis with polyangiitis), while ANCA-negative EGPA may instead have a mucosal/barrier dysfunction origin (Lyons et al., 2019). EGPA is characterized by a Type 2 immune response with elevation of circulating IgG4, which could be mediated by IL-10, and total IgE, mediated by IL-4 (Berti et al., 2020a). In comparison to eosinophilic asthma, elevation of eosinophils are significantly higher, usually above  $1.5 \times 10^9/L$  or 10% of the leukocyte fraction.

EGPA typically develops in three partially overlapping phases: a prodromal phase potentially lasting for years characterized by respiratory tract symptoms, asthma and rhinosinusitis; a second phase with blood eosinophilia, eosinophilic tissue infiltration and eosinophil-induced organ inflammation; and a third phase dominated by systemic necrotizing vasculitis (Groh et al., 2015). Interestingly, the presence of respiratory atopy the year before the diagnosis of vasculitis was more often associated with severe asthma and a greater use of oral corticosteroids (Berti et al., 2018). When the diagnosis of EGPA is established, the clinical course can range from an acute, self-limited process to progressive multiorgan dysfunction with significant morbidity and mortality (Samson et al., 2013).

Surrogates of a vasculitic process include purpura, peripheral nervous system involvement (e.g. mononeuritis multiplex), scleritis, alveolar hemorrhage and glomerulonephritis. In addition to the vasculitic manifestations, other signs and symptoms due to eosinophil-related inflammation are reported as well, including pulmonary infiltrates, pleural effusion, urticarial papules, eosinophilic tubulointerstitial nephritis and eosinophilic myocarditis (Groh et al., 2015). International criteria exists to classify the diagnosis, that needs to be applied in patients with vasculitis (Lanham criteria, ACR criteria, CHCC definition,

ERS and GERM“O”P criteria, MIRRA trial criteria), summarized in (Berti et al., 2020a).

Data reported in the main series of patients highlighted EGPA clinical heterogeneity, suggesting that ANCA-test might distinguish different phenotypes, with MPO-positivity correlating with a “vasculitic” phenotype and an increased relapse rate, while the prognosis of ANCA-negative “eosinophilic” patients seems to be poorer, due to more frequent cardiomyopathy and cardiac death (Samson et al., 2013; Durel et al., 2016; Guillevin et al., 2011; Comarmond et al., 2013). Yet, this dichotomization between “vasculitic” vs. “eosinophilic” phenotypes is perhaps too simplistic, and it is likely that some disease manifestations could rather be due to both pathophysiological processes.

Recent and less recent cohort studies showed a high rate of sequelae and poor long-term outcomes after the resolution of vasculitis, regardless the treatment chosen (Samson et al., 2013; Comarmond et al., 2013). The main sequelae are chronic airway obstruction (65–85%), neurologic damage and especially peripheral neuropathy (40%), chronic rhinitis (35%) and chronic sinusitis (19%), severe lung disease (17%), chronic kidney disease (0–13%) and chronic heart failure (10%) (Rothenberg and Hogan, 2006). In particular, long-term poorly controlled asthma and persistent airflow obstruction may be present in more than 40% of EGPA, thereby leading to significant glucocorticoid-related side effects which negatively affect patient global health status and quality of life (Berti et al., 2020b).

### 3.3. Hypereosinophilic syndromes

Hypereosinophilic syndrome (HES) is a heterogeneous group of conditions, characterize by persistent eosinophilia that is associated with damage to multiple organs (Rosenberg et al., 2013). More specifically, HES can be defined as: hypereosinophilia (circulating eosinophil count  $>1.5 \times 10^9$ /mL documented on at least 2 occasions) or marked tissue eosinophilia, and clinical manifestations directly attributable to the eosinophilia or presumed to be consequence of eosinophilia and for which no alternative cause can be identified (Klion, 2015a).

This definition is broad and it captures all patients with clinical manifestations due to eosinophilia regardless of the underlying etiology. The following classification categories have been proposed (Kay, 2015): myeloproliferative HES, including in FIP1L1-PDGFR-A associated eosinophilic myeloproliferative neoplasms (Lombardi, 1996); lymphocytic variant HES, in which an aberrant or clonal lymphocyte population drives eosinophilia through the production of eosinophilopoietic cytokines (Kanda et al., 2021); overlap HES restricted to a single organ (Choi et al., 2020); secondary HES in the setting of a distinct diagnosis (i.e., parasitic helminth infection, drug hypersensitivity and primary immunodeficiency) in which eosinophilia has been described in a subset of affected patients (Januskevicius et al., 2020); familial HES, a rare autosomal dominant disorder; and (Mesnil et al., 2016) idiopathic HES (Klion, 2015b). This classification system slightly differs from with the 2008 WHO guidelines, which include myeloid and lymphoid neoplasms in the same category, on the basis of a variety of genetic, histopathologic, and clinical criteria.

Regardless this etiological heterogeneity, clinical signs and symptoms greatly overlaps with those defined as “manifestations due to eosinophil-related inflammation” abovementioned for EGPA, thus being often difficult to distinguish between HES and EGPA. Features helping to differentiate between EGPA and HES are a lower prevalence of asthma and a higher prevalence of splenomegaly and lymph nodes enlargement in HES. In a retrospective analysis of 166 patients with blood eosinophilia  $>1.0 \times 10^9$ /L and systemic manifestations, serum C-reactive protein levels could be a reliable biomarker in patients with eosinophilic asthma and systemic manifestations, able to distinguish between EGPA and idiopathic HES. therefore, in the absence of confounding factors (infection, venous thrombosis, malignancy, ...) low C-reactive protein levels (i.e.  $< 36$  mg/L) are suggestive of idiopathic HES rather than EGPA (Leurs et al., 2019).

### 3.4. Other eosinophilic-associated inflammatory conditions

Other less common eosinophilic-associated inflammatory disorders are eosinophilic chronic rhinosinusitis with nasal polyps, eosinophilic esophagitis or gastroenteritis or colitis, eosinophilic pneumonia, eosinophilic cystitis, and eosinophilic fasciitis (Rosenberg et al., 2013; O’Sullivan and Bochner, 2018; Furuta and Katzka, 2015). These disorders are less recognized as separated eosinophilic clinical entities, being often complications of other diseases (e.g. allergies), but when possible causes or confounders are excluded, this definition applies, making these conditions potentially eligible for eosinophilic targeted treatments.

## 4. Implications for biological therapies of eosinophilic-associated inflammatory conditions

The development of biological agents targeting IL-5, IL-4 and IL-13 has provided opportunities to treat patients with eosinophilic-associated inflammatory conditions whose severity or symptoms are driven by eosinophil biology (Table 4). Most studies evaluating these biologics were conducted initially in asthmatic patients by selecting those with raised blood eosinophil numbers (Koenderman et al., 2000). In peripheral blood, eosinophils constitute less than 5% of all leukocytes (Brigden and Graydon, 1997). In absolute counts, blood eosinophils are reported to range from  $0.05\text{--}0.5 \times 10^9$  cells/L (or 5–500 cells/ $\mu$ L), and eosinophilia is traditionally defined by values above the upper normal values (Valent et al., 2012). Large epidemiological study now identifies normal blood eosinophil counts in health to be around 100 cells/ $\mu$ L, once risk factors excluded. In most of these studies on healthy subjects, as many as 75% of the individuals studied had an eosinophil count below 210 cells/ $\mu$ L (Hartl et al., 2020; Vedel-Krogh, 2020).

