

Single Factor Effect of Natural Deep Eutectic Solvent Citric Acid-Glucose Based Microwave-Assisted Extraction on Total Polyphenols Content from *Mitragyna speciosa* Korth. Havil Leaves

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History

- Submission Date: 12-06-2021;
- Review completed: 05-07-2021;
- Accepted Date: 09-07-2021.

DOI : 10.5530/pj.2021.13.143

Article Available online

<http://www.phcogj.com/v13/i5>

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ABSTRACT

Applying green chemistry principles in the exploration of an active secondary metabolite from natural products has continued to experience a significant increase in the last decade, mainly natural deep eutectic solvent (NADES) usage. *Mitragyna speciosa* (Korth.) Havil (*Rubiaceae* family) is a native of East Kalimantan, which is traditionally used as an herb to treat headaches and inflammation by drinking boiled water a few leaves. On the other hand, this plant has an addictive effect. This study aims to determine the single factor effect of NADES citric acid–glucose-based microwave-assisted extraction (MAE) to obtain the yield of total polyphenols content (TPC) from *M. speciosa* leaves. Dried powder of *M. speciosa* leaves was extracted using NADES citric acid-glucose based MAE method with some different conditions such as NADES (citric acid-glucose) ratio (4:1, 5:1, 6:1 g/g), solid-liquid ratio (10:1, 15:1, 20:1 mL/g), microwave power (90, 270, 450 Watts), and extraction time (15, 20, 25 min). The determination of TPC was performed using Folin-Ciocalteu reagent and gallic acid as a standard with different concentration levels. The absorbance measured using spectrophotometer UV-VIS at 746 nm to obtain a regression formula of $Y = 0.0022X - 0.00095$, where $R^2 = 0.9977$. Based on the result, the best of single factor condition effect was obtained the TPC value including NADES ratio of 248.69 mg GAE/g sample (5:1 g/g citric acid-glucose), a solid-liquid ratio of 146.93 mg GAE/g sample (15:1 mL/g solvent-sample), microwave power of 192.20 mg GAE/g sample (270 Watts), and extraction time of 358.59 mg GAE/g sample (15 min). The single factor effect of NADES citric acid-glucose-based MAE shows a difference in TPC value based on various conditions of this method.

Key words: Citric acid-glucose, *Mitragyna speciosa* (Korth.) Havil, Microwave-assisted extraction, Natural deep eutectic solvent, Total polyphenols content.

INTRODUCTION

In the last decade, natural deep eutectic solvents (NADES) have emerged as potential alternative solvents to replace conventional organic solvents, toxic, volatile, and flammable. NADES has been described as a product of hydrogen bonds of two different types of compounds. The appropriate concentration ratio can cause an intense depression at the melting point compared with the one of a single component state.¹⁻⁴ NADES composition is derived from natural secondary metabolites such as amino acids, organic acids, amines, sugar alcohols, and sugars. It can also be combined with water with a certain number of molar ratios. NADES is formed based on the number and position of hydrogen bonds consisting of recipients of hydrogen bonds (HBA) and hydrogen bond donors (HBD). Both positions are very influential on the stability of NADES that are formed. In addition, the addition of water can affect properties such as biocompatibility, viscosity, conductivity, toxicity, and stability of the solvent.^{1-3,5-7}

A combination of citric acid and glucose is one type of NADES composition and successfully utilized as a green solvent to extract the target secondary metabolites from natural products, mainly from plants.^{8,9} Some studies have reported applying this solvent type, such as extraction of total polyphenols

and caffeine from coffee beans,¹⁰ extractions and determination of phenolics in *Cajanus cajan* leaves,¹¹ and extraction and separation of main bioactive flavonoids from *Radix Scutellariae*.⁸ The use of NADES combined with non-conventional extraction methods has proven to be more effective than using conventional solvents because of its non-flammable, non-volatile, non-toxic, biodegradability, environmentally friendly, and edible.^{2,3,12}

Mitragyna speciosa Korth. Havil (*Rubiaceae* family) is a native plant in Southeast Asia, mainly in Indonesia, Malaysia, Thailand, and Vietnam.¹³ In East Kalimantan, the local community uses this plant as an herb to treat headaches and inflammation by drinking boiled water a few leaves. Some studies of *M. speciosa* extract on rats are known to have an active anti-inflammatory of 200 mg/kg. This plant contains several types of glycosides, rich in polyphenols and terpenoids.^{14,15,16} Besides, this plant has activities that can affect the central nervous system due to contains secondary metabolites that have addictive effects, such as mitragynine, 7-hydroxy-mitragynine, painantein, speciesiin, and speciosiliatin.^{17,18} Application of NADES bases microwave-assisted extraction has not been reported.

