

Jurnal Kefarmasian Indonesia

Available online at https://jkefarind.com/index.php/jki **Original Research Article**

Gambier for Diabetes: Comparison of the Antidiabetic Potency between Two Types of Extracts from *Uncaria gambir* **(W. Hunter) Roxb.**

Indah Fajarwati1, Dedy Duryadi Solihin1,2*, Tutik Wresdiyati³*

¹Department of Biology, Faculty of Mathematics and Natural Sciences, IPB University, Bogor, Indonesia ²Biotech Center, IPB University, Bogor, West Java, Indonesia

³Department of Anatomy, Physiology, and Pharmacology, School of Veterinary Medicine and Biomedical Sciences, IPB University, Bogor, Indonesia

ARTICLE INFO **ABSTRACT**

Received 29 September 2023 Revised 24 November 2023 Accepted 17 February 2024 Published online 29 February 2024

*Corresponding author. E-mail: dduryadi@yahoo.com

DOI[: https://doi.org/10.22435/jki.v14i1.6](https://doi.org/10.22435/jki.v14i1.)621

Citation: Fajarwati I, Solihin DD, Wresdiyati T.

Jurnal Kefarmasian Indonesia. 2024;14(1):51-62.

original author and source are credited.

Article history:

Gambier is a dry extract obtained from both the leaves and twigs of the *Uncaria gambir*. It is an agricultural product produced by the local community through traditionally unique methods. The main objective of this study was to evaluate the antidiabetic potency of gambier. The study used two types of gambier, namely traditional aqueous gambier (TAG) from traditional extraction and maceration ethanol gambier (MEG) from maceration extraction. The phytochemical compound, alpha-glucosidase inhibition, and antioxidant activity were analyzed. Moreover, in vivo hypoglycemia activity study was carried out. The results showed that both TAG and MEG contained flavonoids, tannin, and triterpenoids, while saponins were detected in TAG and steroids in MEG only. Both TAG and MEG also contained quercetin and catechin which were higher in the TAG compared to the MEG. The DPPH scavenging activities of the TAG and MEG, measured in IC50 values, were 15.40 ± 0.21 μ g/ml and 12.25 ± 0.04 μ g/ml, respectively. The IC50 values of the alpha-glucosidase inhibition of TAG and MEG were 35.84 ± 1.75 μg/ml and 83.14 ± 1.26 μg/ml, respectively. The best hypoglycemic activity was observed in the TAG group. The group given TAG and high glucose had lower blood glucose (127 mg/dl) compared to the group only given high glucose (178 mg/dl). These results indicated that TAG had a better potential for antidiabetic activities compared to MEG, and it can be proposed for further investigations as a potential antidiabetic agent. Gambier for diabetes: Comparison of the antidiabetic potency between two types of extracts from Uncaria gambir (W. Hunter) Roxb. **Copyright:** © **2024 Fajarwati** *et al*. This is an open-access article distributed under the terms of the **[Creative Commons](https://creativecommons.org/licenses/by/4.0/)** Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the

> *Keywords***:** Alpha-glucosidase inhibitor; Antioxidant; Diabetes; Gambier; Glucose

INTRODUCTION

Diabetes mellitus is a global disease, with the number of prevalences increasing every year. 1-4 Apart from being a health burden, diabetes also poses a significant economic burden on the healthcare systems of some countries, such as USA, China, and Germany. 5-7 Another critical point is that Indonesia ranks fifth worldwide for the highest number of

diabetes patients, with about 19 million in 2021 and estimated to increase to 28.6 million by 2045. ⁸ Managing diabetes mellitus is undoubtedly one of the health priorities worldwide. A potential solution to address this disease is to explore and evaluate various substances that have the potential to act as anti-diabetic agents.

Gambier is a dry extract derived from the leaves and twigs of *Uncaria gambir*. It belongs to the Rubiaceae family. The plant originates from Southeast Asia, particularly Indonesia and Malaysia. 9 Gambier is used to 'menginang' in some areas of Indonesia. The term 'menginang' (sometimes referred to as 'menyirih') is the process of mixing various ingredients, including gambier, wrapped using betel leaves, which is then chewed by the local community as a tradition. The local community produces gambier with a traditionally unique method. The method is similar to the extraction principle using water solvent but differs regarding traditional equipment utilization and heating process. The use of water as a solvent may be more convenient and acceptable for further application based on safety reasons. Water solvents also had an excellent ability to produce pharmacological functions from a particular plant. 10-14 It is also considered the greenest solvent in chemistry from an experimental and an industrial perspective.¹⁵

The hypothesis of the antidiabetic potency of gambier is based on previous studies. *Uncaria tomentosa*, a species native to the Amazon forest, has the same genus as *U. gambir* and could prevent diabetes mellitus. It reduces the blood glucose level, delays diabetes incidence, and protects beta cell mass loss.¹⁶ Some flavonoids were tested to inhibit the intestinal glucose transporter GLUT2. The tests revealed that quercetin could inhibit glucose absorption in the intestines via the transporter GLUT2.¹⁷ Quercetin could also inhibit the alpha-glucosidase enzyme.¹⁸ Another flavonoid, Catechins, also has some beneficial effects in managing diabetes. Catechins could improve insulin resistance, alleviate oxidative stress, regulate mitochondrial function, alleviate endoplasmic reticulum stress, produce anti-inflammatory effects, reduce blood sugar sources, and regulate intestinal function. 19-24 Thus, gambier is strongly expected to have antidiabetic potency, as it was observed to contain both catechin and quercetin.