First, the existence of distinct eosinophils subtypes that are differently involved in these conditions could give important data for better disease management. Importantly, only rEos were described as IL-5 independent cells (Von Hulst, 2021). The data suggest that basal levels of eosinophils left after absolute IL-5 depletion (mepolizumab or

**Table 4**

Biological agents that directly or indirectly target crucial eosinophil pathways, discussed in this article.

Biological Agent	Specifics	Molecular Target(s)	Approved Indications
Mepolizumab	Humanized IgG1 given subcutaneously	IL-5	Asthma (age 12 and older) EGPA (age 6 and older) HES (age 12 and older) CRSwNP (age 18 and older)
Reslizumab	Humanized IgG4 given intravenously	IL-5	Asthma age 18 and older (and others)
Benralizumab	Humanized afucosylated IgG1 given subcutaneously	IL-5R $\alpha$	Asthma (age 12 and older)
Dupilumab	Human IgG4 given subcutaneously	IL-4R $\alpha$ , blocking both IL-4 and IL-13	Atopic dermatitis (age 12 and older) Asthma (age 12 and older) CRSwNP (age 12 and older)
Lebrikizumab	Humanized IgG4 given intravenously	IL-13	Ongoing studies for Atopic Dermatitis

Abbreviations: IL = interleukin; R $\alpha$  = receptor subunit alpha; EGPA = eosinophilic granulomatosis with polyangiitis; HES = hypereosinophilic syndrome; CRSwNP = chronic rhinosinusitis with nasal polyps.

reslizumab) are a steady-state rEos population, and anti-IL-5 treatment affected eosinophils are in inflammatory processes involved iEos (Januskevicius et al., 2020; Mesnil et al., 2016; Abdala-Valencia et al., 2016, 2018). A deeper depletion of eosinophils, eliminating also the rEos population might disturb lung homeostasis. In the field of precision medicine, the different biology of eosinophil subtypes should be taken into account, and a truly targeted therapeutic intervention should be ideally able to distinguish between rEos and iEos. Second, a better characterization of the predominant pathway underlying the eosinophilic inflammation in the single patient could help: there are cases in which asthmatic patients have a modest hypereosinophilia and increased FeNO, or a normalization of peripheral eosinophils during anti-IL-5 treatment with suboptimal asthma control and increased FeNO, suggesting a predominance of IL-13 activation that might respond better to IL-13 targeted treatment (Erzurum and Gaston, 2012). Similarly, high circulating IgE levels (in addition to eosinophilia) in asthmatic patients, may suggest an hyperactivation of IL-4 pathway. Third, markers of type 2 inflammation don't always predict responsiveness to corticosteroids. For example, in patients with eosinophilic asthma, the persistence of sputum eosinophilia despite high dose of corticosteroid identifies a subgroup of patients that despite Th2 inflammation are unresponsive to corticosteroids (17% of the totals in a recent study) (Berthon et al., 2017; Dunican and Fahy, 2017). More in general, these type of patients with persistent circulating or organ-limited eosinophilia despite corticosteroid treatment, can be those that better benefit of anti-eosinophil targeted treatments. In addition, the availability of different agents with different ways of administrations, half-life, and mechanisms of actions is another element that can be weighed to tailor the choice of the anti-eosinophil targeted treatment on the single patient.

Several of these anti-eosinophil targeted treatments have been already approved (especially for asthma), while others are pending. It has been documented that the impact of anti-IL5 biological agents on blood and tissue levels of eosinophils is not equal, likely depending on the different mechanism of action. In recent years two monoclonal antibodies (mepolizumab and reslizumab) directed against IL-5 and one monoclonal antibody directed against the alpha-subunit of the IL-5 receptor (IL-5R) (benralizumab) have been developed.

#### 4.1. ANTI-IL5-RECEPTOR agents

Benralizumab is a monoclonal antibody that binds to IL-5R via its Fab domain, blocking the binding of IL-5 to its receptor and resulting in inhibition of eosinophil differentiation and maturation in bone marrow. In addition, benralizumab is able to bind through its afucosylated Fc domain to the RIIIa region of the Fcγ receptor on NK cells, macrophages, and neutrophils, thus strongly inducing antibody-dependent, cell-mediated cytotoxicity in both rEos and iEos. This double function of benralizumab induces almost complete fast and maintained depletion of eosinophils that is much greater than that induced by other monoclonal antibodies targeting the IL-5 pathway, such as mepolizumab and reslizumab (Kelly et al., 2017). Laviolette et al. documented that in asthmatics treated with the subcutaneous benralizumab (patients randomized to 100 mg or 200 mg, for 3 months) eosinophil counts decrease with a median value of 95.8% in the airways (day 84; placebo, 46.7%;  $P = 0.06$ ), 89.9% in sputum (day 28) and a 100% in blood counts (day 84), even if the extent of decrease in tissue eosinophil showed individual variability (Laviolette et al., 2013). Moreover, eosinophils were not detectable in bone marrow of benralizumab-treated subjects (day 28,  $n = 4$ ) (Laviolette et al., 2013). In addition, a mechanistic study showed that lung eosinophils of bronchoalveolar lavage of patients downregulate IL-5R, therefore it remains unclear if antibody dependent cytotoxicity (ADCC) can occur in lung tissue (Kelly et al., 2017). A deep and persistent depletion of blood eosinophils after chronic administration of benralizumab was observed also in patients with severe, uncontrolled asthma in the BORA study (Busse et al., 2019). Similar results

were observed more recently with the approved sub-cutaneous dose of benralizumab (30 mg), resulting in an almost complete depletion of eosinophils in both blood and sputum after 12 weeks of treatment (Sehmi et al., 2018; Nair et al., 2017).

Of note, benralizumab, has also been investigated in EGPA in a phase II study, facilitating oral corticosteroid reduction and reducing exacerbations in EGPA, while being well tolerate. These positive signals were probably due to its ability to induce profound eosinophil depletion (antibody-dependant cytotoxicity of IL-5Rα-bearing cells) (Guntur et al., 2021). Larger controlled trials are warranted to further evaluate the role of benralizumab in EGPA, currently ongoing.

#### 4.2. ANTI-IL5 agents

Reslizumab, administered intravenously at a dose of 3 mg/kg with some adjustment every 4 weeks, has been approved for severe eosinophilic asthma thanks to two multicenter large phase III trials on uncontrolled asthma with elevated blood eosinophil counts (Castro et al., 2015). In both studies, a significant reduction of asthma exacerbation with reslizumab was observed (rate ratio study 1 0.50 [95% CI 0.37–0.67]; study 2: 0.41 [0.28–0.59]; both  $p < 0.0001$ ) compared with those receiving placebo.