In the present study, a single factor effect of NADES based microwave-assisted extraction on total polyphenols content extraction from *M. speciosa*

Cite this article: Herman, Ibrahim A, Rahayu BP, Arifuddin M, Nur Y, Prabowo WC, et al. Single Factor Effect of Natural Deep Eutectic Solvent Citric Acid-Glucose Based Microwave-Assisted Extraction on Total Polyphenols Content from *Mitragyna speciosa* Korth. Havil Leaves. *Pharmacogn J.* 2021;13(5): 1109-1115.

was performed using different combination factors to obtain the best extraction condition. This study aims to determine the single factor effect of NADES citric acid-glucose-based microwave-assisted extraction to obtain the yield of total polyphenols content from this plant.

MATERIALS AND METHODS

Materials and equipment

M. speciosa leaves were collected from Melak, West Kutai, East Kalimantan, Indonesia. The voucher specimen was identified at the Laboratory of Dendrology Faculty of Forestry, Universitas Mulawarman, Samarinda, East Kalimantan, Indonesia. Citric acid and sucrose were purchased from CV. Chlorogreen Bandung, West Java, Indonesia. Folin-Ciocalteu reagent, gallic acid standard, and sodium carbonate were purchased from Sigma Aldrich USA (via PT. Elo Karsa Utama, Indonesia). Meanwhile, the equipment was used, including Microwave domestic 900 Watts (Modena, USA), spectrophotometer UV-Vis, micropipette, and other glassware.

EXTRACTION PROCESS

NADES Preparation

In this study, a combination of citric acid and glucose as NADES (with three different ratios of 4:1, 5:1, and 6:1 g/g, respectively) were used according to the previous studies,^{10,19,20} with slight modification. Briefly, citric acid and glucose were weighed based on each different ratio. Both materials were melted at a specific temperature using a magnetic stirrer. After melting, aqua demineralization was added and homogenized. Subsequently, the obtained NADES solution was cooled at room temperature and filtered to obtain a homogeneous solution.

NADES Based Microwave-Assisted Extraction

A NADES citric acid-glucose-based microwave-assisted extraction was applied to obtain total polyphenols content (TPC) value from *M. speciosa* leaf using different condition factors based on some literature.^{9,10,19-24} Briefly, dried powder of *M. speciosa* leaf (5 g) was extracted using NADES citric acid-glucose base microwave-assisted extraction method using different conditions (including NADES ratio, solid-liquid ratio, microwave power, and extraction time) as can be seen in Table 1. Then, the extract solution and residue were separated using the Buchner funnel. The obtained extract solution was stored at room temperature and until ready to analyze.

Determination of Total Polyphenols Content

The total polyphenols content (TPC) value was determined using Folin-Ciocalteu reagent according to some literature.^{25,26,27,28,29,30,31} The absorbance was measured using spectrophotometer UV-Vis at 746 nm with a bit of adjustment. Briefly, the standard and sample solution (1 mL) were mixed with 5 mL of distilled water and 0.5 mL Folin-ciocalteu reagents, then homogenized for 5 min. Subsequently, a 2 mL NaCO₃ solution was added and homogenized, and incubated for 30 min until ready to analyze. A standard solution of gallic acid with various concentrations from 12.5 to 200 µg/mL was used to obtain the

Table 1: Design experimental of single factor condition of NADES based microwave-assisted extraction on TPC value from *M. speciosa* leaves.

| Factor variable | Unit | Range and level | | |
|----------------------|---------|-----------------|--------|------|
| | | Low | Medium | High |
| NADES Ratio | g/g | 4 | 5 | 6 |
| Solid – liquid ratio | mL/g | 10 | 15 | 20 |
| Microwave Power | Watts | 90 | 270 | 450 |
| Extraction Time | Minutes | 10 | 15 | 20 |

linear regression formula of $Y = 0.0022X - 0.00095$, where $R^2 = 0.9977$. The TPC value of extract samples was determined using this formula.