A study reported antioxidant activity and alpha-glucosidase inhibition of gambier.²⁵ However, the study was conducted using the ethanol extract of commercial gambier from a local market in Indonesia. The extract was produced from a re-extraction of traditional gambier extract, which may cause the loss of some active compounds.

This study will be the first to evaluate the antidiabetic potency of traditional gambier extract. This study compared the antidiabetic potency of the two types of gambier extract, namely traditional aqueous gambier (TAG) made using the traditional extraction method and maceration ethanol gambier (MEG) made using the maceration extraction method. Furthermore, the in vivo study of these extracts has yet to be previously reported with regard to assessing the antidiabetic potency of gambier. Thus, this study aimed to analyze the phytochemical compounds, antioxidant activity, alpha-glucosidase inhibition, and in vivo hypoglycemic activity of two gambier extracts.

METHODS

Plant materials

Leaves and twigs of *Uncaria gambir* were harvested in September in Pesisir Selatan District, West Sumatera, Indonesia. The validation of the *U. gambir* was done by identifying the plant in the herbarium of Andalas University (UNAND), Padang, West Sumatera, Indonesia.

Animals

Six weeks old, Sprague Dawley male rats were purchased from Indoanilab, a private company in Bogor, Indonesia. All animals received standard commercial rat food and water *ad libitum*. The Animals were acclimatized for two weeks before experiments. Clinically healthy animals with a body weight of 200-230 grams were used for the study. Ethical clearance was obtained from The Veterinary Teaching Hospital, Faculty of Veterinary Medicine, Bogor Agricultural University (No. 29-2016 ACUC RSHP FKH-IPB).

Extraction

Gambier extract was made using two different methods: traditional extraction yielding traditional aqueous gambier (TAG) and maceration extraction yielding maceration ethanol gambier (MEG). In the traditional extraction, 36.5 kg of fresh leaves and twigs were boiled and then pressed using a jack press to get the liquid extract. Liquid extract was filtered and then deposited in a wooden container called *paraku* for 24 hours. In *paraku*, the extract was allowed to cool into the form of a paste. The extract was then drained and pressed again with stone ballast to allow the paste to be denser and immediately printable. The extract was then blocked and sun-dried. In the maceration extraction, 14 kg of fresh leaves and twigs were ground, producing a coarse, smaller sample. The sample was extracted using the standard procedure of maceration in ethanol 70% for three days at room temperature. The resulting filtrate was then filtered and evaporated to produce the solid extract.

Phytochemical profile

Phytochemical tests were carried out using qualitative and quantitative methods. Qualitative methods to check alkaloids, flavonoids, saponins, quinones, tannins, triterpenoids, and steroids following the standard procedure.²⁶ Quercetin was determined using High-Performance Liquid Chromatography (HPLC-UV/Vis Hitachi), according to a previous study.²⁷ The quantitative analysis of catechin was carried out by spectrophotometry at a wavelength of 279 nm according to the Indonesian National Standard.

DPPH (2,2-Diphenyl-1-picrylhydrazyl) assay

The assay of DPPH was conducted according to a modified method from the previous study.²⁸ A series of sample solutions of different concentrations were prepared at 100 μL by serial dilution method. The concentrations were 100, 50, 25, 12.5, 6.25, 3.125 μg/ml. Next, 100 μL DPPH (125 µM in ethanol) was added to each sample solution (S1). The control sample only contained 100 μL of the sample solution and 100 μL of ethanol (S0). A blank solution consisting of 100 μL ethanol was added to the DPPH of 100 μL (B1). The blank control contained ethanol only (B0). The final volume of each solution was 200 μL. These solutions were incubated at 37oC in a dark condition for 30 minutes. Absorbance was measured using an ELISA reader with a wavelength of 517 nm. The scavenging capacity (SC) was calculated as follows: $SC\% = [(B-S)/B] \times 100\%$ where B was the absorbance of black corrected (B1- B0), and S was the absorbance of the sample corrected (S1-S0). All treatments were done in triplicate.