Mepolizumab is a fully humanized monoclonal antibody IgG1 specific for human IL-5. Mepolizumab blocks the binding of human IL-5, impairing its ligation to the α-chain of the IL-5Rα complex, which is mainly expressed on the eosinophil cell surface. Flood Page et al. already demonstrated in a 2003 paper that mepolizumab produced a median decrease from baseline of 55% for airway eosinophils, 52% for bone marrow eosinophils, and 100% for blood eosinophils in twenty-four patients with mild asthma that received three intravenous doses of either 750 mg of mepolizumab or placebo in a randomized, double-blind, parallel-group fashion over 20 weeks (Flood-Page et al., 2003). Therefore, the administration at high doses of mepolizumab does not deplete airway or bone marrow eosinophils. Blood pharmacodynamic responses showed that a subcutaneous dose of 100 mg of mepolizumab administered subcutaneously provides the desired pharmacology to neutralize serum IL-5 levels and reduce blood eosinophil counts to approximately 40 cells/mL in asthmatic patients, which corresponds to a low normalized blood eosinophil count (Yancey et al., 2017). Indeed, similar data have also been observed in the multicenter, open-label, long-term, Phase IIIb COSMEX study. In this study patients received mepolizumab 100 mg subcutaneously every 4 weeks as add-on therapy for up to 172 weeks with eosinophils blood levels constantly detected at levels of about 50 cells/μL (Geometric Mean Blood Eosinophil Count) (Khurana et al., 2019). Pavord et al. demonstrated in the DREAM study that mepolizumab, at the 750-mg intravenous dose, has determined comparable reductions of 88% in blood and sputum eosinophil counts; while for the 250-mg intravenous dose, the reduction in blood eosinophil counts was 86% compared with 65% for sputum, and for the 75 mg intravenous dose, the reduction in blood eosinophil counts was 78% compared with 32% for sputum (Pavord et al., 2012). A similar trend on the decrease of eosinophils in induced sputum was also observed in the study of Pouliquen et al. (2015). Furthermore, mepolizumab does not alter levels of eosinophils, T cells, and mast cells in the duodenal mucosa in eosinophilic esophagitis (Conus et al., 2010). Mepolizumab had no effect on the duodenal infiltration of eosinophils, T cells, and mast cells in subjects participating in the first placebo-controlled clinical study performed in adult patients with eosinophilic esophagitis (Straumann et al., 2010). Whereas “inflammatory” eosinophils at the site of inflammation are reduced by approximately 50%, physiologic eosinophil infiltration, at least in the duodenum, is not affected, even at the high 1500 mg dose level used in this study. In addition, the anti-IL-5 therapy did not change the expression levels of IL-5Ra and the eosinophil activation markers CD25 and IL-13, suggesting that the relative proportions of potential functionally different eosinophil subgroups also did not change. Finally, in another study that assessed outcomes after



cessation of mepolizumab in severe eosinophilic asthmatic patients, the blood eosinophils count resulted higher in 0–3 months ( $p < 0.001$ ) in treated patients, demonstrating that the inhibitory effect of mepolizumab on eosinophils is rapidly reversible upon its cessation (Haldar et al., 2014). Overall, this indicates that Mepolizumab, which targets the IL-5 molecule does not impact on tissue eosinophils as strong as benralizumab, which target the IL-5 receptor.

Having demonstrated efficacy in severe asthma, mepolizumab have been tested in patients with nasal polyposis with or without asthma, showing to reduce nasal and sinus nasal polyp burden monitored by nasal endoscopy and to improve nasal symptoms. The study SYNAPSE enrolled 407 patients were randomized to 100 mg mepolizumab subcutaneously or placebo once every 4 weeks, in addition to standard of care. This study demonstrate efficacy in larger patient populations with current, refractory severe chronic rhinosinusitis with nasal polyps, leading to the registration for the indication nasal polyps (Han et al., 2021).

Mepolizumab has been approved for the treatment of refractory or relapsing EGPA by EMA in September 2021. One of the most important studies involved the subcutaneous administration of 300 mg every four weeks, i.e. 3 times higher than the asthma dose (Wechsler et al., 2017). Interestingly, no specific dose-finding studies were performed for EGPA. This multicenter, randomized controlled trial enrolled 136 subjects with stable disease on a stable prednisolone or prednisone dose along with standard EGPA care. Treatment with mepolizumab led to a greater likelihood of having 24 weeks of disease remission (28% on active drug versus 3% on placebo, with similar proportions at 48 weeks), which was the primary endpoint, and an average daily dose of prednisolone or prednisone of  $\leq 4$  mg per day during the last four weeks of the 52 of observations was achieved in 44% of the mepolizumab group versus 7% of placebo. This highlights the steroid sparing potential of this treatment in EGPA. Even if better than placebo, remission did not occur in 47% of the participants in the mepolizumab group versus 81% of those in the placebo group. Only 13% of patients in both arms were ANCA positive at enrollment (19% were ANCA-positive at some time in the course of the disease), which clearly represents a limitation of this study. In addition, none of the patients received mepolizumab as first-line therapy, and overall, mepolizumab's ability to curb vasculitis manifestations remains unclear.

Similarly, a phase III trial led to mepolizumab approval by EMA for FIP1-like-1-platelet-derived growth factor receptor  $\alpha$ -negative HES, given subcutaneously at a dose of 300 mg every 4 weeks versus placebo, and added on HES therapy (Roufosse et al., 2020). While as safe as standard of care, the proportion of patients experiencing 1 or more flares/withdrawing from the study was 50% lower with mepolizumab versus placebo, and the open-label extension of the trial confirmed long-term control of disease flares, blood eosinophil counts, plus reductions in corticosteroid use (Gleich et al., 2021).

#### 4.3. ANTI-IL4/IL13 agents

Dupilumab is a recombinant human IgG4 antibody to the IL-4 receptor. There are 2 types of IL-4 receptors: the type 1 receptor, which is composed of the IL-4 chain (IL-4R $\alpha$ ) and a  $\gamma$  chain ( $\gamma$ C), and the type 2 receptor, which is composed of the IL-4R $\alpha$  chain and the  $\alpha$ 1 chain of the IL-13 receptor (IL-13R $\alpha$ 1) (Sastre and Dávila, 2018). The type 1 receptor can be activated by IL-4 and the type 2 receptor can be activated by both IL-4 and IL-13. By blocking the IL-4R alpha subunit, dupilumab inhibits IL-4 and IL-13 cytokine-induced responses, including the release of proinflammatory cytokines, chemokines, and immunoglobulin E. Transient eosinophilia is a known side effect of dupilumab and transient increases in eosinophils have been observed in dupilumab clinical studies. The treatment with dupilumab in asthmatic subjects was associated with an increase in blood eosinophils with a maximum increase occurring at approximately 16–20 weeks after starting therapy, and the mean percent increase of approximately 10% and fifty-two of 1264

treated subjects (4.1%) developed a treatment emergent eosinophilia (Castro et al., 2018). All subjects with an eosinophil response recovered, some of them with systemic corticosteroid therapy and dupilumab discontinuation and others improving despite continuation of the dupilumab. In subjects who remained on dupilumab, the eosinophil count generally decreased to baseline by 28–52 weeks. In a French multicentre, retrospective *real-life* cohort study (treated patients: 64) a hypereosinophilia  $\geq 1500/\text{mm}^3$  was observed at least once during follow-up in 16 patients (25%), persisting after 6 months in 8 (14%) of them. Increase in blood eosinophil count did not modify the clinical response during the study period (Dupin et al., 2020). Transient eosinophilia is felt to be self-limited due to the drug's effect on the Th2 immune response and its subsequent effect on blunting migration of eosinophils into target tissues. As a result, it is not commonly thought to cause eosinophilic disease processes, due to the subsequent sequestration of eosinophils in the blood and inhibition of their migration into target tissues. Two theories have been proposed for this finding of increased eosinophilia in response to dupilumab: (a) eosinophilia could represent a transient rebound elevation in response to eosinophil-promoting mediators such as IL-5, as a result of blocking IL-4 and IL-13 (Chung, 2016), and (b) as IL-4 and IL-13 recruit and facilitate eosinophil migration into tissues, and their inhibition may cause eosinophil accumulation in the peripheral blood (Darveaux and Busse, 2015; Barranco et al., 2017). However, there have been some cases of marked increase of eosinophilia and tissue accumulation of eosinophils during dupilumab therapy. One patient developed hypereosinophilic syndrome; the treatment was discontinued, and corticosteroid therapy was administered with immediate improvement (Wenzel et al., 2013). It has been also reported a case of eosinophilic pneumonia with sustained hypereosinophilia (maximum eosinophil count of 2080/ $\mu\text{L}$ ) associated with the use of dupilumab (Menzella et al., 2019).