RESULTS AND DISCUSSION

Single Factor Experiment on Process

In this study, an extraction method-based microwave-assisted extraction (MAE) was developed using a combination of citric acid and glucose as natural deep eutectic solvent (NADES) to extract the total polyphenol content from *M. speciosa* leaves. Based on studies that have been reported previously,^{8,10} some extraction condition factors (Table 1) that affect the ability to separate the target of secondary metabolite contained in the sample matrix. The extraction time and the microwave power can be adjusted to the modified domestic microwave device by adding a condenser to prevent the solvent from evaporating and reducing the pressure in the microwave during the extraction process. At the same time, the NADES ratio and the solid-liquid ratio were obtained from the combination of the two types of NADES and the comparison between samples and solvents. The use of NADES as an alternative solvent to replace conventional organic solvents because of its non-toxic nature, environmentally friendly, and inexpensive.^{3,32} The combination of the MAE method with NADES is very suitable for extracting polyphenol compounds in natural product plants.

Single Factor Effect of Natural Deep Eutectic Solvent-based Microwave-Assisted Extraction on Total Polyphenols Content

Effect of citric acid-glucose combination

In the present study, citric acid-glucose with the ratio of 6:1, 5:1, and 4:1 g/g were selected according to previous research,¹⁰ where both NADES components act as hydrogen bond acceptor (HBA) and hydrogen bond donor (HBD), respectively.^{33,34,35} The distilled water was added to accelerate NADES preparation and reduce viscosity. Meanwhile, other condition factors, such as solid-liquid ratio, microwave power, and extraction time, are carried out in constant form (10:1 mL/g, 450 watts, and 10 min, respectively).

Table 2 shows the measurement results of total polyphenol levels from each extract obtained in the range of 200 to 250 mg GAE/g sample. The three variations of the NADES concentration show that the 5:1 NADES ratio has a higher TPC value than the other ratios (Figure 1). It shows that the 5:1 g/g NADES ratio was the most optimum ratio, and these results were in accordance with studies that have been previously reported by Ahmad *et al.*, 2018¹⁰ and Wei *et al.*, 2015.¹¹

Effect of Solid-Liquid Ratio

The solid-liquid ratio is the one-factor ratio between the number of solid samples (dried simplicia) and the solvent. This factor has an essential role during the extraction process and is related to the contact area between the sample (solid) and solvent (liquid) that affects extraction efficiency. The contact area can reach optimum conditions when the liquid phase is saturated to solid. The use of large quantities of solvents can cause improvidence and much waste, while the use of small amounts results in an incomplete extraction process.^{36,37}

In this study, the ratio of 10:1, 15:1, and 20:1 mL/g with other factors in a constant state such as NADES ratio of 5:1 g/g, extraction time of 10 min, and microwave power of 450 Watts. As can be seen in Table 2 shows that the TPC value varies based on the difference in solid-liquid ratio. Optimum conditions were obtained at a solid-liquid ratio of 15:1 mL/g (Figure 2).

Table 2: Effect of Citric acid – glucose ratio on TPC value.

| Citric Acid-Glucose ratio (g/g) | Absorbance | TPC (mgGAE/g) | Average TPC (mgGAE/g) | Deviation Standard |
|---------------------------------|------------|---------------|-----------------------|--------------------|
| 4:1 | 0.075 | 228.08 | 232.07 | 4.57 |
| | 0.076 | 231.08 | | |
| | 0.078 | 237.06 | | |
| 5:1 | 0.082 | 261.30 | 248.69 | 10.92 |
| | 0.076 | 242.39 | | |
| | 0.076 | 242.39 | | |
| 6:1 | 0.069 | 199.42 | 203.23 | 2.29 |
| | 0.071 | 205.13 | | |
| | 0.071 | 205.13 | | |

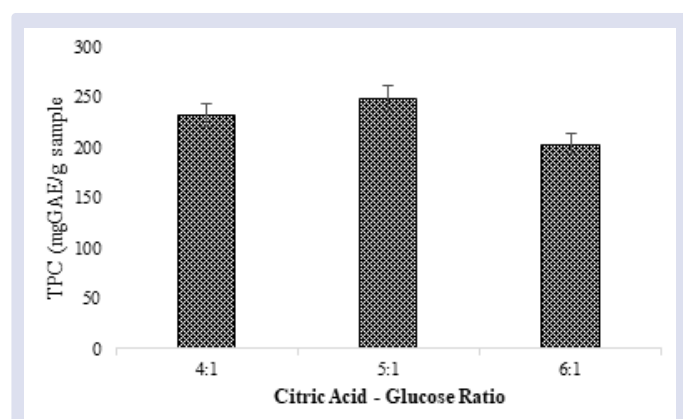


Figure 1: The optimum single factor of NADES composition ratio on polyphenols extraction from *M. speciosa* leaves.