Alpha-glucosidase inhibition assay

The inhibitory activity of the alphaglucosidase enzyme was determined based on the method in the previous study. 29 Briefly, the reagent mixture contained 50 μL of Phosphate buffer, 25 μL of pnitrophenyl α-D glucopyranoside 0.5 mM, ten μL of the test sample in DMSO with a concentration range of 7.8125 μg/ml - 500 μg/ml. 25 μL enzyme solution was added to the solution (S1). Control samples contained 10 μL of the sample solution, 25 μL of p-nitrophenyl α-D-glucopyranoside and 75 μL of phosphate buffer (S0). The blank solution contained 25 μL of enzyme solution, 25 μL of p-nitrophenyl α-Dglucopyranoside, 10 μL of DMSO, and 50 μL of phosphate buffer (B1). The blank control contained 25 μL of p-nitrophenyl α-D-glucopyranoside, 10 μL of DMSO, and 75 μL of phosphate buffer (B0). The mixed solution to be incubated was 100 μL. The reaction mixture was then incubated at 37°C for 30 minutes. The reaction was stopped by the addition of 100 μL of Na2CO3 0.2 M. Substrate enzymatic hydrolysis was monitored through the amount of p-nitrophenol released in the reaction mixture at 410 nm using an ELISA reader. All treatments were triplicated. The inhibitory activity (IA) was calculated as IA% = $[(B-S)/B] \times 100\%$, where B was the corrected blank absorbance (B1-B0), and S was the corrected sample absorbance (S1- S0).

Hypoglycemic activity test

The hypoglycemic activity of gambier was examined using an Oral Glucose Tolerant Test (OGTT) according to a modified method from the previous study.³⁰ The test was performed on five rats from each group. There were nine groups as follows: G1 (negative control group), G2 (positive control group), G3 (commercial drug group treated with Glucobay© at a dose of 4.5 mg/kg BW), G4 to G6 (TAG groups treated with TAG at doses of 100 mg/kg BW, 200 mg/kg BW, and 300 mg/kg BW, respectively), and G7 to G9 (MEG groups treated with MEG at doses of 100 mg/kg BW, 200 mg/kg BW, and 300 mg/kg BW, respectively). Rats fasted overnight before treatment. They were then given gambier extracts, except positive and negative control groups that were given distilled water and the commercial drug group that was given Glucobay. A glucose load was then orally administered via a gavage (90% glucose solution, 1 ml), except for the negative control group that was given distilled water as a placebo. The blood glucose levels were then measured every 30 minutes for two hours. Blood samples were taken from rat tails cleaned with alcohol (70%). The blood glucose was measured using an Accu-Check® Active glucometer (Roche, Germany). The area under the glucose curve (AUC) during the OGTT was determined using the trapezoidal method.

Statistical analysis

Antioxidant activity, alphaglucosidase inhibition and glucose AUC results were presented as mean ± SD. IC50 values from log regression were used to present antioxidant activity and alphaglucosidase inhibition. We used the Mann-Whitney test to compare quantitative phytochemical content, DPPH scavenging activity and alpha-glucosidase inhibition of two gambier extracts. Glucose AUC was analyzed using ANOVA followed by

Duncan's multiple range tests (p<0.05 or p<0.01).

RESULTS AND DISCUSSION

Natural products have been targeted for many years to cope with various diseases, including diabetes, for their medicinal value. This study evaluated the antidiabetic potency of two types of extract of *Uncaria gambir*. They are traditional aqueous gambier (TAG), which uses water as the solvent, and maceration ethanol gambier (MEG), which uses ethanol as the solvent. Both aqueous and ethanolic solvents were less toxic than other organic solvents. Thus, further application of the extracts could be more acceptable.³¹

The extraction yield of *Uncaria gambier* leaves and twigs is shown in Table 1. This study showed that 36.5 kg of leaves and twigs of *U. gambir* produced 3.50 kg of TAG (9.59%). Meanwhile, 14 kg of *U. gambir* produced 1.92 kg of MEG (13.71%). The higher yield of MEG compared to TAG suggests that the type of solvent could influence the yield of each extract. The MEG obtained using the ethanol solvent (containing 70% ethanol as an organic solvent and 30% water) may facilitate the extraction of both water-soluble chemicals and an organic solvent.³² Also, the organic compounds are generally non-polar, and most of them are insoluble in water as water is a polar solvent. Most carbohydrates and pigments as organic compounds elute with ethanol.³³ It is possible that these factors contributed to the higher yield of MEG compared to TAG.

In this study, both TAG and MEG were evaluated for their phytochemical compounds as shown in Table 1. It can be used for estimating a plant's pharmacological function. A qualitative phytochemical analysis was conducted on both TAG and MEG, revealing the presence of flavonoids, tannins, and triterpenoids in both. However, only TAG contained saponins, and MEG contained steroids. In the quantitative analysis, TAG had higher levels of both quercetin and catechin compared to MEG. Specifically, TAG contained 2.65 mg/g of quercetin, while MEG contained 1.69 mg/g. Additionally, TAG had 80.56%wt of catechin, while MEG had 71.73%wt. These findings suggest that TAG and MEG differ qualitatively and quantitatively in terms of phytochemical compounds. The extraction temperature, extraction time, and type of solvents may influence these variations, which could affect the antidiabetic potency of TAG and MEG. It is important to determine the amount of catechin present in the gambier, as it is used as a marker in trade and establishes the quality of the gambier.³⁴ The quantification of quercetin may also provide valuable information for evaluating the antidiabetic potency of gambier, as previous studies have reported its effectiveness.35,36 However, the contribution of other compounds should also be considered.

