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Review Article

Understanding the challenges to COVID-19 vaccines and treatment options, herd immunity and probability of reinfection



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الملخص

على عكس الأوبئة في الماضي، شهد تقشي مرض كوفيد-19 الذي ينتقل بسرعة في جميع أنحاء العالم نهجا مختلفا تماما للضوابط والتدابير التي تم تنفيذها في جميع البلدان المتضررة. لا يزال عدم فهم الطبيعة الأساسية لتقشي المرض يجعل من كوفيد-19 تحديا لكل من ممارسي الرعاية الصحية والمجتمع العلمي. لا تزال التحديات التي تواجه تطوير اللقاح وتقييمه، والخيارات العلاجية الحصرية، وعلاج البلازما في فترة النقاهة، ومناعة القطيع، فضلا عن ظهور العدوى مرة أخرى والمتغيرات الجديدة، هي العقبات الرئيسية أمام مكافحة كوفيد-19. تناقش هذه المراجعة هذه التحديات في إدارة كوفيد-19 بشكل مفصل وتسلط الضوء على الآيات اللازمة لمساعدتنا على فهم هذا الوباء بشكل أفضل.

الكلمات المفتاحية: سارس؛ كوفيد-19؛ فيروس كورونا؛ عودة العدوى؛ علاج البلازما؛ مناعة القطيع

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Abstract

Unlike pandemics in the past, the outbreak of coronavirus disease 2019 (COVID-19), which rapidly spread worldwide, was met with a different approach to control and measures implemented across affected countries. The lack of understanding of the fundamental nature of the outbreak continues to make COVID-19 challenging to manage for both healthcare practitioners and the scientific community. Challenges to vaccine development and evaluation, current therapeutic options, convalescent plasma therapy, herd immunity, and the emergence of reinfection and new variants remain the major obstacles to combating COVID-19. This review discusses these challenges in the management of COVID-19 at length and highlights the mechanisms needed to provide better understanding of this pandemic.

Keywords: 2019-nCoV; Convalescent plasma therapy; Coronavirus; Herd immunity; Reinfection; SARS-CoV-2

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Introduction

Since it was first reported in Wuhan, China, in December 2019, communicable coronavirus disease 2019 (COVID-19) caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) continues to impose health and socioeconomic threats globally. SARS-CoV-2 belongs to the coronavirus family but is different from previous human coronaviruses (e.g., SARS-CoV and Middle East respiratory syndrome CoV [MERS-CoV]) in that it results in disease progression that is difficult to predict and is varied. SARS-CoV-2 has a receptor domain with high affinity toward angiotensin-converting enzyme 2 (ACE-2), which is abundant in many organs (Figure 1), thus explaining the varying symptoms and severity of COVID-19.

The medical community initially expected that reaching the vaccination stage would not be feasible for a very long time. Consequently, research studies and clinical trials were initiated to assess the potential effects of several existing drugs against COVID-19.4 Furthermore, owing to its extensive history of efficacy against human respiratory viral infections, convalescent plasma (CP) therapy was used to treat COVID-19.5 This treatment option was used primarily for severely ill patients hospitalized with COVID-19, and has been shown to be effective for reducing the mortality risk of COVID-19. However, given the unprecedented speed of community transmission, the CP therapy did not assisted in achieving herd immunity to COVID-19, which requires a sufficient number of people in a population to have developed protective antibodies against SARS-CoV-2 through either vaccination or natural infection. Hence, the

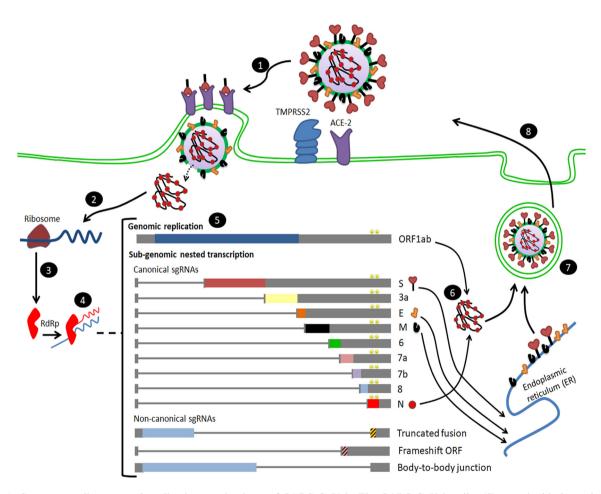


Figure 1: Structure, cell entry and replication mechanisms of SARS-CoV-2. The SARS-CoV-2 spike (S) protein binds angiotensin-converting enzyme 2 (ACE2). The transmembrane protease serine 2 (TMPRSS2) protease enzyme cleaves the S protein, thus allowing viral entry into the host cell cytosol after fusion of the viral and cellular membranes (1). SARS-CoV-2 genomic RNA is translated into non-structural proteins (NSPs) from two open reading frames (ORF1a and ORF1b) (2). Some of the NSPs have RNA-dependent RNA polymerase (RdRp) activity (nsp12) (3). Negative-sense RNA intermediates are generated to serve as the templates for the synthesis of positive-sense genomic RNA (gRNA) (4) and sub-genomic RNAs (sgRNAs) (5). The gRNA is packaged by the structural proteins to assemble progeny virions. Shorter sgRNAs encode conserved structural proteins (S, E, M and N) and several accessory proteins. SARS-CoV-2 is known to have at least six accessory proteins (3a, 6, 7a, 7b, 8 and 10), and non-canonical sgRNAs are also shown in the subfigure (6). The AAGAA-type modification clusters in gRNA and sgRNAs are shown with yellow star annotations. Thereafter, budding and exocytosis of the virus occurs (7 and 8) (Adapted from Hatmal et al., 2020³).

world awaited COVID-19 vaccines that offered hope to defeat the COVID-19 pandemic.

Currently, six vaccines (Pfizer-BioNTech, Moderna, AstraZeneca-Oxford, Johnson & Johnson, Sputnik V, Sinopharm and Sinovac) are authorized and available for immunization against COVID-19. However, no evidence indicates that recovered people with existing SARS-CoV-2 antibodies are protected against a second infection of COVID-19, and a growing number of studies report COVID-19 reinfections. This comprehensive review provides a broad overview of the challenges associated with COVID-19 vaccines and treatment options, including the potential of CP therapy, herd immunity, and the probability of reinfection.