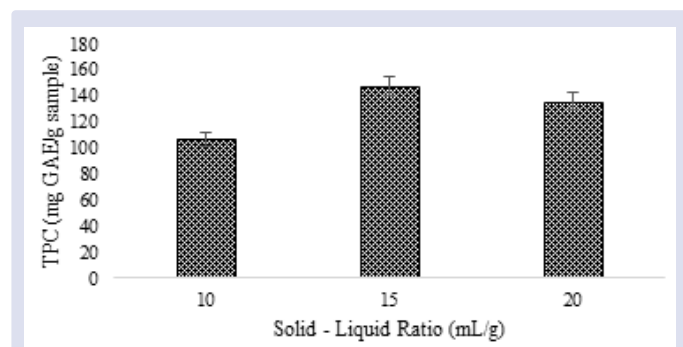


Figure 2: The optimum single factor condition of solid-liquid ratio on polyphenol extraction from *M. speciosa* leaves.

Effect of Microwave Power

The microwave power from the microwave-assisted extraction method can affect the rate of equilibrium and interactions between solvents and solutes (secondary metabolites) in the sample matrix and control the partition of solutes between the sample and the extraction phase.³³ To determine the effect of microwave power on the extraction efficiency of TPC, extraction was carried out at 90, 270, and 450 Watts, respectively, with an of 10 min extraction time, 5:1 g/g NADES ratio, and the solid-liquid ratio of 10:1 mL/g sample.

The results obtained (Table 4) show that the microwave power from 90 to 270 watts has an increased TPC value. This is due to the separation of the secondary metabolite constituents from the sample matrix due to the increase in temperature and pressure due to the microwave power. In contrast, the microwave power of 270 to 450 Watts tends to decrease due to the possibility of secondary metabolite compounds being

damaged due to excessive temperature and pressure. Figure 3 shows the optimum TPC value at 270 watts of microwave power because, in these conditions, there is an equilibrium between temperature and microwave pressure with the extraction process.

Effect of Extraction Time

In Table 5, the effect of extraction time was performed using different the time include 10, 15, and 20 min. In range 10-15 minutes of the efficiency of extraction time shows a tendency to increase in TPC value. The phenomenon indicates that an increase in extraction time can cause an acceleration of the mass transfer of the analyte and a decrease in the viscosity of the extraction solvent.^{38,39} Whereas, if the extraction time was extended by more than 15 min (precisely for 20 min), the TPC value has decreased. This was due to the high microwave power, and the longer extraction time can damage the structure of the components of the target secondary metabolite.⁴⁰ Therefore, According to Figure 4 shows that the optimum extraction time is obtained at the 15th min.

Based on this study's result, the single factor effect of NADES citric acid-glucose-based microwave-assisted extraction with different conditions includes NADES ratio, solid-liquid ratio, microwave power, and extraction time. These results are preliminary data to optimize the

Table 3: Effect of Solid-Liquid Ratio on TPC value.

| Solid-Liquid ratio (mL/g) | Absorbance | TPC (mgGAE/g) | Average TPC (mgGAE/g) | Deviation Standard |
|---------------------------|------------|---------------|-----------------------|--------------------|
| 10:1 | 0.057 | 107.25 | 106.63 | 1.071 |
| | 0.056 | 105.39 | | |
| | 0.057 | 107.25 | | |
| 15:1 | 0.035 | 158.70 | 146.93 | 11.113 |
| | 0.030 | 136.62 | | |
| | 0.044 | 145.45 | | |
| 20:1 | 0.023 | 145.59 | 135.457 | 12.657 |
| | 0.019 | 121.27 | | |
| | 0.055 | 139.51 | | |

Table 4: Effect of different microwave power on TPC value.

| Microwave Power (Watt) | Absorbance | TPC (mgGAE/g) | Average TPC (mgGAE/g) | Deviation Standard |
|------------------------|------------|---------------|-----------------------|--------------------|
| 90 | 0.051 | 175.95 | 156.76 | 17.383 |
| | 0.041 | 142.08 | | |
| | 0.053 | 152.24 | | |
| 270 | 0.065 | 194.16 | 192.20 | 3.399 |
| | 0.063 | 188.28 | | |
| | 0.065 | 194.16 | | |
| 450 | 0.059 | 188.84 | 185.69 | 30.048 |
| | 0.048 | 154.20 | | |
| | 0.058 | 214.04 | | |

Table 5: Effect of different extraction time on TPC value.