Statistically significant differences are indicated by a, b $(p \leq$ 0.05).

Abbreviations: Traditional Aqueous Gambier (TAG); Maceration Ethanol Gambier (MEG).

This study evaluated the antioxidant activity of two types of gambier extract using the DPPH method. The results of the DPPH are expressed in IC50 values and presented in Table 2. The DPPH IC50 values of the TAG and MEG were 15.40 μg/ml and 12.25 μg/ml, respectively. The IC50 value of MEG was lower than that of TAG. It indicated that TAG has slightly higher antioxidant activity than the MEG. However, both of them were categorized as very strong antioxidants. Antioxidant activity is classified as very strong (IC50 <50 μg/ml), strong (IC50 50-100 μg/ml), moderate (IC50 100-150 μg/ml), and weak (IC50 150-200 μ g/ml. The information on the antioxidant activity of gambier is essential, with increasing evidence showing that oxidative stress contributes to the development and complications of diabetes.37,38 Thus, the antioxidant is expected to inhibit these two aspects of diabetes. Additionally, several studies reported that various plants that have antioxidant properties showed antidiabetic activities. 39-41

Two types of gambier extracts, TAG and MEG, were studied for the inhibitory activity of the alpha-glucosidase enzyme (Table 2). It was surprising to find that TAG showed more significant inhibition of the enzyme, with roughly twice the potency of MEG. The IC50 values for TAG and MEG were 35.84 μg/ml and 83.14 μg/ml, respectively. Recent research has identified phytochemical compounds such as quercetin and catechin with alphaglucosidase inhibition activity.42-45 These compounds were found to be more abundant in TAG than MEG. Additionally, saponins were also reported to have alphaglucosidase inhibition activities, but they were not detected in MEG.46,47

The alpha-glucosidase inhibition of gambier was found to be dose-dependent. It was elevated as the concentration of gambier extracts increased. However, when the gambier extract concentration gradually increased, the inhibition activity slowed. It could be a consequence of the enzyme molecules being occupied, becoming a limiting factor. The inhibition became plateau regardless of the increased extract concentration, suggesting that all enzyme molecules are occupied, and gambier extracts as an inhibitor were in excess. 48

Table 2. Alpha-glucosidase inhibition and antioxidant activity of gambier extracts

Notes: *Percentage of DPPH scavenging or alpha-glucosidase inhibition activity. #Concentrations of extract that scavenged or inhibited 50% system, equipped with R-value from log regression. Statistically significant differences between the means of IC50 are indicated by a, b ($p < 0.05$).

Abbreviations: Traditional Aqueous Gambier (TAG); Maceration Ethanol Gambier (MEG).

The hypoglycemic activity of gambier extracts was analyzed using the Oral Glucose Tolerant Test (OGTT). OGTT is widely utilized to investigate and evaluate the hypoglycemic potential of various plants.49-53 The test could be the initial approach in considering the properness of a plant for further study and development as an antidiabetic agent. The anti-diabetic properties of plants, especially alphaglucosidase inhibitors, are expected to reduce glucose absorption in the intestine. 54-57 As a result, during the OGTT test, the blood glucose levels of rats given the antidiabetic candidate were lower than the control group.

The results of the OGTT test are displayed in Figure 1 and Table 3. Blood glucose levels rose above the normal range 30 minutes after administering glucose, particularly in the G2 group. The G2 group had blood glucose levels that nearly reached 180 mg/dl. However, all treated gambier groups (G4 to G9) showed lower blood glucose levels compared to G2. Table 3 shows the AUC glucose during the OGTT, which confirms that the gambier groups (G4 to G9) are significantly different from the untreated group (G2). The AUC glucose demonstrated that TAG groups (G4 to G6) at doses of 100 mg/kg BW, 200 mg/kg BW, and 300 mg/kg BW

had lower AUC glucose, which correlated with blood glucose levels over two hours compared to MEG groups (G7 to G9) using the same extract doses. Among the TAG groups, 200 mg/kg BW (G5) produced the best hypoglycemic activity. This test proved that TAG had better hypoglycemic activity than MEG.

The exact mechanism of TAG lowers blood glucose levels still needs to be fully understood. The hypoglycemic effects of gambier can be attributed to its various phytochemical components. Some of these components, such as catechin, quercetin, and saponin, are found in gambier. Among these components, catechin and quercetin are more prevalent in TAG, while saponin is only present in TAG, not MEG. According to a study, saponins have been known to enhance insulin sensitivity both in vitro and in vivo. 23,58-60 While MEG has been found to have hypoglycemic properties, it has some limitations. For instance, in a study involving rats, those given MEG had higher blood glucose levels than those given TAG. Additionally, the MEG could not return blood glucose levels to normal range at the 120-minute mark. It may also be due to the presence of antagonistic substances, thereby inhibiting gambier's hypoglycemic activity.