SARS-CoV-2 variants

Since the early stages of the COVID-19 pandemic, multiple strains of SARS-CoV-2 have emerged and spread worldwide. In open-source platforms such as Pango, ¹⁰ Nexstrain (3103 genome sequences), ¹¹ GISAID (10,298,748 genome sequences), ¹² and NCBI Virus (4,616,599 nucleotide sequences), ¹³ more than 5 million genome sequences have been provided. According to their phylogenetic analyses, clades have been determined according to a collection of common mutations ¹⁴ and the ancestral clade known as clade O. ¹⁵ Other clades such as L, S and A2a, as well as clade G, have become more prevalent after 2020. ^{16–18}

According to the WHO, ¹⁹ the European Centre for Disease Prevention and Control (ECDC) ²⁰ and Centers for Disease Control and Prevention (CDC), ²¹ a classification system has been developed to differentiate variants of concern (VOCs), variants of interest (VOIs), variants under monitoring (VUM), variants being monitored (VBM), formerly monitored variants (FMV) and variants of high consequence (VOHC). ²² After the emergence of variants that posed a greater risk to global public health in late 2020, characterization of specific VOCs, VUMs, VOIs and VOHCs became necessary to prioritize research and global monitoring, and to influence the continuing response to the COVID-19 pandemic. ¹⁹

COVID-19 variants that meet specific criteria, such as high transmission or adverse alterations in COVID-19 distribution; changes in clinical illness presentation or elevated clinical virulence; diminished effectiveness of social interventions and public health measures; or diminished effectiveness of diagnostics, treatments and vaccinations are defined as VOCs. ^{19,23} VOCs are mutant strains that are relatively more transmissible, elude antibody neutralization, induce more severe disease progression, confound identification and increase the incidence of mortality.²² Particular VOCs may have selective benefits, on the basis of the existence of the same mutations repeatedly emerging in many virus strains different geographic locations (i.e., convergent evolution). 11,20,24,25 Most VOCs studied since the SARS-CoV-2 pandemic have had relatively higher transmissibility and virulence; these were the Alpha, Delta, Gamma and Beta variants. Although Gamma, Alpha, and Beta are no longer designated as VOCs, studying the causes that led to their emergence in populations is crucial.²² According to the SARS-CoV-2 Interagency Group of the United States government, the Omicron variant is considered a VOC. 21 VOCs

include Delta (1.617.2) and Omicron (B.1.1.529, BA.1, BA.2, BA.3, BA.4, BA.5, XE [a recombinant of Omicron BA.1 and BA.2] and descendent lineages). ^{19,21}

In contrast, VOIs are characterized as genetically mutated variants with predicted or known effects on viral characteristics, such as disease severity, transmissibility, immunological breakout, and therapeutic or diagnostic escape.²³ In addition, these variants are characterized by the presence of specific genetic markers that may be associated with high transmissibility or virulence, the ability to escape detection, a decrease in neutralizing antibodies (NAbs) acquired via natural infection or vaccination, or a decrease in the effectiveness of vaccination or therapeutics.² Furthermore, these variants have been identified as causing substantial and unique COVID-19 outbreak clusters or community transmission in various countries, with corresponding increases in the prevalence rates or other clear epidemiological effects, thus indicating a rising risk to global public health. 19 VOIs include Omicron (BA.2.75) and Omicron (BA.2 + L452X (x)). 2

VUMs are viral variants that bear genetic modifications suspected to affect the characteristics of the virus and that show indications of future risk to human health. 19 Moreover, these variants are associated with more severe disease or greater transmission, but are either no longer detectable or are circulating at extremely low levels. However, the evidence of epidemiological or phenotypic influence is now uncertain, thus necessitating increased monitoring and reassessment in the absence of additional evidence. A VOI or VOC may be relegated to this category if its national and regional percentages have decreased significantly and consistently over time, or if other evidence suggests that a variant does not constitute a major risk to public health.²¹ This category includes former VOIs/VOCs that will continue to be tracked for an extended length of time and will retain the WHO label assigned to them until further notice. Some evidence suggests that they may have characteristics identical to those of VOCs; however, the evidence is either insufficient or has not yet been evaluated by the ECDC. According to the ECDC, VUMs include Omicron (BA.3).²⁰ The WHO has established a new category in its variant tracking system, designated "Omicron subvariants under monitoring," to indicate to public health authorities worldwide which VOC lineages may warrant prioritization and monitoring, in light of the broad transmission of the Omicron VOC worldwide and the expected increase in its viral diversity. Omicron subvariants under monitoring include BA.4 and BA.5, as well as lineages that share the same cluster of mutations in the spike, namely BA.2.12.1, BA.2.9., BA.2.11, BA.2.13 and BA.2.75, and the following variations seen outside the spike: BA.4: ORF7b:L11F, N:P151S; BA.5: M:D3N; ORF1a:del141/143 and ORF6:D61L. 19

Since September 2021, the ECDC has categorized some variants as de-escalated variants,²⁰ although these variants had been classified as VUMs by the CDC and as VOIs and/or VOCs by the WHO.^{19,21} These variants meet at least one of the following criteria: (1) the variant has spread for a long period of time without causing any overall epidemiological effects; (2) the variant is no longer circulating; and (3) scientific research has demonstrated that the variant does not contain any concerning features.

Recently, on the basis of ECDC advice, the de-escalated variants have come to include Alpha (B.1.1.7), Beta (B.1.351), Gamma (P.1), Epsilon (B.1.427 and B.1.429), Zeta (P.2), Eta (B.1.525), Theta (P.3), Iota (B.1.526), Kappa (B.1.617.1), B.1.640, Lambda (C.37) and Mu (B.1.621).