| Extraction Time (Minutes) | Absorbance | TPC (mgGAE/g) | Average TPC (mgGAE/g) | Deviation Standard |
|---------------------------|------------|---------------|-----------------------|--------------------|
| 10 | 0.043 | 194.09 | 176.42 | 15.922 |
| | 0.036 | 163.18 | | |
| | 0.038 | 172.01 | | |
| 15 | 0.085 | 371.56 | 358.59 | 11.438 |
| | 0.080 | 349.94 | | |
| | 0.081 | 354.26 | | |
| 20 | 0.039 | 194.09 | 173.48 | 17.847 |
| | 0.037 | 163.18 | | |
| | 0.036 | 173.261 | | |

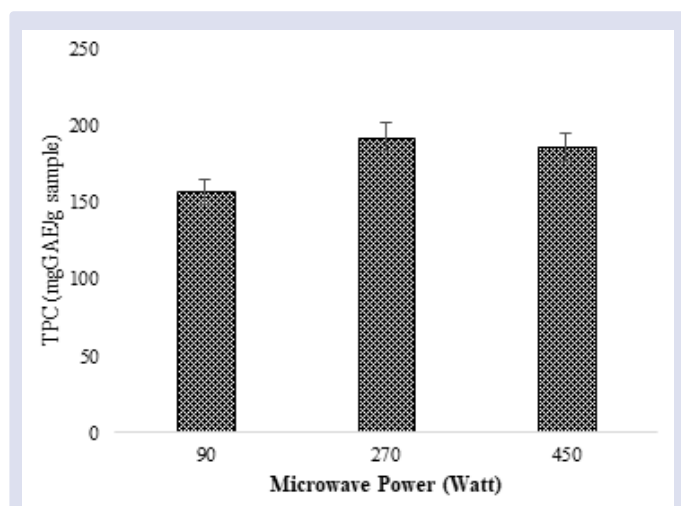


Figure 3: The optimum single factor condition of microwave power on polyphenols extraction from *M. speciosa*.

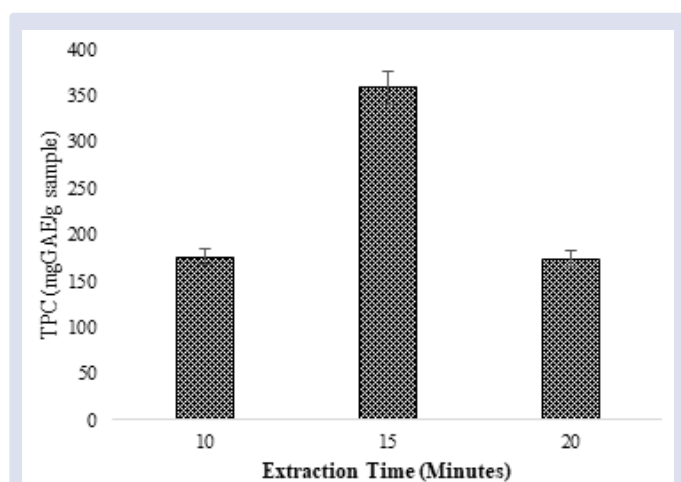


Figure 4: The optimum single factor of extraction time on polyphenolics extraction from *M. speciosa*.

extraction method using the NADES based on microwave-assisted extraction method to obtain the optimum target secondary metabolite from *M. speciosa* leaves quickly, easily, efficiently, and environmentally friendly.

ACKNOWLEDGMENTS

This study was funded and supported by the Ministry of Research, Technology, and Higher Education, Republic of Indonesia and Institute of Research and Community Service, Universitas Mulawarman (LP2M UNMUL) via a grant "Hibah Penelitian Dasar Unggulan Perguruan Tinggi (PDUPT) 2019- 2020.

CONFLICTS OF INTEREST

The authors declared no conflicts of interest.