The dupilumab trial findings on uncontrolled asthma are in contrast to those of two large international phase 3 studies assessing lebrikizumab (LAVOLTA I and II), in which patients were randomized to receive placebo or lebrikizumab (Hanania et al., 2016). Overall primary endpoint was not met, i.e. the reduction in exacerbations over a one-year period in subjects with high blood eosinophil counts or periostin levels: one study reached statistical significance while the other one did not. Taken all together, one could speculate that in asthma targeting both IL-4 and IL-13 as dupilumab does is better than targeting IL-13 alone as done by lebrikizumab, or alternatively that targeting the receptor alpha rather than the cytokine could impact more on eosinophil biology.

#### 5. Conclusions and future perspectives

Overall, data suggest that eosinophils are heterogeneous and that different eosinophil subpopulations exist *in vivo*, i.e. rEos and iEos. Besides their roles in Th2-oriented diseases, eosinophils also regulate homeostatic processes at steady state, thereby challenging the exclusive paradigm of the eosinophil as a destructive and inflammatory cell (Mesnil et al., 2016). These findings may thus have an impact on the efficacy of eosinophil-targeted therapy.

Distinctive eosinophil subpopulations exert different functions and the identification of functional subgroups and specific markers is a mandatory research objective. Application of precision medicine by proteomics, transcriptomics, and metabolomics to eosinophil biology may help to promote further advances in this research field.

Therapeutic intervention with biological agents that totally deplete tissues and circulating eosinophils or, *vice versa*, maintain a minimal proportion of eosinophils, particularly those residents in tissues or rEos, could therefore have a very different impact, especially when considering the administration of these therapies for prolonged periods. Similarly, a wider use of routine biomarkers of Type 2 inflammation (circulating eosinophils, and organ-specific eosinophils levels such as eosinophil count in sputum, bronchoalveolar lavage, tissue biopsy; total circulating IgE levels or the use of FeNO) can help in to choose the best

eosinophil-targeted approach among the increasing therapeutic armamentarium of biological agents.

The identification of predictors for the efficacy of anti-eosinophil treatments is therefore a key objective and might further result in the identification of new functional eosinophil subsets as well as new categories of eosinophil-associated clinical disorders.

## Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

## Authors' contributions

Writing the original draft, investigation, literature analysis, conceptualization, figures: CL. Contribution to writing and conceptual framing of the article: AB. All the Authors critically review and approved the final manuscript.

## Availability of data and materials

Not applicable.

## Ethics approval and consent to participate

Not applicable.

## Consent for publication

Not applicable.

## Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## References

- Abdala-Valencia, H., Loffredo, L., Misharin, A., Berdnikov, S., 2016. Phenotypic plasticity and targeting of Siglec-F high CD 11c low eosinophils to the airway in a murine model of asthma. *Allergy* 71, 267–271.
- Abdala-Valencia, H., Coden, M.E., Chiarella, S.E., Jacobsen, E.A., Bochner, B.S., Lee, J.J., et al., 2018 Jul. Shaping eosinophil identity in the tissue contexts of development, homeostasis, and disease. *J. Leukoc. Biol.* 104 (1), 95–108.
- Adamko, D.J., Yost, B.L., Gleich, G.J., Fryer, A.D., Jacoby, D.B., 1999 Nov. Ovalbumin sensitization changes the inflammatory response to subsequent parainfluenza infection. Eosinophils mediate airway hyperresponsiveness, m(2) muscarinic receptor dysfunction, and antiviral effects. *J. Exp. Med.* 190 (10), 1465–1478.
- Amelink, M., de Groot, J.C., de Nijs, S.B., et al., 2013. Severe adult-onset asthma: a distinct phenotype. *J. Allergy Clin. Immunol.* 132, 336–341.
- Aoki, A., Hirahara, K., Kiuchi, M., Nakayama, T., 2021 Jan. Eosinophils: cells known for over 140 years with broad and new functions. *Allergol. Int.* 70 (1), 3–8. <https://doi.org/10.1016/j.alit.2020.09.002>.
- Bagnasco, D., Ferrando, M., Varricchi, G., et al., 2017. Anti-interleukin 5 (IL-5) and IL-5Ra biological drugs: efficacy, safety, and future perspectives in severe eosinophilic Asthma. *Front. Med.* 4, 135. <https://doi.org/10.3389/fmed.2017.00135>.
- Barnes, P.J., 2008. Immunology of asthma and chronic obstructive pulmonary disease. *Nat. Rev. Immunol.* 8 (3), 183–192.
- Barranco, P., Phillips-Angles, E., Dominguez-Ortega, J., Quirce, S., 2017. Dupilumab in the management of moderate-to-severe asthma: the data so far. *Therapeut. Clin. Risk Manag.* 13, 1139–1149.
- Berek, C., 2015. Eosinophils: important players in humoral immunity. *Clin. Exp. Immunol.* 183, 57–64.
- Berthon, B.S., Gibson, P.G., Wood, L.G., et al., 2017. A sputum gene expression signature predicts oral corticosteroid response in asthma. *Eur. Respir. J.* 49, 1700180.
- Berti, A., Volcheck, G.W., Cornec, D., Smyth, R.J., Specks, U., Keogh, K.A., 2018. Severe/uncontrolled asthma and overall survival in atopic patients with eosinophilic granulomatosis with polyangiitis. *Respir Med [Internet]* 142, 66–72.
- Berti, A., Boukhhal, S., Groh, M., Cornec, D., 2020a. Eosinophilic granulomatosis with polyangiitis: the multifaceted spectrum of clinical manifestations at different stages of the disease-. *Expet Rev. Clin. Immunol.* 16 (1), 51–61. <https://doi.org/10.1080/1744666X.2019.1697678>.
- Berti, A., Cornec, D., Casal Moura, M., Smyth, R.J., Dagna, L., Specks, U., Keogh, K.A., 2020b. Eosinophilic Granuloma-tosis with polyangiitis: clinical predictors of long-term Asthma severity. *May Chest* 157 (5), 1086–1099. <https://doi.org/10.1016/j.chest.2019.11.045>. Epub 2020 Jan 17.
- Brigden, M., Graydon, C., 1997. Eosinophilia detected by automated blood cell counting in ambulatory North American outpatients. Incidence and clinical significance. *Arch. Pathol. Lab Med.* 121, 963–967.
- Busse, W.W., Bleeker, E.R., FitzGerald, J.M., Ferguson, G.T., Barker, P., Sproul, S., et al., 2019. Long-term safety and efficacy of benralizumab in patients with severe, uncontrolled asthma: 1-year results from the BORA phase 3 extension trial. *The Lancet Respiratory* 7 (Issue 1), 46–59. [https://doi.org/10.1016/S2213-2600\(18\)30406-5](https://doi.org/10.1016/S2213-2600(18)30406-5).
- Castro, M., Zangrilli, J., Wechsler, M.E., Bateman, E.D., Brusselle, G.G., Bardin, P., Murphy, K., Maspero, J.F., O'Brien, C., Korn, S., 2015 May. Reslizumab for inadequately controlled asthma with elevated blood eosinophil counts: results from two multicentre, parallel, double-blind, randomised, placebo-controlled, phase 3 trials. *Lancet Respir. Med.* 3 (5), 355–366. [https://doi.org/10.1016/S2213-2600\(15\)00042-9](https://doi.org/10.1016/S2213-2600(15)00042-9). Epub 2015 Feb 23.
- Castro, M., Corren, J., Pavord, I.D., et al., 2018. Dupilumab efficacy and safety in moderate-to-severe uncontrolled asthma. *N. Engl. J. Med.* 378, 2486–2496.
- Choi, Y., Sim, S., Park, H.-S., 2020. Distinct functions of eosinophils in severe asthma with type 2 phenotype: clinical implications. *Korean J. Intern. Med. (Korean Ed.)* 35, 823–833.
- Chu, V.T., Beller, A., Rausch, S., et al., 2014. Eosinophils promote generation and maintenance of immunoglobulin-A-expressing plasma cells and contribute to gut immune homeostasis. *Immunity* 40, 582–593.
- Chung, K.F., 2016. Dupilumab: a potential new treatment for severe asthma. *Lancet* 388 (10039), 3–4.
- Churg, J., Strauss, L., 1951. Allergic granulomatosis, allergic angiitis, and periarteritis nodosa. *Am. J. Pathol.* 27 (2), 277–301, 1951/03/01.
- Comarmond, C., Pagnoux, C., Khellaf, M., Cordier, J.F., Hamidou, M., Viallard, J.F., et al., 2013. Eosinophilic granuloma-tosis with polyangiitis (Churg-Strauss): clinical characteristics and long-term follow-up of the 383 patients enrolled in the French Vasculitis Study Group cohort. *Arthritis Rheum.* 65 (1), 270–281, 2012/10/10.
- Conus, S., Straumann, A., Bettler, E., Simon, H.-U., 2010. Mepolizumab does not alter levels of eosinophils, T cells, and mast cells in the duodenal mucosa in eosinophilic esophagitis. *J. Allergy Clin. Immunol.* 126 (1), 175–177.
- Darveau, J., Busse, W.W., 2015. Biologics in asthma – the next step toward personalized treatment. *J. Allergy Clin. Immunol. Pract.* 3 (2), 152–160.
- de Groot, J.C., Storm, H., Amelink, M., et al., 2016. Clinical profile of patients with adult-onset eosinophilic asthma. *ERJ Open Res.* 2 (2) <https://doi.org/10.1183/23120541.00100-2015>, 00100-2015-100-2015.
- Drake, M.G., Bivins-Smith, E.R., Proskocil, B.J., Nie, Z., Scott, G.D., Lee, J.J., et al., 2016. Human and mouse eosinophils have antiviral activity against parainfluenza virus. *Am. J. Respir. Cell Mol. Biol.* 55, 387–394.
- Duez, C., Dakkhama, A., Tomkinson, A., Marquillies, P., Balhorn, A., Tonnel, A.B., et al., 2004. Migration and accumulation of eosinophils toward regional lymphnodes after airway allergen challenge. *J. Allergy Clin. Immunol.* 114, 820–825.
- Dunican, E.M., Fahy, J.V., 2017 Jun 29. Asthma and corticosteroids: time for a more precise approach to treatment. *Eur. Respir. J.* 49 (6), 1701167. <https://doi.org/10.1183/13993003.01167-2017>.
- Dupin, C., Belhadi, D., Guilleminault, L., Gamez, A.-S., Berger, P., De Blay, F., et al., 2020 Jul. Effectiveness and safety of dupilumab for the treatment of severe asthma in a real-life French multi-centre adult cohort. *Clin. Exp. Allergy* 50 (7), 789–798. <https://doi.org/10.1111/cea.13614>. Epub 2020 May 29.
- Durel, C.A., Berthiller, J., Caboni, S., Jayne, D., Ninet, J., Hot, A., 2016. Long-term followup of a multicenter cohort of 101 patients with eosinophilic granulomatosis with polyangiitis (Churg-Strauss). *Arthritis Care Res.* 68 (3), 374–387, 2015/09/01.
- Erzurum, S.C., Gaston, B.M., 2012 Sep. Biomarkers in asthma: a real hope to better manage asthma. *Clin. Chest Med.* 33 (3), 459–471.
- Flood-Page, P.T., Menzies-Gow, A.N., Kay, A.B., Robinson, D.S., 2003. Eosinophil's role remains uncertain as anti-interleukin-5 only partially depletes numbers in asthmatic airway. *Am. J. Respir. Crit. Care Med.* 167, 199–204.
- Furuta, G.T., Katzka, D.A., 2015 Oct 22. Eosinophilic esophagitis. *N. Engl. J. Med.* 373 (17), 1640–1648. <https://doi.org/10.1056/NEJMra1502863>.
- Gleich, G.J., Roufosse, F., Chupp, G., Faguer, S., Walz, B., Reiter, A., Yancey, S.W., Bentley, J.H., Steinfeld, J., HES Mepolizumab Study Group, 2021 Aug 10. S2213-2198 J. Allergy Clin. Immunol. Pract. (21). <https://doi.org/10.1016/j.jaip.2021.07.050>, 00891-6.
- Gonlugur, U., Gonlugur, T.E., 2006. Non-allergic eosinophilic inflammation. *Immunol. Invest.* 35 (1), 29–45. <https://doi.org/10.1080/08820130500496779>.
- Groh, M., Pagnoux, C., Baldini, C., Bel, E., Bottero, P., Cottin, V., et al., 2015. Eosinophilic granulomatosis with polyangiitis (Churg-Strauss) (EGPA) Consensus Task Force recommendations for evaluation and management. *Eur. J. Intern. Med.* 26 (7), 545–553, 2015/05/15.
- Guillemin, L., Pagnoux, C., Seror, R., Mahr, A., Mouthon, L., Le Toumelin, P., 2011. The Five-Factor Score revisited: assessment of prognoses of systemic necrotizing vasculitides based on the French Vasculitis Study Group (FVSG) cohort. *Med* 90 (1), 19–27, 2011/01/05.
- Guntur, V.P., Manka, L.A., Denson, J., Dunn, R.M., Dollin, Y.T., Gill, M., Kolakowski, C., Strand, M.J., Wechsler, M.E., 2021 Mar. Benralizumab as a steroid-sparing treatment option in eosinophilic granulomatosis with polyangiitis. *J. Allergy Clin. Immunol. Pract.* 9 (3), 1186–1193. <https://doi.org/10.1016/j.jaip.2020.09.054>. Epub 2020 Oct 14.