In this section, we discuss the main COVID-19 variants along with their severity, transmissibility and immunity characteristics (Table 1). Although some SARS-CoV-2 variants (e.g., B.1.1.7, P.1 and 501Y. V2) have been identified to be more transmissible variants and are already spreading rapidly worldwide, ^{26–28} Delta and Omicron currently have the highest transmissibility and virulence. 22 Although Delta has dispersed worldwide, Omicron is growing at a faster rate than Delta, and it bears an increased risk of transmission, particularly among contacts outside the household. 29,30 Consequently, although early indications have suggested that individuals may be less likely to require hospitalization, a much greater number of people are anticipated to become infected. Regular contact tracing analyses have revealed that transmission rates among people infected with BA.2 are expected to be higher (13.4%) than those among household contacts of other people infected with Omicron (10.3%).30 The higher transmissibility of these variants has been associated with the mutations that they carry, including mutations that alter their S proteins (particularly the receptor-binding domain), the main target of COVID-19 vaccines and therapeutics.³ These mutations may decrease the binding affinity of anti-SARS-CoV-2 antibodies.³¹ Therefore, the growing number of more transmissible SARS-CoV-2 variants may cause more reinfections. In addition, existing vaccines may require future updates to improve their efficacy, particularly if vaccine-resistant SARS-CoV-2 variants emerge.

Vaccine development against COVID-19

Vaccines are artificial or natural biological preparations used to stimulate the immune response for antibody production against a particular disease. A vaccine typically consists of a pathogenic agent (either the microorganism itself, or its specific antigen epitope, surface protein, toxin or a synthetic substitute) treated to act as an antigen without causing disease in a host. A vaccine works by "educating" the immune system to recognize and fight against microbial pathogens, i.e., either viruses or bacteria. Thus, a particular molecule (e.g., antigen epitope, surface protein or toxin) from the pathogen must be presented to the host to induce an immune response. The production of antibodies from the given antigen provides immunity, which helps the immune system become stronger when it encounters a real infection.

Vaccines aid in developing immunity against particular pathogens by imitating an infection caused by the given pathogen. Although this type of imitated infection almost never causes illness, it stimulates the immune system to generate T cell populations and antibodies. As the body builds immunity, minor symptoms, such as fever, may develop and are considered normal. After the period of the imitated infection passes, the body is left with a population of memory T cells and B cells, which provide resistance against the particular disease in the future. Although the

steps in the traditional method for developing new vaccines tend to be sequential and well established (Figure 2), some overlap exists between the stages. Because each phase is costly, particularly later phases, and every phase enhances understanding of whether the next step might be successful, the objective of this sequential strategy is partly to lessen the financial risk involved. The process can be halted at any point for a variety of reasons, including the detection of adverse events, such as major adverse effects, or if the data indicate that the vaccine is likely to be ineffective.^{89,90} Combining phases of vaccine development can accelerate the process in situations such as the upheaval created by the COVID-19 pandemic and the urgent need for a vaccine that is both effective and widely available worldwide. 91 As of October 27, 2022, on the basis of the recent COVID-19 landscape report by the WHO, a total of 172 and 199 vaccine candidates are currently in clinical development and preclinical development worldwide, respectively. 92 As shown in the same report, these vaccine candidates have been developed by using different platforms including, but not limited to, protein subunits (32.2%), RNA (23.4%), nonreplicating viral vectors (13.5%), inactivated viruses (12.9%), DNA (9.4%), virus-like particles (3.5%) and replicating viral vectors (2.3%). Figure 3 summarizes the immunological defense mechanisms involved in developing immunity to SARS-CoV-2 in either infected or vaccinated individuals. 93-99

COVID-19 vaccine challenges

Infectious diseases can have enormous effects and devastating consequences on population health and country economies. Because prevention is always a better option than treatment, vaccines provide the best solution to combat infectious disease. Beyond providing long-term protection against infections, the ideal types of vaccines should be safe, easy to administer, simple and inexpensive to manufacture. 100 Careful evaluations at each step are required to develop novel vaccines while ensuring effectiveness and safety. Hence, the pipeline for traditional vaccine development usually requires more than 10 years before reaching the general public.¹⁰¹ However, the highly infectious SARS-CoV-2 and the ensuing morbidity and mortality required urgent development of vaccines despite the challenges in vaccine development. The development of COVID-19 vaccines faces several policy challenges, as shown in Figure 4. Therefore, the Government Accountability Office in the USA has highlighted nine policy options that might help overcome challenges to vaccine development technologies and methods, as well as economic challenges (Figure 4).

Immune responses

Antigen selection for vaccine development requires a critical evaluation of whether to use whole-cell antigens (killed or weakened) or fragments of their viral materials (subunits). For the COVID-19 vaccine, vaccinologists are currently focusing on the S protein, which is believed to facilitate cell membrane fusion and viral entry. ¹⁰³ Of note, vaccination should trigger a proper robust immune response to ensure long-term protection while posing no