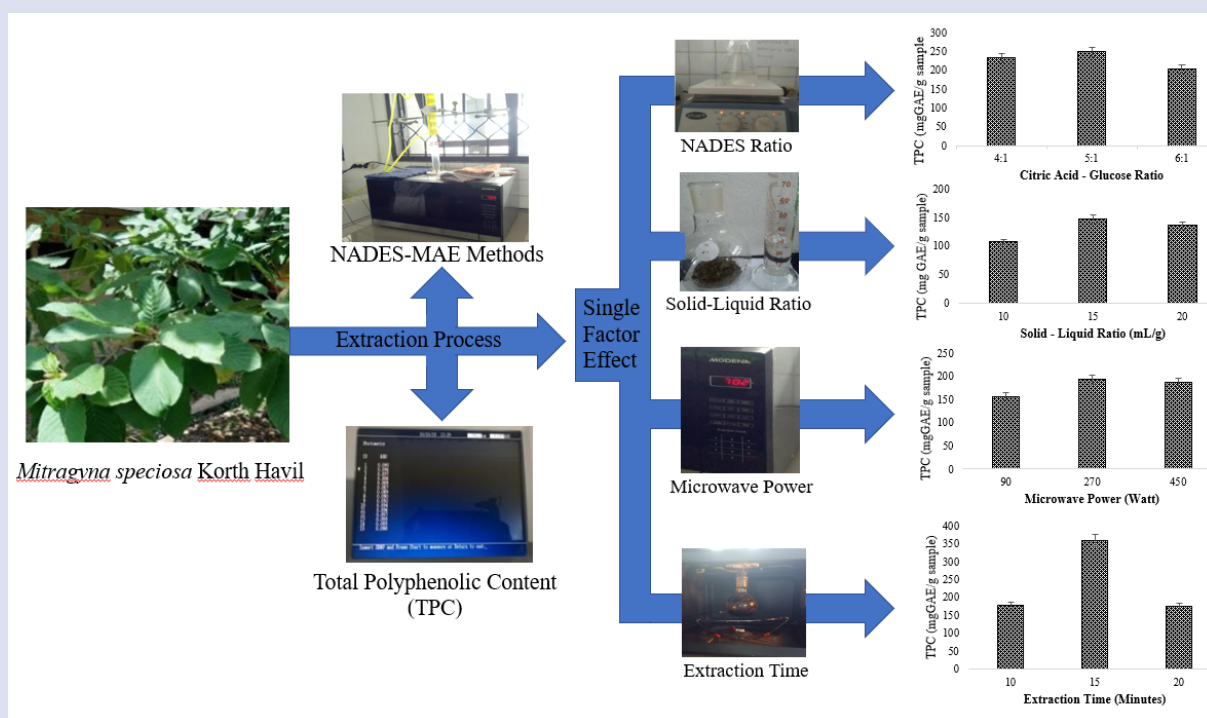
REFERENCES

1. Bubalo MC, Vidovi S, Radoj I, and Joki S. Green solvents for green technologies. *Journal of Chemistry Technology and Biotechnology*. 2015; 90: 1631-1639.
2. Castro VIB, Craveiro R, Silva JM, Reis RL, Paiva A, and Duarte ARC. Natural deep eutectic systems as alternative nontoxic cryoprotective agents. *Cryobiology*. 2018; 83: 15-26.

3. Gomez FJV and Espino M. A greener approach to prepare natural deep eutectic solvents. *Analytical Chemistry*. 2018; 3: 6122-6125.
4. Martins MAR, Pinho SP, Coutinho JAP. Insights into the nature of eutectic and deep eutectic mixtures. *Journal of Solution Chemistry*. 2019; 48(7): 962-982.
5. Craveiro R, Aroso I, Flammia V, Carvalho T, Viciosa MT, Dionisio M, Barreiros S, Reis RL, Duarte ARC, and Paiva A. Properties and thermal behavior of natural deep eutectic solvents. *Journal of Molecular Liquids*. 2016; 215: 534-540.
6. Dai Y, Witkamp GJ, Verpoorte R, and Choi YH. Tailoring properties of natural deep eutectic solvents with water to facilitate their applications. *Food Chemistry*. 2015; 187: 14-19.
7. Dwamena A. Recent advances in hydrophobic deep eutectic solvents for extraction. *Separations*. 2019; 6(1): 1-15.
8. Wei ZF, Wang XQ, Peng X, Wang W, Zhao CJ, Zu YG, and Fu YJ. Fast and green extraction and separation of main bioactive flavonoids from *Radix Scutellariae*. *Industrial Crops and Products*. 2015; 63: 175-181.
9. Savi L, Dias M, Carpine D, Waszczyński N, Ribani R, and Haminiuk C. Natural deep eutectic solvents (NADES) based on citric acid and sucrose as a potential green technology : a comprehensive study of water inclusion and its effect on thermal, physical and rheological properties. *International Journal of Food Science and Technology*. 2018; 54(3): 898-907.
10. Ahmad I, Pertiwi AS, Kembaren YH, Rahman A, and Mun'in A. Application of natural deep eutectic solvent-based ultrasonic assisted extraction of total polyphenolic and caffeine content from Coffee Beans (*Coffea Beans* L.) for instant food products. *Journal Applied of Pharmaceutical Sciences*. 2018; 8(8): 138-143.