- Haldar, P., Brightling, C.E., Singapuri, A., Hargadon, B., Gupta, S., Monteiro, W., et al., 2014. Outcomes after cessation of mepolizumab therapy in severe eosinophilic asthma: a 12-month follow-up analysis. *J. Allergy Clin. Immunol.* 133 (3), 921–923.
- Han, J.K., Bachert, C., Fokkens, W., Desrosiers, M., Wagenmann, M., Lee, S.E., Smith, S. G., Martin, N., Mayer, B., Yancey, S.W., Sousa, A.R., Chan, R., Hopkins, C., 2021 Oct. SYNAPSE study investigators. Mepolizumab for chronic rhinosinusitis with nasal polyps (SYNAPSE): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Respir. Med.* 9 (10), 1141–1153. [https://doi.org/10.1016/S2213-2600\(21\)00097-7](https://doi.org/10.1016/S2213-2600(21)00097-7). Epub 2021 Apr 16. PMID: 33872587.
- Hanania, N.A., Korenblat, P., Chapman, K.R., Bateman, E.D., Kopecky, P., Paggiaro, P., et al., 2016. Efficacy and safety of lebrikizumab in patients with uncontrolled asthma (LAVOLTA I and LAVOLTA II): replicate, phase 3, randomised, double-blind, placebo-controlled trials. *Lancet Respir. Med.* 4, 781–796.
- Hansel, T.T., Braunstein, J.B., Walker, C., Blaser, K., Bruijnzeel, P.L., Virchow Jr., J.C., et al., 1991. Sputum eosinophils from asthmatics express ICAM-1 and HLA-DR. *Clin. Exp. Immunol.* 86, 271–277.
- Hara, T., Miyajima, A., 1996. Function and signal transduction mediated by the interleukin 3 receptor system in hematopoiesis. *Stem Cell.* 14, 605–618.
- Hartl, S., Breyer, M.-K., Burghuber, O.C., et al., 2020. Blood eosinophil count in the general population: typical values and potential confounders. *Eur. Respir. J.* 55, 1901874. <https://doi.org/10.1183/13993003.01874-2019>.
- Impellizzeri, G., Marasco, G., Eusebia, L.H., Salfib, N., Bazzolia, F., Zagari, R.M., 2019 Jun. Eosinophilic colitis: a clinical review. *Dig. Liver Dis.* 51 (6), 769–773.
- Jacobsen, E.A., Helmers, R.A., Lee, J.J., Lee, N.A., 2012. The expanding role(s) of eosinophils in health and disease. *Blood* 120 (19), 3882–3890. <https://doi.org/10.1182/blood-2012-06-330845>.
- Januskevicius, A., Jurkeviciute, E., Janulaityte, I., Kalinauskaitė -Zukauske, V., Miliauskas, S., Malakauskas, K., 2020. Blood eosinophils subtypes and their survivability in asthma patients. *Cells* 9, 1248–1264.
- Johansson, M.W., 2014. Activation states of blood eosinophils in asthma. *Clin. Exp. Allergy* 44, 482–498.
- Jung, Y., 2015. Comparative analysis of dibutyric cAMP and butyric acid on the differentiation of human eosinophilic leukemia EoL-1 cells. *Immune Netw* 15, 313–318.
- Jung, Y., Rothenberg, M.E., 2014. Roles and regulation of gastrointestinal eosinophils in immunity and disease. *J. Immunol.* 193, 999–1005.
- Kanda, A., Yun, Y., Van Bui, D., Nguyen, L.M., Kobayashi, Y., Suzuki, K., et al., 2021 Jan. The multiple functions and subpopulations of eosinophils in tissues under steady-state and pathological conditions. *Allergol. Int.* 70 (1), 9–18. <https://doi.org/10.1016/j.alit.2020.11.001>.
- Kay, A.B., 2015. The early history of the eosinophil. *Clin. Exp. Allergy* 45, 575–582.
- Kelly, K.A., Esnault, S., Liu, L.Y., Evans, M.D., Johansson, M.W., Mathur, S., Mosher, D.F., Denlinger, L.C., Jarjour, N.N., 2017 Dec 1. Mepolizumab attenuates airway eosinophil numbers, but not their functional phenotype, in asthma. *Am. J. Respir. Crit. Care Med.* 196 (11), 1385–1395.
- Khurana, S., Brusselle, G.G., Bel, E.H., FitzGerald, Masoli, M., Korn, S., Kato, M., Albers, F.C., Bradford, E.S., Gilson, M.J., Price, R.G., Humbert, M., 2019. Long-term safety and clinical benefit of mepolizumab in patients with the most severe eosinophilic asthma: the COSMEX study. *Clin. Therapeut.* 41 (Number 10), 2041–2056.
- Kim, H.J., Jung, Y.J., 2020. The emerging role of eosinophils as multifunctional leukocytes in health and disease. *Immune netw* 20 (3), e24.
- Klion, A., 2015a. Eosinophilia: a pragmatic approach to diagnosis and treatment. *Hematology Am. Soc. Hematol Educ. Prog.* 2015, 92–97. <https://doi.org/10.1182/asheducation-2015.1.92>.
- Klion, A.D., 2015b. How I treat hypereosinophilic syndromes. *Blood* 126 (9), 1069–1077.
- Klose, C.S., Artis, D., 2016. Innate lymphoid cells as regulators of immunity, inflammation and tissue homeostasis. *Nat. Immunol.* 17, 765–774.
- Koenderman, L., Hassani, M., Mukherjee, M., Nair, P., 2000. Monitoring eosinophils to guide therapy with biologics in asthma: does the compartment matter? *Allergy*. <https://doi.org/10.1111/all.14700> dec. 10.
- Kroegel, C., Liu, M.C., Hubbard, W.C., Lichtenstein, L.M., Bochner, B.S., 1994. Blood and bronchoalveolar eosinophils in allergic subjects after segmental antigen challenge: surface phenotype, density heterogeneity, and prostanoid production. *J. Allergy Clin. Immunol.* 93, 725–734.
- Kuo, H.P., Yu, T.R., Yu, C.T., 1994. Hypodense eosinophil number relates to clinical severity, airway hyperresponsiveness and response to inhaled corticosteroids in asthmatic subjects. *Eur. Respir. J.* 7, 1452–1459.
- Lavine, K.J., 2020. Eosinophils confer protection following myocardial infarction. *JACC (J. Am. Coll. Cardiol.): Basic Translat. Sci.* 5 (7), 682–684.
- Lavoilette, M., Gossage, D.L., Gauvreau, G., Leigh, R., Olivenstein, R., Katial, R., Busse, W.W., Wenzel, S., Wu, Y., Datta, V., Kolbeck, R., Molino, N.A., 2013 Nov. Effects of benralizumab on airway eosinophils in asthmatic patients with sputum eosinophilia. *J. Allergy Clin. Immunol.* 132 (5), 1086–1096. <https://doi.org/10.1016/j.jaci.2013.05.020> e5.
- Lee, J.J., Jacobsen, E.A., McGarry, M.P., Schleimer, R.P., Lee, N.A., 2010. Eosinophils in health and disease: the LIAR hypothesis. *Clin. Exp. Allergy* 40, 563–575.
- Leurs, A., Chenivresse, C., Lopez, B., Gibier, J.-B., Clément, G., Groh, M., et al., 2019. C-Reactive protein as a diagnostic tool in differential diagnosis of hypereosinophilic syndrome and antineutrophil cytoplasmic antibody-negative eosinophilic granulomatosis with polyangiitis. *J. Allergy Clin. Immunol. Pract.* 7 (4), 1347–1351. <https://doi.org/10.1016/j.jaip.2018.10.002>. Apr.
- Li, N., Hua, J., 2017 Jan 10. Immune cells in liver regeneration. *Oncotarget* 8 (2), 3628–3639. <https://doi.org/10.18632/oncotarget.12275>.