WHO label	Lineage (Pango)	GISAID clade	Other names	Most common countries (earliest date)	Status from September 2021	Effect on severity	Effect on transmissibility	Effect on immunity	References
Delta	B.1.617.2 (Indian variant)	G/478K. V1	VUI-21APR-02	USA 21.0%, India 18.0%, UK 13.0%, Turkey 11.0%, Germany 5.0%	WHO: previously circulating VOC CDC: VBM ECDC: de- escalated	Increased	Increased	Increased	32–36
Alpha	B.1.1.7 (British variant)	GRY	VOC 202012/01, 20I/501Y.V1	UK 24.0%, USA 20.0%, Germany 9.0%, Sweden 6.0%, Denmark 6.0%	WHO: previously circulating VOC CDC: VBM ECDC: de-escalated	Increased	Identical to previous variants in that it was unlikely to have an effect	Increased	37–40
Beta	B.1.351 (South African variant)	GH/501Y. V2	20H/501Y. V2	South Africa 19.0%, Philippines 9.0%, USA 9.0%, Sweden 8.0%, Germany 7.0%	WHO: previously circulating VOC CDC: VBM ECDC: de-escalated	Increased	Increased	Increased	41,40,42-45
Gamma	P.1 (Brazilian variant)	GR/501Y. V3	20J/501Y. V3	Brazil 56.0%, USA 29.0%, Chile 3.0%, Argentina 2.0%, Spain 1.0%	WHO: previously circulating VOC CDC: VBM ECDC: de-escalated	Increased	Increased	Increased	40,46-48
Epsilon	B.1.427 and B.1.429 (Californian variant)	GH/452R. V1	CAL. 20C/L452R	B.1.427: USA 98.0%, Mexico 1.0%, Aruba 0.0%, Argentina 0.0%, Canada 0.0% B.1.429: USA 97.0%, Canada 2.0%, Mexico 1.0%, South Korea 0.0%, Chile 0.0%	WHO: previously circulating VOI CDC: VBM ECDC: de-escalated	Unclear	Increased	No evidence	20,49-51
Eta	B.1.525	G/484K. V3	-	Canada 19.0%, USA 15.0%, Germany 9.0%, France 8.0%, Denmark 7.0%	WHO: previously circulating VOI CDC: VBM ECDC: de-escalated	No evidence	Increased	No evidence	20,52-54
Theta	P.3	GR/1092K. V1	-	Philippines 83.0%, USA 3.0%, Germany 2.0%, Malaysia 2.0%, UK 2.0%	WHO: previously circulating VOI CDC: N/A ECDC: de-escalated	Increased	Increased	No evidence	38,53
Kappa	B.1.617.1	G/452R. V3	_	India 72.0%, UK 6.0%, Canada 6.0%, USA 5.0%, Ireland 3.0%	WHO: previously circulating VOI CDC: VBM ECDC: de-escalated	Increased	Increased	No evidence	55-58
Zeta	P.2	GR/484K. V2	VUI- 202101/01, B.1.1.28.2	Brazil 57.0%, USA 24.0%, Canada 4.0%, Argentina 2.0%, Paraguay 2.0%	WHO: previously circulating VOI CDC: VBM ECDC: de-escalated	No evidence	Increased	No evidence	20, 53, 59
Lambda	C.37	GR/452Q. V1	_	Peru 44.0%, Chile 19.0%, Argentina 13.0%, USA 12.0%, Ecuador 3.0%	WHO: previously circulating VOI CDC: N/A ECDC: de-escalated	No evidence	Increased	No evidence	60-62

Mu	B.1.621	GH	_	USA 40.0%, Colombia 27.0%, Chile 8.0%, Spain 4.0%, Panama 4.0%	WHO: previously circulating VOI CDC: VBM	Increased	Increased	No evidence	38,53,63
Iota	B.1.526	GH/253G. V1	-	USA 97.0%, Ecuador 1.0%, Canada 1.0%, Puerto Rico 1.0%, Spain 0.0%	ECDC: de-escalated WHO: previously circulating VOI CDC: VBM	No evidence	Increased	No evidence	20,53,64
N/A	XD (France variant)	_	_	France 68.0%, Denmark 28.0%, Netherlands 4.0%	ECDC: de-escalated WHO: FMV CDC: N/A ECDC: de-escalated	No evidence	No evidence	No evidence	20,65
N/A	XE (British	_	_	UK 99.0%, USA 1.0%, Ireland	WHO: VOC CDC: N/A	No evidence	No evidence	No	20,66
N/A	variant) XF (British variant)	_	-	0.0%, Denmark 0.0% UK 100.0%	ECDC: N/A WHO: N/A CDC: N/A ECDC: de-escalated	No evidence	No evidence	evidence No evidence	20,67
Omicron	BA.1 (B.1.1.529)	GR/484A	_	USA 38.0%, UK 29.0%, Canada 5.0%, Germany 4.0%,	WHO: VOC CDC: VOC	Increased	Increased	Decreased	68-75
Omicron	BA.2 (B.1.1.529)	GR/484A	-	Norway 3.0% UK 67.0%, Germany 8.0%, USA 4.0%, Denmark 3.0%,	ECDC: VOC WHO: VOC CDC: VOC	Increased	Increased	Decreased	69,76-80
Omicron	BA.3 (B.1.1.529)	GR/484A	_	France 3.0% UK 35.0%, Poland 24.0%, Germany 17.0%, South Africa	ECDC: VOC WHO: VOC CDC: VOC	No evidence	No evidence	No evidence	20,81
Omicron	BA.4 (B.1.1.529)	GR/484A	_	5.0%, USA 4.0% UK 28.0%, USA 27.0%, South Africa 8.0%, Israel 5.0%,	ECDC: VUM WHO: VOC CDC: VOC	No evidence	Increased	No evidence	20,82
Omicron	BA.5 (B.1.1.529)	GR/484A	_	Denmark 5.0% USA 24.0%, UK 19.0%, Denmark 9.0%, France 5.0%,	ECDC: VOC WHO: VOC CDC: VOC	No evidence	Increased	No evidence	20,83
Omicron	BA.2 + L452X	GR/484A	-	South Africa 5.0% UK 32.0%, Germany 13.0%, Denmark 13.0%, USA 11.0%,	ECDC: VOC WHO: VOC CDC: VOC	No evidence	Increased	No evidence	76,84
Omicron	BA.2.75	GR/484A	_	France 5.0% India 76.0%, UK 8.0%, USA 4.0%, New Zealand 3.0%, Canada 2.0%	ECDC: VOI WHO: VOC CDC: VOC ECDC: VOI	No evidence	No evidence	No evidence	84,85

VOC, variants of concern; VOI, variants of interest; VUM, variants under monitoring; VBM, variant being monitored; FMV, formerly monitored variants; N/A, not applicable.