11. Wei Z, Qi X, Li T, Luo M, Wang W, Zu Y, and Fu Y. Application of natural deep eutectic solvents for extraction and determination of phenolics in *Cajanus cajan* leaves by ultra performance liquid chromatography. *Separation and Purification Technology*. 2015; 149: 237-244.
12. Duarte ARC, Ferreira ASD, Barreiros S, Cabrita E, Reis RL, and Paiva A. A comparison between pure active pharmaceutical ingredients and therapeutic deep eutectic solvents : Solubility and permeability studies. *European Journal of Pharmaceutics and Biopharmaceutics*. 2017; 114: 296-304.
13. Ahmad K and Aziz Z. *Mitragyna speciosa* use in the northern states of Malaysia: A cross-sectional study. *Journal of Ethnopharmacology*. 2012; 141(1): 446-450.
14. Chittrakarn S, Keawpradub N, Sawangjaroen K, Kansanalak S and Janchawee B. The neuromuscular blockade produced by pure alkaloid, mitragynine and methanol extract of kratom leaves (*Mitragyna speciosa* Korth.). *Journal of Ethnopharmacology*. 2010; 129(3): 344-349.
15. Tohar N, Shilpi JA, Sivasothy Y, Ahmad S and Awang K. Chemical constituents and nitric oxide inhibitory activity of supercritical carbon dioxide extracts from *Mitragyna speciosa* leaves. *Arabian Journal of Chemistry*. 2016; 12(3): 350-359.
16. Brown PN, Lund JA and Murch SJ. A botanical, phytochemical and ethnomedicinal review of the genus *Mitragyna* korth: Implications for products sold as kratom. *Journal of Ethnopharmacology*. 2017; 202: 302-325.
17. Horie S, Koyama F, Takayama H, Ishikawa H, Aimi N, Ponglux D, Matsumoto K, and Murayama T. Indole alkaloids of a Thai medicinal herb, *Mitragyna speciosa*, that has opioid agonistic effect in guinea-pig ileum. *Planta Medica*. 2005; 71(3): 231-236.
18. Takayama H. Chemistry and pharmacology of analgesic indole alkaloids from the rubiaceous plant, *Mitragyna speciosa*. *Chemical and Pharmaceutical Bulletin*. 2004; 52(8): 916-928.