Figure 1. Hypoglycemic effect of gambier extracts. Values are expressed as means of blood glucose levels (n = 5 per group). G1 and G2: negative control and positive control group, respectively; G3: glucobay followed by 90% glucose 1 ml; G4, G5, G6: TAG at 100, 200, 300 mg/kg, respectively, followed by glucose 90% 1 ml; MEG at 100, 200, 300 mg/kg, respectively, followed by 90% glucose 1 ml. **Abbreviations:** Traditional Aqueous Gambier (TAG); Maceration Ethanol Gambier (MEG).

Table 3. The area under the 2hr glucose curve (AUC) for each group during OGTT

Groups	AUC-Glucose
G1-Negative control	178.70 ± 7.398 ^a
G ₂ - Positive control	276.85 ± 26.484 ^d
G3-Glucobay	237.45 ± 15.923 bc
$G4 - TAG$ 100	239.85 ± 12.260 bc
$G5 - TAG$ 200	224.35 ± 15.417 ^b
$G6 - TAG300$	242.40 ± 6.035 bc
$G7 - MEG$ 100	253.30 ± 6.542 cd
G8-MEG 200	259.20 ± 13.654 ^{cd}
G9 - MEG 300	263.30 ± 11.226 cd
۰.	$\ddot{}$

The same letter in the same column means not significantly different (p *<* 0.05).

Abbreviations: Traditional Aqueous Gambier (TAG); Maceration Ethanol Gambier (MEG).

Another previous study had studied the antidiabetic potential of gambier through DPPH scavenging activity and alpha-glucosidase inhibition evaluation tests. However, the study was conducted using re-extracted commercial gambier from a local market in Indonesia, which was different to aqueous and ethanol extractions from fresh leaves and twigs of *U. gambir*, refers to gambier.²⁵ The reextraction process is a method reported to

increase catechin content in gambier, as catechin is a primary active compound. In comparison, this study revealed that gambier in the form of crude extracts (TAG and MEG) had IC50 values of both DPPH scavenging activity and alpha-glucosidase inhibition, which were similar to those of the re-extraction process. Further study is required to determine whether the phytochemical contribution of gambier is individual or synergistic. So, this extract or more purified derivatives could be applied as an antidiabetic agent.

CONCLUSION

Two types of gambier extracts (traditional aqueous gambier and maceration ethanol gambier) were examined in this study. Both extracts showed intense DPPH scavenging activity and inhibited the alpha-glucosidase enzyme. They also reduced rat blood glucose levels. However, traditional aqueous gambier showed better antidiabetic potency. Further studies on the animal model of diabetes are being carried

out to precisely understand the antidiabetic mechanism of TAG and determine its maximum potency in treating diabetes.

Conflict of Interest

The authors declare no conflict of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them. All authors contributed equally to this work.

Acknowledgments

The authors would like to thank the Ministry of Research, Technology, and Higher Education of the Republic of Indonesia for supporting this study through "Penelitian Pendidikan Magister Menuju Doktor Sarjana Unggul (PMDSU)" with under No. 330/SP2H/LT/DRPM/IX/2016.

REFERENCES

- 1. Safiri S, Karamzad N, Kaufman JS, Bell AW, Nejadghaderi SA, Sullman MJM, et al. Prevalence, Deaths and Disability-Adjusted-Life-Years (DALYs) Due to Type 2 Diabetes and Its Attributable Risk Factors in 204 Countries and Territories, 1990-2019: Results From the Global Burden of Disease Study 2019. Front Endocrinol (Lausanne). 2022;13(February):1–14.
- 2. Gregory GA, Robinson TIG, Linklater SE, Wang F, Colagiuri S, de Beaufort C, et al. Global incidence, prevalence, and mortality of type 1 diabetes in 2021 with projection to 2040: a modelling study. Lancet Diabetes Endocrinol. 2022;10(10):741–60.
- 3. Akhtar S, Nasir JA, Ali A, Asghar M, Majeed R, Sarwar A. Prevalence of type-2 diabetes and prediabetes in Malaysia: A systematic review and meta-analysis. PLoS One [Internet]. 2022;17(1 January):1–14. Available

from:

http://dx.doi.org/10.1371/journal.po ne.0263139

- 4. Liu J, Ren Z, Qiang H, Wu J, Shen M, Zhang L, et al. Trends in the incidence of diabetes mellitus : results from the Global Burden of Disease Study 2017 and implications for diabetes mellitus prevention. BMC Public Health. 2020;(20):1–12.
- 5. Standl E, Khunti K, Hansen TB, Schnell O. The global epidemics of diabetes in the 21st century: Current situation and perspectives. Eur J Prev Cardiol. 2019;26(2_suppl):7–14.
- 6. Sugandh F, Chandio M, Raveena F, Kumar L, Karishma F, Khuwaja S, et al. Advances in the Management of Diabetes Mellitus: A Focus on Personalized Medicine. Cureus. 2023;15(8).
- 7. Tomic D, Shaw JE, Magliano DJ. The burden and risks of emerging complications of diabetes mellitus. Nat Rev Endocrinol. 2022;18(9):525–39.
- 8. IDF. IDF Diabetes Atlas. 10th editi. Boyko EJ, Karuranga DJMS, Piemonte L, Saeedi PRP, Boyko HSJ, Karuranga DJMS, et al., editors. Diabetes Research and Clinical Practice. International Diabetes Federation; 2021. 37 p.
- 9. Munggari IP, Kurnia D, Deawati Y, Julaeha E. Current Research of Phytochemical, Medicinal and Non-Medicinal Uses of Uncaria gambir Roxb.: A Review. Molecules. 2022;27(19).
- 10. Nguyen NH, Pham QT, Luong TNH, Le HK, Vo VG. Potential antidiabetic activity of extracts and isolated compound from adenosma bracteosum (Bonati). Biomolecules. 2020;10(2).
- 11. Abdel-Aal RA, Abdel-Rahman MS, Al Bayoumi S, Ali LA. Effect of stevia aqueous extract on the antidiabetic activity of saxagliptin in diabetic rats. J Ethnopharmacol. 2021;265(January):1– 6.
- 12. Zofou D, Matumamboh EE, Shu G, Tsague MFP, Teugwa MC, Sofeu FDD, et al. In vivo assessment of antidiabetic

activity and safety of polyherbal teas from selected Cameroonian medicinal plants: Persea americana, Ageratum conyzoides and Mangifera indica. Investig Med Chem Pharmacol. 2023;6(2):1–9.

- 13. Rahhal BM, Jaradat N, Hawash M, Qadi M, Issa L, Yahya A, et al. Phytochemical Screening, Antioxidative, Antiobesity, Antidiabetic and Antimicrobial Investigations of Artemisia scoparia Grown in Palestine. Processes. 2022;10(10).
- 14. Saxena M, Prabhu SV, Mohseen M, Pal AK, Alarifi S, Gautam N, et al. Antidiabetic Effect of Tamarindus indica and Momordica charantia and Downregulation of TET-1 Gene Expression by Saroglitazar in Glucose Feed Adipocytes and Their Involvement in the Type 2 Diabetes-Associated Inflammation in Vitro. Biomed Res Int. 2022;2022:1–10.
- 15. Lajoie L, Fabiano-Tixier AS, Chemat F. Water as Green Solvent: Methods of Solubilisation and Extraction of Natural Products—Past, Present and Future Solutions. Pharmaceuticals. 2022;15(12).
- 16. Domingues A, Sartori A, Golim MA, Valente LMM, Da Rosa LC, Ishikawa LLW, et al. Prevention of experimental diabetes by Uncaria tomentosa extract: Th2 polarization, regulatory T cell preservation or both? J Ethnopharmacol [Internet]. 2011;137(1):635–42. Available from: http://dx.doi.org/10.1016/j.jep.2011. 06.021
- 17. Kwon O, Eck P, Chen S, Corpe CP, Lee J, Kruhlak M, et al. Inhibition of the intestinal glucose transporter GLUT2 by flavonoids. FASEB J. 2007;21(2):366–77.
- 18. Barber E, Houghton MJ, Williamson G. Flavonoids as human intestinal αglucosidase inhibitors. Foods. 2021;10(8).
- 19. Wen L, Wu D, Tan X, Zhong M, Xing J, Li W, et al. The Role of Catechins in Regulating Diabetes: An Update

Review. Nutrients. 2022;14(4681):1–19.

- 20. Chen B, Zhang W, Lin C, Zhang L. A Comprehensive Review on Beneficial Effects of Catechins on Secondary Mitochondrial Diseases. Int J Mol Sci. 2022;23(19).
- 21. Kim JM, Heo HJ. The roles of catechins in regulation of systemic inflammation. Food Sci Biotechnol [Internet]. 2022;31(8):957–70. Available from: https://doi.org/10.1007/s10068-022- 01069-0
- 22. Basu T, Selman A, Reddy AP, Reddy PH. Current Status of Obesity: Protective Role of Catechins. Antioxidants. 2023;12(2):1–21.
- 23. Shi Y, Liu Z, Gai L, Gao Y, He Y, Liu C, et al. The preventive effect of total saponins from Panax japonicus on inflammation and insulin resistance in adipose tissue of mice induced by a high-fat diet. J Funct Foods [Internet]. 2021;78:104369. Available from: https://doi.org/10.1016/j.jff.2021.104 369
- 24. Monika P, Chandraprabha MN, Murthy KNC. Catechin, epicatechin, curcumin, garlic, pomegranate peel and neem extracts of Indian origin showed enhanced anti-inflammatory potential in human primary acute and chronic wound derived fibroblasts by decreasing TGF-β and TNF-α expression. BMC Complement Med Ther [Internet]. 2023;23(1):1–16. Available from: https://doi.org/10.1186/s12906-023- 03993-y
- 25. Apea-Bah F, Hanafi M, Dewi RT, Fajriah S, Darwaman A, Artanti N, et al. Assessment of the DPPH and glucosidase inhibitory potential of gambier and qualitative identification of major bioactive compound. J Med Plants Res. 2009;3(10):736–57.
- 26. Harborne JB. Methods of Plant Analysis. Third. Phytochemical Methods. International Thomson Publishing; 1973. 1–32 p.
- 27. Hertog MGL, Hollman PCH, Venema DP. Optimization of a Quantitative