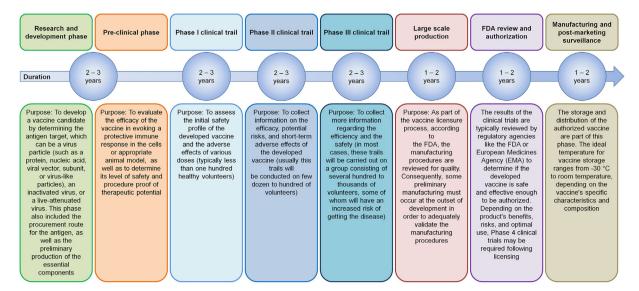


Figure 2: Conventional vaccine development process. The conventional process for developing vaccines includes several phases, beginning with research and development; then pre-clinical studies; then phase I trials in a relatively small number of control volunteers; and finally more extensive phase II and phase III trials. After that, the Food and Drug Administration or the European Medicines Agency evaluates the findings of the clinical trials to determine whether the vaccine is sufficiently effective and safe to be authorized and is also affordable. The final phase is manufacturing and post-marketing surveillance, which is completed after the vaccine has been made available for use by the general public and after it has been monitored for its overall effectiveness among the population. Any negative adverse effects that may arise from the vaccination programs are also documented in the vaccine reports in this phase.

harm to people. However, the emergence of several new strains and mutant variants has resulted in infections in fully vaccinated people and even reinfections among people who have recovered from COVID-19. Thus, to select the antigen with the optimal immune response, many studies are needed to fully understand how the immune system responds to SARS-CoV-2 ⁵⁵. However, developing novel SARS-CoV-2 vaccines may be challenging because of the many unknowns regarding this virus. Therefore, further investigation is warranted.

Vaccine adjuvant and delivery system

In the early years of vaccine development, Louis Pasteur introduced inactivated pathogens (attenuated viruses) to stimulate the immune system. 104 Since then, mainstream global vaccine development has evolved into a more minimalist design approach, wherein only one subunit of the peptide-based composition of the antigenic material is used in the formulation instead of the whole virus. Despite the higher safety and usually lower production cost for this type of vaccines, adjuvants are required because they are less immunogenic than inactivated whole-cell vaccines. 104,105 Adjuvants are often designed to function as both immunostimulators and delivery systems to enhance vaccine immunogenicity. To date, aluminum salts, which are very inexpensive to produce, have been the most commonly used vaccine adjuvant. They have been approved by the Food and Drug Administration since the 1930s because of their safety and long-term protection. Despite showing an excellent safety profile and the longest history of use, aluminum adjuvant-containing vaccines also have some limitations. For example, aluminum-containing salts have been reported to induce strong humoral responses but not cellular responses. 105 An ideal immunization for adaptive immunity must provide long-term protection that involves both humoral and cellular immunity for a rapid response and immunological memory to fight future infections by pathogenic agents.⁸⁷ In addition, cellular immune responses have been suggested to be valuable for respiratory tract infections such as SARS-CoV-2, owing to the late onset of the production of antibodies after infection. ¹⁰⁸ In adaptive immunity, the CD8⁺ T cell subset is involved primarily in the cellular response, whereas CD4+ T cells are responsible for longterm protection against subsequent infection. 108 Hence, the limited number of clinically approved adjuvants has restricted the choices for novel vaccines under development and prolonged the vaccine pipeline. The development of COVID-19 vaccines should, of course, meticulously consider every feature, including the type of adjuvants, to ensure optimal immunostimulation and antigen delivery. These considerations are complicated by the incomplete understanding of the exact mechanism of action of SARS-CoV-2, given its relatively recent emergence.

Financial commitment

The development of new vaccines such as those for COVID-19 is costly, particularly during this pandemic. As described, the typical timeline for previous vaccine development processes was 10 years or more. However, during the emerging pandemic disease, shortening of the timeline for the vaccine development was necessary. For instance, the mRNA-1273 vaccine candidate from Moderna entered a phase 1 clinical trial on March 16, 2020, less than 10 weeks after China had released the genetic sequence of SARS-CoV-2 ⁵⁵. However, accelerating the process, such as by conducting research phases in parallel with other phases, poses a

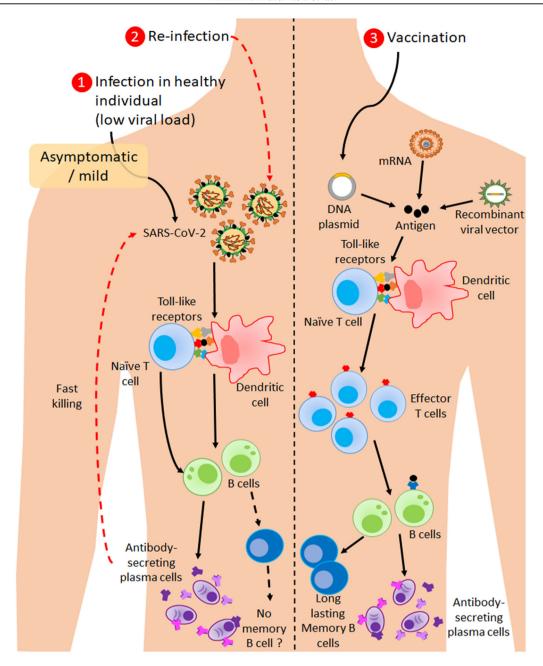


Figure 3: Schematic representation of the mechanisms of development of immunity to SARS-CoV-2 in three situations. 1) Infection in a healthy individual (low viral load). 2) Reinfection with SARS-CoV-2 when previously infected people did not have an adequate neutralizing IgG response, which formed only after their secondary infection. Usually, the innate immune response regulates the immediate host response to SARS-CoV-2 infection, which is then accompanied by an early antibody response. Development of high titers of IgM, IgA, and IgG virus-specific antibodies peaks 3-4 weeks after symptom onset, then decreases. In some cases, IgM and IgA levels exceed baseline levels 2 months after onset, whereas IgG levels remain elevated for as many as 3 months in most patients. Intriguingly, the frequency and durability of the COVID-19 antibody response tend to be correlated with the severity of the disease. In patients who experienced asymptomatic infection or mild illness, short-lived immunity has been noted, thus indicating that a low neutralizing antibody response has been established, with the virus effectively cleared by the innate and early antibody response. In asymptomatic/mild cases of COVID-19, the viral-clearing antibody response wanes very quickly (<90 days). Therefore, these people may be vulnerable to reinfection. High titers of NAbs have been found in the more extreme cases of COVID-19, and notably, no cases of reinfection of immunocompetent individuals have been identified. 3) Potential effects of different types of vaccines (DNA-based, RNA-based and non-replicating viral vector) after administration to the host. A vaccine works by imitating an infection caused by a particular pathogen for which the vaccine is developed. After the vaccine is injected into the host, the antigen inside the vaccine component is captured by antigen-presenting cells (APC), such as dendritic cells (DC), by phagocytosis. Subsequently, the DCs travel to a secondary lymphoid organ, such as lymph nodes, where immature T and B cells are situated. The antigen processed by the DCs is bound to the major histocompatibility complex (MHC) class II receptor on the DC surface. The antigen is then presented to immature T helper cells through binding the MHC class II to the T cell receptor (1st signal). Later, these T lymphocytes proliferate and mature. T helper cells activate B cells, which then proliferate into antibody-secreting plasma cells (2nd signal). These memory T and B cells are formed after the imitation infection by the vaccine ends.