- Li, Y., Wang, W., Lv, Z., et al., 2018. Elevated expression of IL-33 and TSLP in the airways of human asthmatics in vivo: a potential biomarker of severe refractory disease. *J. Immunol.* 200, 2253–2262.
- Lim, H.F., Nair, P., 2018. Airway inflammation and inflammatory biomarkers. *Semin. Respir. Crit. Care Med.* 39, 56–63, 3. Licari A, Castagnoli R, Brambilla I, et al. Asthma endotyping and biomarkers in childhood asthma. *Pediatr Allergy Immunol Pulmonol* 2018;31:44-5.
- Locksley, R.M., 2010. Asthma and allergic inflammation. *Cell* 140, 777–783.
- Lombardi, C., 1996. Eosinofili ed eosinofilia. *Fisiopatologia e nosografia clinica*, first ed. Città Studi Edizioni. 88-251-0108-9.
- Lombardi, C., Passalacqua, G., 2003 Jun 9. Eosinophilia and diseases: clinical revision of 1862 cases. *Arch. Intern. Med.* 163 (11), 1371–1373. <https://doi.org/10.1001/archinte.163.11.1371-b>.
- Lyons, P.A., Peters, J.E., Alberici, F., Liley, J., Coulson, R.M.R., Astle, W., et al., 2019. Genome-wide association study of eosinophilic granulomatosis with polyangiitis reveals genomic loci stratified by ANCA status. *Nature Communications* 10 (1), 5120. <https://doi.org/10.1038/s41467-019-12515-9>. Nov 12.
- Menzella, Francesco, et al., 2019. A case of chronic eosinophilic pneumonia in a patient treated with dupilumab. *Therapeut. Clin. Risk Manag.* 15, 869–871.
- Mesnil, C., Raulier, S., Paulissen, G., Xiao, X., Birrell, M.A., Pirotin, D., et al., 2016. Lung-resident eosinophils represent a distinct regulatory eosinophil subset. *J. Clin. Invest.* 126, 3279–3295.
- Metcalfe, D.D., Pawankar, R., Ackerman, S.J., Akin, C., Clayton, F., Falcone, F.H., et al., 2016. Biomarkers of the involvement of mast cells, basophils and eosinophils in asthma and allergic diseases. *World Allergy Organ J.* 9, 7.
- Mitchell, P.D., O'Byrne, P.M., 2017. Epithelial-derived cytokines in asthma. *Chest* 151, 1338–1344. <https://doi.org/10.1016/j.chest.2016.10.042>.
- Miyasato, M., Tsuda, S., Nakama, T., Kato, K., Kitamura, N., Nagaji, J., et al., 1996. Serum levels of eosinophil cationic protein reflect the state of in vitro degranulation of blood hypodense eosinophils in atopic dermatitis. *J. Dermatol.* 23, 382–388.
- Mould, A.W., Matthaei, K.I., Young, I.G., Foster, P.S., 1997. Relationship between interleukin-5 and eotaxin in regulating blood and tissue eosinophilia in mice. *J. Clin. Invest.* 99, 1064–1071.
- Nair, P., Wenzel, S., Rabe, K.F., Bourdin, A., Lugogo, N.L., Kuna, P., et al., 2017. Oral glucocorticoid-sparing effect of benralizumab in severe asthma. *N. Engl. J. Med.* 376, 2448–2458.
- O'Sullivan, J.A., Bochner, B.S., 2018 Feb. Eosinophils and eosinophil-associated diseases: an update. *J. Allergy Clin. Immunol.* 141 (2), 505–517.
- Pavord, I.D., Korn, S., Howarth, P., Bleeker, E.R., Buhl, R., Keene, O.N., Ortega, H., Chanez, P., 2012. Mepolizumab for severe eosinophilic asthma (DREAM): a multicentre, double-blind, placebo-controlled trial. *Lancet* 380, 651–659.
- Percopo, C.M., Dyer, K.D., Ochkur, S.I., Luo, J.L., Fischer, E.R., Lee, J.J., et al., 2014 Jan. Activated mouse eosinophils protect against lethal respiratory virus infection. *Blood* 123 (5), 743–752.
- Petsky, H.L., Cates, C.J., Kew, K.M., et al., 2018. Tailoring asthma treatment on eosinophilic markers (exhaled nitric oxide or sputum eosinophils): a systematic review and meta-analysis. *Thorax* 73, 1110–1119.
- Phipps, S., Lam, C.E., Mahalingam, S., Newhouse, M., Ramirez, R., Rosenberg, H.F., et al., 2007 Sep. Eosinophils contribute to innate antiviral immunity and promote clearance of respiratory syncytial virus. *Blood* 110 (5), 1578–1586.
- Poullieux, I.J., Kornmann, O., Barton, S.V., Price, J.A., Ortega, H.G., 2015. Characterization of the relationship between dose and blood eosinophil response following subcutaneous administration of mepolizumab. *Int. J. Clin. Pharm. Ther.* 53 (12), 1015–1027.
- Prin, L., Capron, M., Tonnel, A.B., Blety, O., Capron, A., 1983. Heterogeneity of human peripheral blood eosinophils: variability in cell density and cytotoxic ability in relation to the level and the origin of hypereosinophilia. *Int. Arch. Allergy Appl. Immunol.* 72, 336–346.
- Reichman, H., Moshkovits, I., Itan, M., Pasmannik-Chor, M., Vogl, T., Roth, J., et al., 2017 Aug. Transcriptome profiling of mouse colonic eosinophils reveals a key role for eosinophils in the induction of s100a8 and s100a9 in mucosal healing. *Sci. Rep.* 7 (1), 7117.
- Riggoni, C., Comberiat, P., Giovannini, M., Agache, I., Akdis, M., Alves-Correia, M., et al., 2020. A compendium answering 150 questions on COVID-19 and SARS-CoV-2. *Allergy* 75, 2503–2541.
- Roca, E., Ventura, L., Zattra, C.M., Lombardi, C., 2020 Sep 2. Eosinopenia: an early, effective, and relevant COVID-19 biomarker? *QJM*. <https://doi.org/10.1093/qjmed/hcaa259>.
- Rosenberg, H.F., Dyer, K.D., Foster, P.S., 2013. Eosinophils: changing perspectives in health and disease. *Nat. Rev. Immunol.* 13, 9–22.
- Roth, N., Städler, S., Lemann, M., Hösl, S., Simon, H.U., Simon, D., 2011 Nov. Distinct eosinophil cytokine expression patterns in skin diseases - the possible existence of functionally different eosinophil subpopulations. *Allergy* 66 (11), 1477–1486.
- Rothenberg, M.E., Hogan, S.P., 2006. The eosinophil. *Annu. Rev. Immunol.* 24, 147–174.
- Roufosse, F., Kahn, J.-E., Rothenberg, M.E., Wardlaw, A.J., Klion, A.D., Kirby, S.Y., Gilson, M.J., Bentley, J.H., Bradford, E.S., Yancey, S.W., Steinfeld, J., Gleich, G.J., 2020. Efficacy and safety of mepolizumab in hypereosinophilic syndrome: a phase III, randomized, placebo-controlled trial. *J. Allergy Clin. Immunol.* 146 (6), 1397–1405. <https://doi.org/10.1016/j.jaci.2020.08.037>. Epub 2020 Sep. 18.
- Rugeles, M.T., Trube, C.M., Bedoya, V.I., Pinto, L.A., Oppenheim, J.J., Rybak, S.M., et al., 2003 Mar. Ribonuclease is partly responsible for the HIV-1 inhibitory effect activated by HLA alloantigen recognition. *AIDS* 17 (4), 481–486.
- Samson, M., Puechal, X., Devilliers, H., Ribl, C., Cohen, P., Stern, M., et al., 2013. Long-term outcomes of 118 patients with eosinophilic granulomatosis with polyangiitis (Churg-Strauss syndrome) enrolled in two prospective trials. *J. Autoimmun.* 43, 60–69, 2013/04/18.