HPLC Determination of Potentially Anticarcinogenic Flavonoids in Vegetables and Fruits. J Agric Food Chem. 1992;40(9):1591–8.

- 28. Salazar-Aranda R, P´erez-L´opez LA, L´opez-Arroyo J, Alan´ıs-Garza BA, Torres NW de. Antimicrobial and antioxidant activities of plants from northeast of Mexico. Evidence-based Complement Altern Med. 2011;2011.
- 29. Sancheti S, Sancheti S, Seo SY. Chaenomeles sinensis: A potent α-and β-glucosidase inhibitor. Am J Pharmacol Toxicol. 2009;4(1):8–11.
- 30. Wresdiyati T, Sa'diah S, Winarto A, Febriyani V. Alpha-Glucosidase Inhibition and Hypoglycemic Activities of Sweitenia mahagoni Seed Extract. HAYATI J Biosci [Internet]. 2015;22(2):73–8. Available from: http://dx.doi.org/10.4308/hjb.22.2.73
- 31. Febrinda AE, Yuliana ND, Ridwan E, Wresdiyati T, Astawan M. Hyperglycemic control and diabetes complication preventive activities of Bawang Dayak (Eleutherine palmifolia L. Merr.) bulbs extracts in alloxandiabetic rats. Int Food Res J. 2014;21(4):1405–11.
- 32. Do QD, Angkawijaya AE, Tran-Nguyen PL, Huynh LH, Soetaredjo FE, Ismadji S, et al. Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of Limnophila aromatica. J Food Drug Anal [Internet]. 2014;22(3):296–302. Available from: http://dx.doi.org/10.1016/j.jfda.2013. 11.001
- 33. McEvoy FA, Lynn WS. Chloroplast Membrane Proteins. J Biol Chem. 1973;248(13):4568–73.
- 34. Arumsari AG. Analysis of Catechins in Gambir West Sumatra. ARRUS J Eng Technol. 2021;1(2):47–54.
- 35. Azeem M, Hanif M, Mahmood K, Ameer N, Chughtai FRS, Abid U. An insight into anticancer, antioxidant, antimicrobial, antidiabetic and antiinflammatory effects of quercetin: a review. Polym Bull [Internet]. 2023;80(1):241–62. Available from:

https://doi.org/10.1007/s00289-022- 04091-8

- 36. Hasan AA, Tatarskiy V, Kalinina E. Synthetic Pathways and the Therapeutic Potential of Quercetin and Curcumin. Int J Mol Sci. 2022;23(22).
- 37. Masenga SK, Kabwe LS, Chakulya M, Kirabo A. Mechanisms of Oxidative Stress in Metabolic Syndrome. Int J Mol Sci. 2023;24(9).
- 38. Singh A, Kukreti R, Saso L, Kukreti S. Mechanistic Insight into Oxidative Stress-Triggered Signaling Pathways and Type 2 Diabetes. Molecules. 2022;27(3).
- 39. Purwaningsih I, Maksum IP, Sumiarsa D, Sriwidodo S. A Review of Fibraurea tinctoria and Its Component, Berberine, as an Antidiabetic and Antioxidant. Molecules. 2023;28(3):1– 38.
- 40. Daniel AI, Gara TY, Ibrahim YO, Muhammad FM, Salisu FE, Tsado R, et al. In vivo antidiabetic and antioxidant activities of chloroform fraction of Nelsonia canescens Leaf in Alloxaninduced Diabetic Rats. Pharmacol Res Mod Chinese Med [Internet]. 2022;3(April):100106. Available from: https://doi.org/10.1016/j.prmcm.202 2.100106
- 41. Wulandari L, Nugraha AS, Himmah UA. Penentuan Aktivitas Antioksidan dan Antidiabetes Ekstrak Daun Matoa (Pometia pinnata J.R. Forst. & G. Forst.) secara In Vitro. J Kefarmasian Indones. 2021;11(2):132–41.
- 42. Aghababaei F, Hadidi M. Recent Advances in Potential Health Benefits of Quercetin. Pharmaceuticals. 2023;16(7):1–31.
- 43. Shen H, Wang J, Ao J, Cai Y, Xi M, Hou Y, et al. Inhibitory kinetics and mechanism of active compounds in green walnut husk against αglucosidase: Spectroscopy and molecular docking analyses. Lwt [Internet]. 2022;172(July):114179. Available from: https://doi.org/10.1016/j.lwt.2022.11 4179
- 44. Orita T, Chogahara S, Okuda M, Sakao

K, Hou D-X. Extraction Efficiency and Alpha-Glucosidase Inhibitory Infusion Methods. Foods. 2023;(12):1–16.