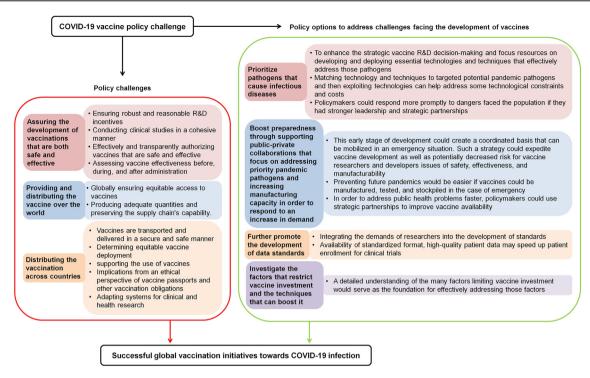


Figure 4: Challenges and policy options to address the challenges in the development of COVID-19 vaccines. The three primary "D" dimensions of obtaining broad global COVID-19 immunity by vaccinations are development, dissemination and deployment, which ensure continuing development of effective and safe vaccines, providing and distributing the vaccine globally, and deploying the vaccine across countries. To achieve these goals, 11 challenges must be addressed: maintaining robust and reasonable R&D incentives; conducting cohesive clinical trials; authorizing effective and safe vaccines; surveilling effectiveness after and during vaccination; ensuring equitable availability of the vaccine worldwide; producing adequate amounts and sustaining supply chain ability; safely transferring and supplying vaccines; deploying vaccines equitably; supporting vaccine use; addressing ethical concerns re: vaccine passports and other obligations; and adapting systems for clinical and health research. From the beginning of vaccine R&D until the adaption of the clinical system, financial and ethical decisions must be made. Decause vaccine development methods and techniques face numerous challenges, the Government Accountability Office has outlined several policy options for overcoming these obstacles. Policymakers, including those in the US Congress, state and local governments, federal agencies, academic and research organizations, and private industry, could take one of several new actions as a result of these policy options.

substantial financial risk, particularly in view of the many unknowns surrounding COVID-19. Nevertheless, the cost for the R&D of a new vaccine has not been disclosed by the pharmaceutical company, which has lacked transparency regarding the actual costs. However, on the basis of common costs for new drug discovery, the range for the true cost has been estimated to have been approximately \$2.6 billion. 109 Beyond the financial cost of R&D, advanced technologies, which are often available to only high-income nations, are required for the development of COVID-19 vaccines. Consequently, the development of novel COVID-19 vaccines is very challenging, particularly in low-and middle-income countries (LMICs), owing to the enormous financial commitments required for R&D.

Because of global inequality in terms of vaccine development and production capacity, programs must be developed to enable the distribution of vaccines on the basis of population needs rather than wealth or social status. Previously, operation Warp Speed, a US program, aimed to develop vaccines faster than ever before, to make them accessible for emergency use by the end of 2020 and to produce billions of doses by the end of the 2021. However, vaccine manufacturing and procurement cost

billions of dollars in high-income nations, thus resulting in an uneven distribution of vaccines to LICs. ¹¹¹ Despite the recommendations made at the Global Vaccination Summit in favor of equal vaccine allocation, concerns remain that some governments might want to secure their citizens' vaccine supplies before sharing vaccines globally. ¹¹² Several countries, such as the United States and Europe, have indicated that vaccines will initially be delivered to their own citizens because of the high demand for vaccines. However, questions have been raised regarding the ethics of distributing resources unequally. Nonetheless, positive steps have been made, including by the COVAX initiative and AstraZeneca's announcement of a partnership with an Indian institute to provide appropriate doses to LMICs. ¹¹³

The unequal distribution of vaccine supplies worldwide has substantially affected progress in vaccination in LICs. According to a United Nations report released on March 28, 2022, vaccination distribution has increased worldwide but still shows inequality. Only 1% of the 10 billion doses distributed worldwide went to LICs, thus leaving about 2.8 billion individuals waiting for their first dose. 114,115 To date, slightly more than 3% of people have been vaccinated with at least one dose in LICs, as compared with 60% in high-

income countries. As of the middle of 2022, the United Nations health agency sought vaccination for 70% of the world's population, ¹¹⁴ a goal scarcely attainable. Approximately 39 high-income nations have already met the 70% target as of January 2022, whereas the Republic of Congo, Ethiopia, South Sudan and Nigeria still seek to acquire sufficient vaccines to vaccinate 10% of their populations. A total of US\$19,87 billion in gross domestic product has been lost as a result of these four countries failing to meet the 40% target for vaccination by the year 2021. ¹¹⁶