- Sastre, J., Dávila, I., 2018. Dupilumab: a new paradigm for the treatment of allergic diseases. *J. Investig. Allergol. Clin. Immunol.* 28 (3), 139–150. <https://doi.org/10.18176/jiaci.0254>.
- Sehmi, R., Lim, H.F., Mukherjee, M., Huang, C., Radford, K., Newbold, P., et al., 2018. Benralizumab attenuates airway eosinophilia in prednisone-dependent asthma. *J. Allergy Clin. Immunol.* 141, 1529–15232 e8.
- Shah, K., Ignacio, A., McCoy, K.D., Harris, N.L., 2020. The emerging roles of eosinophils in mucosal homeostasis. *Mucosal Immunol.* 13, 574–583.
- Shen, Z.J., Malter, J.S., 2015 Feb. Determinants of eosinophil survival and apoptotic cell death. *Apoptosis* 20 (2), 224–234. <https://doi.org/10.1007/s10495-014-1072-2>.
- Simon, H.-U., Yousefi, S., Germic, N., Arnould, I.C., Haczk, A., Karaulov, A.V., Simon, D., Rosenberg, H.F., 2020. The cellular functions of eosinophils: collegium Internationale Allergologicum (CIA) update 2020. *Int. Arch. Allergy Immunol.* 181, 11–23. <https://doi.org/10.1159/000504847>.
- Sinico, R.A., Di Toma, L., Maggiore, U., Bottero, P., Radice, A., Tosoni, C., et al., 2005. Prevalence and clinical significance of antineutrophil cytoplasmic antibodies in Churg-Strauss syndrome. *Arthritis Rheum.* 52 (9), 2926–2935. <https://doi.org/10.1002/art.21250>. Sep.
- Smith, S.G., Chen, R., Kjarsgaard, M., et al., 2016. Increased numbers of activated group 2 innate lymphoid cells in the airways of patients with severe asthma and persistent airway eosinophilia. *J. Allergy Clin. Immunol.* 137, 75–86.
- Stolarski, B., Kurowska-Stolarska, M., Kewin, P., Xu, D., Liew, F.Y., 2010. IL-33 exacerbates eosinophil-mediated airway inflammation. *J. Immunol.* 185, 3472–3480.
- Strandmark, J., Rausch, S., Hartmann, S., 2016. Eosinophils in homeostasis and their contrasting roles during inflammation and helminth infections. *Crit. Rev. Immunol.* 36 (3), 193–238. <https://doi.org/10.1615/CritRevImmunol.2016018726>.
- Straumann, A., Kristl, J., Conus, S., Vassina, E., Spichtin, H.P., Beglinger, C., et al., 2005 Aug. Cytokine expression in healthy and inflamed mucosa: probing the role of eosinophils in the digestive tract. *Inflamm. Bowel Dis.* 11 (8), 720–726.
- Straumann, A., Conus, S., Grzonka, P., Kita, H., Kephart, G., Bussmann, C., et al., 2010. Anti-interleukin-5 antibody treatment (mepolizumab) in active eosinophilic oesophagitis: a randomized, placebo-controlled, double-blind trial. *Gut* 59, 21–30.
- Tagaya, E., Tamaoki, J., 2007. Mechanisms of airway remodeling in asthma. *Allergol. Int.* 56, 331–340.
- Valent, P., Klion, A.D., Horny, H.P., et al., 2012. Contemporary consensus proposal on criteria and classification of eosinophilic disorders and related syndromes. *J. Allergy Clin. Immunol.* 130, 607–612 e609.
- Vedel-Krogh, S., 2020. The search for the “healthy” blood eosinophil count. *Eur Respir J.* 55, 2000473 <https://doi.org/10.1183/13993003.00473-2020>.
- Von Hulst, G., 2021 sept. Anti-IL5 mepolizumab minimally influences residual blood eosinophils in severe asthma. *Eur. Respir. J.* 2, 2100935.
- Wechsler, M.E., Akuthota, P., Jayne, D., Khoury, P., Klion, A., Langford, C.A., Merkel, P. A., Moosig, F., Specks, U., Cid, M.C., Luqmani, R., Brown, J., Mallett, S., Philipson, R., Yancey, S.W., Steinfeld, J., Weller, P.F., Gleich, G.J., EGPA Mepolizumab Study Team, 2017. Mepolizumab or placebo for eosinophilic granulomatosis with polyangiitis. *N. Engl. J. Med.* 376 (20), 1921–1932. <https://doi.org/10.1056/NEJMoa1702079>. May 18.
- Weller, P.F., Spencer, L.A., 2017. Functions of tissue-resident eosinophils. *Nat. Rev. Immunol.* 17, 746–760.
- Wenzel, S., Ford, L., Pearlman, D., Spector, S., Sher, L., Skobieranda, F., et al., 2013. Dupilumab in persistent asthma with elevated eosinophil levels. *N. Engl. J. Med.* 368 (26), 2455–2466.
- Wong, C.K., Hu, S., Cheung, P.F., Lam, C.W., 2010. Thymic stromal lymphopoietin induces chemotactic and pro-survival effects in eosinophils: implications in allergic inflammation. *Am. J. Respir. Cell Mol. Biol.* 43, 305–315.
- Wu, D., Molofsky, A.B., Liang, H.E., Ricardo-Gonzalez, R.R., Jouihan, H.A., Bando, J.K., Chawla, A., Locksley, R.M., 2011. Eosinophils sustain adipose alternatively activated macrophages associated with glucose homeostasis. *Science* 332, 243–247.
- Yancey, S.W., Keene, Albers, F.C., Ortega, H., Bates, S., Bleecker, E.R., Pavord, I., 2017. Biomarkers for severe eosinophilic asthma. *J. Allergy Clin. Immunol.* 140, 1509–1518.
- Yang, B.G., Seoh, J.Y., Jang, M.H., 2017. Regulatory eosinophils in inflammation and metabolic disorders. *Immune Network* 17 (No. 1), 41–47.
- Yousefi, S., Gold, J.A., Andina, N., Lee, J.J., Kelly, A.M., Kozłowski, E., et al., 2008 Sep. Catapult-like release of mitochondrial DNA by eosinophils contributes to antibacterial defense. *Nat. Med.* 14 (9), 949–953.
- Yu, C., Cantor, A.B., Yang, H., Browne, C., Wells, R.A., Fujiwara, Y., Orkin, S.H., 2002. Targeted deletion of a high-affinity GATA-binding site in the GATA-1 promoter leads to selective loss of the eosinophil lineage in vivo. *J. Exp. Med.* 195, 1387–1395.
- Yu, S., Kim, H.Y., Chang, Y.J., et al., 2014. Innate lymphoid cells and asthma. *J. Allergy Clin. Immunol.* 133, 943–950.
- Yun, Y., Kanda, A., Kobayashi, Y., Van Bui, D., Suzuki, K., Sawada, S., et al., 2020. Increased CD69 expression on activated eosinophils in eosinophilic chronic rhinosinusitis correlates with clinical findings. *Allergol. Int.* 69, 232–238.