- 45. Choudhary DK, Chaturvedi N, Singh A, Mishra A. Characterization, inhibitory activity and mechanism of polyphenols from faba bean (gallicacid and catechin) on α-glucosidase: insights from molecular docking and simulation study. Prep Biochem Biotechnol [Internet]. 2020;50(2):123– 32. Available from: https://doi.org/10.1080/10826068.20 19.1679171
- 46. Van Cong P, Tuan Anh H Le, Vinh LB, Han YK, Trung NQ, Minh BQ, et al. Alpha-Glucosidase Inhibitory Activity of Saponins Isolated from Vernonia gratiosa Hance. J Microbiol Biotechnol. 2023;33(6):797–805.
- 47. Nabil M, Ghaly NS, Kassem IAA, Grace MH, Melek FR. Two triterpenoid saponins with αglucosidase inhibitory activity from Harpullia pendula seed extract. Pharmacogn J. 2019;11(6):1386–90.
- 48. Watcharachaisoponsiri T, Sornchan P, Charoenkiatkul S, Suttisansanee U. The α-glucosidase and α-amylase inhibitory activity from different chili pepper extracts. Int Food Res J. 2016;23(4):1439–45.
- 49. Sai K, Chhetri SBB, Devkota SR, Khatri D. Evaluation of the Hypoglycemic Potential of Leaves Extract of Spondias pinnata (L.f.) Kurz. From Nepal. Sci World J. 2021;2021(Figure 1).
- 50. Gebremeskel L, Tuem KB, Teklu T. Evaluation of antidiabetic effect of ethanolic leaves extract of becium grandiflorum lam. (lamiaceae) in streptozotocin-induced diabetic mice. Diabetes, Metab Syndr Obes. 2020;13:1481–9.
- 51. Noora RR, Deb P, Jahan R, Mahamud R Al, Jannat K, Rahmatullah M. Oral Glucose Tolerance Test with Methanolic Extract of Homalomena aromatica (Araceae) Whole Plant. EC Pharmacol Toxicol. 2020;11:26–30.
- 52. Alyahya ARAI, Asad M, Alhussaini MS, Abdelsalam KEA, Alenezi EA. The

antidiabetic effect of methanolic extract of Holarrhena pubescens seeds is mediated through multiple mechanisms of action. Saudi Pharm J [Internet]. 2023;31(6):824–33. Available from:

https://doi.org/10.1016/j.jsps.2023.04 .009

- 53. Cunha J da SM da, Alfredo TM, Santos JM dos, Junior VVA, Rabelo LA, Lima ES, et al. Antioxidant, antihyperglycemic, and antidiabetic activity of Apis mellifera bee tea. PLoS One. 2018;13(6):1–17.
- 54. Dirir AM, Daou M, Yousef AF, Yousef LF. A review of alpha-glucosidase inhibitors from plants as potential candidates for the treatment of type-2 diabetes. Phytochem Rev [Internet]. 2021;21(4):1049–79. Available from: https://doi.org/10.1007/s11101-021- 09773-1
- 55. Kashtoh H, Baek KH. Recent Updates on Phytoconstituent Alpha-Glucosidase Inhibitors: An Approach towards the Treatment of Type Two Diabetes. Plants. 2022;11(20).
- 56. Alssema M, Ruijgrok C, Blaak EE, Egli L, Dussort P, Vinoy S, et al. Effects of alpha-glucosidase-inhibiting drugs on acute postprandial glucose and insulin responses: a systematic review and meta-analysis. Nutr Diabetes [Internet]. 2021;11(1). Available from: http://dx.doi.org/10.1038/s41387- 021-00152-5
- 57. Assefa ST, Yang EY, Chae SY, Song M, Lee J, Cho MC, et al. Alpha glucosidase inhibitory activities of plants with focus on common vegetables. Plants. 2020;9(1).
- 58. Cui J, Duan J, Chu J, Guo C, Xi M, Li Y, et al. Chikusetsu saponin IVa protects pancreatic β cell against intermittent high glucose-induced injury by activating Wnt/β-catenin/TCF7L2 pathway. Aging (Albany NY). 2020;12(2):1591–609.
- 59. Zhou Y, Xu B. New insights into antidiabetes effects and molecular mechanisms of dietary saponins. Crit Rev Food Sci Nutr. 2022;1–6.

60. Jiang S, Xu L, Xu Y, Guo Y, Wei L, Li X, et al. Antidiabetic effect of Momordica charantia saponins in rats induced by high-fat diet combined with STZ.
Electron J Biotechnol [Internet]. Biotechnol [Internet]. 2020;43:41–7. Available from: https://doi.org/10.1016/j.ejbt.2019.12 .001