The COVAX initiative stands out as an international project designed to acquire and distribute vaccines in an equitable manner. However, it has not been able to procure sufficient doses to achieve its modest objective for 2021 of covering 20% of the LICS population. 111,117 For example, Pfizer agreed to sell only 40 million doses to COVAX, and slightly more than 1 million doses had been delivered by mid-May 2021. 117 By the end of May 2021, 34 countries worldwide had contributed doses, and many more had committed to the goals of COVAX. 118 However, LMICs make up 6.5 billion of the world population, and high-income countries have purchased only 5.9 billion doses. 119 As of January 12, 2022, 84 vaccines were in late developmental phases (phase II or III), and another 33 were in post-registration. Of the 28 vaccine manufacturers. Pfizer/BioNTech has made a public commitment to provide the most doses, exceeding 5.1 billion, followed by AstraZeneca, with approximately 3.6 billion doses. 120 Highincome countries have reserved most doses from Pfizer/Bio-NTech (83%) and Moderna (73%), but no bilateral agreements with low-income countries have been revealed. In contrast, AstraZeneca/Oxford, Johnson & Johnson and Novavax have had a larger distribution in LMICs. Vaccines produced in middle-income countries (India, Cuba, China and Russia) are restricted primarily to the country where the vaccine was developed or other middle-income countries. Meanwhile, LICs have conserved relatively smaller quantities of each of these vaccine candidates bilaterally. 120 Therefore, AstraZeneca and the Serum Institute of India have collaborated to develop and supply vaccines to LMICs, in an attempt to partially avert this situation. Several organizations, such the WHO-led COVAX, African Vaccine Acquisition Task Team of African Union Africa in Africa, 121 Serum Institute of India, Bharat Biotech, Premas Bio-tech and Zydus Cadila in India, 122 have been established to work on vaccine development and procurement. 111 In addition, donations could have assisted in vaccination of health professionals and vulnerable communities worldwide if they had been made more quickly and achieved more widespread distribution.113

This disparity between developed and the developing countries in terms of the prospects of achieving a timely solution for the COVID-19 pandemic echoes the commonly observed inverse relationship between income and tendency to contract the disease, ¹²³ given the pervasive outbreaks among low-income communities primarily working in manufacturing and the safety of online jobs available for the higher-income populace. ¹²⁴ Therefore, COVID-19 discourse is expected to increasingly pose important questions that address the principles of socioeconomic disparities related not only to the healthcare industry but also to broader aspects of our lives; it is also likely to provide a platform for

recognizing, addressing and ameliorating the aforementioned disparities.

Vaccine hesitancy and adverse effects

The WHO defined vaccine hesitancy as a reluctance to be vaccinated despite the availability of vaccines. ¹²⁵ In 2019, the WHO designated hesitancy to be vaccinated as one of the ten threats affecting global health. ¹²⁶ Because the success of vaccination programs depends largely on the public's willingness to accept the vaccine, many studies have reported vaccine acceptance and hesitancy rates in various countries. ^{127,128} Furthermore, several research groups have aimed to assess the determinants of, and factors driving, vaccine hesitancy and acceptance. ^{129,130} Notably, many COVID-19 vaccine-associated studies have focused on healthcare workers (HCWs) in the early stages of the pandemic, because many countries prioritized vaccination of HCWs at the frontline of the pandemic response, who helped safeguard the lives of vulnerable populations. ¹³¹

Although vaccine hesitancy and acceptance rates vary among populations, ^{132,133} several studies from different countries have shown that vaccine hesitancy is associated with a central concern regarding the lack of trust in COVID-19 vaccine safety and efficacy, particularly given that the COVID-19 vaccines were developed at an unprecedentedly rapid pace. 132,134,135 In addition, some psychosocial predictors, prior experience preconceptions about vaccines, social media influence and trust in governments have been reported to be comodulators of vaccine hesitancy/acceptance across different populations. Moreover, since early 2021, multiple SARS-CoV-2 variants have emerged with reports of waning vaccine-induced immunity in various countries worldwide, thus leading to a surge in the demand for booster doses of COVID-19 vaccines. Studies have shown that booster immunization acceptance is attributable to the belief that boosters are necessary and efficient, 140 as well as to altruistic reasons, such as family, patient and community health protection. 141-143

In contrast, many post-authorization observational studies have been performed to assess the safety and effectiveness of COVID-19 vaccines. Independent studies not supported by vaccine development corporations have helped enhance public confidence in the safety of COVID-19 vaccines, thereby increasing vaccination rates in targeted populations.¹ Generally, although post-vaccination adverse effects have been reported by most participants, studies have shown that serious safety issues of COVID-19 vaccines are extremely rare, and long-term adverse effects are unlikely. 145-147 The most commonly reported post-vaccination adverse effects are fatigue, chills, dizziness, fever, headache, joint pain and myalgia. 148-151 In addition, post-vaccination adverse effects are mostly resolved within 3 days without medical intervention. 133,152 However, the overall prevalence and severity of post-vaccination adverse effects may vary depending on the type of the vaccine. 133,144,153,154

COVID-19 therapeutics

The Milken Institute's COVID-19 treatment and vaccine tracker¹⁵⁵ lists a total of 309 potential treatments from different

categories, including repurposed drugs and drugs assessed in different stages of development against COVID-19, 195 of which have reached the clinical stage. In addition, five treatments (remdesivir, REGN-COV2, bamlanivimab, olumiant and chloroquine/hydroxychloroquine) have received emergency authorization. The chloroquine/hydroxychloroquine authorization was revoked thereafter, and 31 treatments have reached phase III trials.

This repurposing of available drugs has involved several types of antimalarial drugs (e.g., chloroquine), antimalarial and antibiotic combinations (e.g., hydroxychloroquine and azithromycin), antiviral drugs (e.g., nafamostat, camostat, bromhexine, favipiravir, remdesivir and lopinavir) and antihelminthic/antiparasitic agents (e.g., nitazoxanide and ivermectin), which have shown potential antiviral effects against SARS-CoV-2. ^{156–158} Although prior studies did not identify specific antiviral drugs against SARS-CoV or MERS-CoV, reports have recently suggested that several available drugs may act as antiviral agents against SARS-CoV-2, particularly hydroxychloroquine, chloroquine and nafamostat. ^{159,160} However, the clinical effectiveness of these drugs has not yet been fully evaluated, and several clinical trials remain underway. ¹⁶⁰

Furthermore, other types of repurposed drugs are expected to have regulatory effects on the immunity of patients with COVID-19, particularly those with severe disease. Anticoagulant treatments (e.g., heparin and nafamostat), for example, inhibit the cytokine storm and increase the percentage of lymphocytes. ^{159,161,162} Some immune-based therapies (e.g., tocilizumab and interferon alpha-2B) are also expected to have similar effects but continue to await experimental evaluation. ¹⁵⁶ Other types of drugs have been repurposed and are currently being studied, such as antihypertensive drugs and non-steroidal anti-inflammatory drugs; however, no scientific evidence demonstrating the effectiveness of any drug or therapeutic compound against COVID-19 has been reported to date.