19. Dai Y, van Spreonsen J, Vitkamp GJ, Verpoorte R, and Choi YH. Ionic liquids and deep eutectic solvents in natural products research: Mixtures of solids as extraction solvents. *Journal of Natural Products*. 2013; 76(11): 2162–2173.
20. Dai Y, Witkamp GJ, Verpoorte R, and Choi YH. Natural deep eutectic solvents as a new extraction media for phenolic metabolites in *Carthamus tinctorius* L. *Analytical Chemistry*. 2013; 85(13): 6272-6278.
21. Cui Q, Peng X, Yao XH, Wei ZF, Luo M, Wang W, Xhao CJ, Fu YF, and Zu YG. Deep eutectic solvent-based microwave-assisted extraction of genistin, genistein and apigenin from pigeon pea roots. *Separation and Purification Technology*. 2015; 150: 63-72.
22. Ahmad I, Yanuar A, Mulia K, and Mun'im A. Extraction of polyphenolic content from *Peperomia pellucida* (L) Kunth herb with 1-ethyl-3- methylimidazolium bromide as a green solvent. *Indian Journal of Pharmaceutical Sciences*. 2017; 79(6): 1013-1017.
23. Trinovita E, Sutriyo, Saputri FC, and Mun'im A. Enrichment of the gamma oryzanol level from Rice Bran by addition of inorganic salts on ionic liquid 1-butyl-3- methylimidazolium hexafluorophosphate ([BMIM]PF₆) extraction. *Journal of Young Pharmacists*. 2017; 9(4): 555-558.
24. Mun'im A, Nurpriantia N, Setyaningsih R, and Syahdi RR. Optimization of microwave-assisted extraction of active compounds, antioxidant activity and angiotensin converting enzyme (ACE) inhibitory activity from *Peperomia pellucida* (L.) Kunth. *Journal of Young Pharmacists*. 2017; 9(1): 73-78.
25. Bobo-García G, Davidov-Pardo G, Arroqui C, Marín-Arroyo MR, and Navarro M. Intra-laboratory validation of microplate methods for total phenolic content and antioxidant activity on polyphenolic extracts, and comparison with conventional spectrophotometric methods. *Journal of Science Food and Agriculture*. 2014; 95(1): 204-209.
26. Margraf T, Karnopp AR, Rosso ND and Granato D. Comparison between Folin-Ciocalteu and Prussian Blue assays to estimate the total phenolic content of juices and teas using 96-Well microplates. *Journal of Food Sciences*. 2015; 80(11): 2397-2403.
27. Vongsak B, Sithisarn P, and Mangmool S. Maximizing total phenolics, total flavonoids contents and antioxidant activity of *Moringa oleifera* leaf extract by the appropriate extraction method. *Industrial Crops and Products*. 201; 44: 566-571.
28. Tawaha K, Alali FQ, Gharaibeh M, Mohammad M, El-Elimat T, and Elelimat T. Antioxidant activity and total phenolic content of selected Jordanian plant species. *Food Chemistry*. 2007; 104(4): 1372-1378.
29. Sanchez-Rangel JC, Benavides J, Heredia JB, Cisneros-Zevallos L, and Jacobo-Velasquez DA. The Folin-Ciocalteu assay revisited: improvement of its specificity for total phenolic content determination. *Analytical Methods*. 2013; 21: 1-10.
30. Falleh H, Ksouri R, Lucchessi ME, Abdelly C, and Magné C. Ultrasound-assisted extraction: Effect of extraction time and solvent power on the levels of polyphenols and antioxidant activity of *Mesembryanthemum edule* L. Aizoaceae Shoots. *Tropical Journal of Pharmaceutical Research*. 2012; 11(2): 243-249.
31. Filip S, Pavli B, Vidovi S, Vladi J, and Zekovi Z. Optimization of microwave-assisted extraction of polyphenolic compounds from *Ocimum basilicum* by response surface methodology. *Food Analytical Methods*. 2017; 2017: 1-11.
32. Yin-Leng K and Suyin G. Natural deep eutectic solvent (NADES) as a greener alternative for the extraction of hydrophilic (polar) and lipophilic (non-polar) phytonutrients. *Key Engineering Materials*. 2019; 797: 20-28.
33. Lu W, Alam MA, Pan Y, Wu J, Wang Z, and Yuan Z. A new approach of microalgal biomass pretreatment using deep eutectic solvents for enhanced lipid recovery for biodiesel production. *Bioresources Technology*. 2016; 218: 123-128.
34. Fernández M de los Á, Espino M, Gomez FJV, and Silva MF. Novel approaches mediated by tailor-made green solvents for the extraction of phenolic compounds from agro-food industrial by-product. *Food Chemistry*. 2017; 239: 671-678.
35. Wang H, Ma X, Cheng Q, Xi X, and Zhang L. Deep eutectic solvent-based microwave-assisted extraction of Baicalin from *Scutellaria baicalensis* Georgi. *Journal of Chemistry*. 2018; 2018: 1-10.
36. Ma W, Lu Y, Hu R, Chen J, Zhang Z, and Pan Y. Application of ionic liquids based microwave-assisted extraction of three alkaloids N-nornuciferine, O-nornuciferine, and nuciferine from lotus leaf. *Talanta*. 2010; 80(3): 1292-1297.
37. Ahmad I, Yanuar A, Mulia K, and Mun'im A. Ionic liquid-based microwave-assisted extraction: Fast and green extraction method of secondary metabolites on medicinal plant. *Pharmacognosy Reviews*. 2018; 12(23): 20-26.
38. Cheng Z, Song H, Yang Y, Liu Y, Liu Z, Hu H, and Zhang Y. Optimization of microwave-assisted enzymatic extraction of polysaccharides from the fruit of *Schisandra chinensis* Baill. *International Journal of Biological Macromolecules*. 2015; 76: 161-168.
39. Handayani D, Mun'im A, and Ranti AS. Optimization of green tea waste extraction using microwave assisted extraction to yield green tea extract. *Traditional Medicine Journal*. 2014; 19(1): 29-35.
40. Xiong W, Chen X, Lv G, Hu D, Zhao J, and Li S. Optimization of microwave-assisted extraction of bioactive alkaloids from lotus plumule using response surface methodology. *Journal of Pharmaceutical Analysis*. 2016; 6(6): 382-388.

GRAPHICAL ABSTRACT



SUMMARY

- The best of single factor condition effect of NADES-MAE method was obtained the TPC value including:
 - NADES ratio of 248.69 mg GAE/g sample (5:1 g/g citric acid-glucose)
 - A solid-liquid ratio of 146.93 mg GAE/g sample (15:1 mL/g solvent-sample)
 - Microwave power of 192.20 mg GAE/g sample (270 Watts)
 - Extraction time of 358.59 mg GAE/g sample (15 min).
- The single factor effect of NADES citric acid-glucose-based MAE shows a difference in TPC value based on various conditions of the NADES-MAE method.

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