Another proposed therapeutic strategy for COVID-19 is using potent NAbs against SARS-CoV-2. Several studies have shown promising results supporting this strategy. 163–165 One study has reported that two human monoclonal antibodies isolated from a convalescent patient with COVID-19 demonstrated potent SARS-CoV-2-specific neutralization activity in vitro. One of these antibodies showed inhibitory therapeutic and prophylactic activity against SARS-CoV-2 in rhesus monkeys. 163 Structural studies of this antibody have indicated its ability to recognize an epitope that overlaps with ACE2-binding sites in the RBD of SARS-CoV-2 S protein. 163 However, NAbs are not produced exclusively against the RBD of S protein regions. Another study has shown that 19 of 61 monoclonal antibodies isolated from five patients with COVID-19 potently neutralized authentic SARS-CoV-2 in vitro. Epitope mapping has indicated that half of these antibodies were directed against the RBD of S protein, whereas other antibodies were directed against the N-terminal domain of S protein¹⁶⁴; therefore, both the RBD and N-terminal domain are immunogenic regions of S protein. However, further research remains needed before this therapeutic strategy can enter the clinical stage.

Combined and supplementary therapies have been suggested as potential therapeutic approaches. For example, an *in vitro* study has shown that using a combination of zinc

ionophore (pyrithione) with Zn²⁺ potently inhibits SARS-CoV RNA replication in cultured Vero cells as well as SARS-CoV RNA-dependent RNA polymerase (RdRp) elongation. 166 On the basis of the high sequence similarity between SARS-CoV and SARS-CoV-2 (79% at the genome level and 98% at the RdRp level), researchers have hypothesized that the antiviral effects of zinc or the aforementioned combination might be similarly effective against SARS-CoV-2. 167 An interesting approach has suggested that combining zinc with chloroquine or hydroxychloroquine-which inhibit SARS-CoV-2 replication after an increase in the pH of the intracellular vesicles and also inhibit the intracellular virus particle delivery—might provide a promising treatment for COVID-19.168 Chloroquine or hydroxychloroquine may specifically target extracellular zinc and activate its entrance into the intracellular lysosomes, where it interferes with SARS-CoV-2 RNA replication and RdRp elongation activity, thereby enhancing the efficacy of these repurposed drugs. Furthermore, zinc, vitamin D, vitamin C and N-acetylcysteine have been suggested to have a potential role in the prevention and treatment of COVID-19. However, clinical evidence supporting the use of those supplements for these purposes is insufficient. 169,170 Taking over-the-counter doses of these supplements in the absence of a confirmed nutritional deficiency might pose risks to the treatment outcomes. 169 Thus, further research is required before doses above those recommended for these supplements could be approved.

Overall, massive efforts have been made worldwide toward the development of COVID-19 therapeutics, and many clinical trials have aimed at evaluating the effectiveness of repurposed drugs. Given that their chances of reaching very advanced phases are moderate to high, we summarize the potential challenges of repurposed drugs¹⁷¹ in Figure 5.

Convalescent plasma therapy for COVID-19

According to the WHO, most studies have shown that people who have recovered from COVID-19 have antibodies

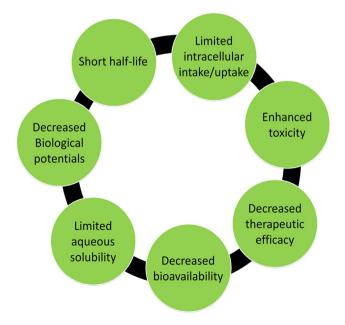


Figure 5: Challenges regarding repurposed drugs against COVID-19.

against the virus; however, some of those people have very low levels of NAbs in their blood.^{6,172} CP is a convalescent blood product obtained from patients who have recovered from a viral infection and have developed humoral immunity against a particular pathogen.¹⁷³ The isolated plasma from the whole blood of recovered patient donors has a high concentration of NAbs against the virus. Hence, the administration of CP to patients with severe SARS-CoV-2 infection might possibly decrease viral replication and disease severity via specific antigen—antibody binding.

In general, CP is a passive antibody therapy that depends on the transfer of pathogen-specific antibodies from a recovered patient to aid in the prevention or treatment of disease. Antibody treatment can also be used to treat individuals already exhibiting symptoms with varying severity. In contrast, passive antibody therapy is most successful when applied prophylactically, or when used immediately after the onset of signs and symptoms. Passive antibody administration through transfusion of CP may be the only short-term method available for providing direct immunity to susceptible patients in a timely manner. Numerous cases have been reported in which CP has been used successfully for treatment and/or postexposure prophylaxis against infectious diseases, including in prior coronavirus outbreaks (i.e., SARS-CoV-1 and MERS-CoV).

The development and implementation of CP therapy is less complicated than that of vaccines and monoclonal antibodies. In addition, CP can adapt to a variety of conditions. The only requirements are the availability of disease survivors who can donate plasma and a standard blood collection infrastructure to collect and distribute the CP. ¹⁷⁵ Individuals who are eligible to donate CP must have a history of COVID-19, as confirmed by approved molecular testing (e.g., nasopharyngeal swab) or the presence of antibodies against SARS-CoV-2. In addition, symptoms including fever, cough and shortness of breath must have resolved at least 14 days before donation of CP.175 In light of the emergence of variant SARS-CoV-2 strains, CP donated by survivors of infections with variant strains is an immediately deployable treatment option for patients who have been identified as having a variant infection, whereas other immune treatments may require (re)development of more specific mechanisms to target new viral strains. 176

Three essential principles of passive antibody therapy must be met by CP. To be effective against SARS-CoV-2 infection: i) CP must contain specific antibodies against the SARS-CoV-2 virus; ii) a sufficient quantity of anti-SARS-CoV-2 antibodies must be produced in the plasma to improve the outcome of the disease; iii) for optimal benefits, the plasma must be administered early after the onset of symptoms. ¹⁷⁹

CP therapy was first recognized as a potential antiviral therapy during the Spanish influenza pandemic approximately 100 years ago. 180 CP administration substantially decreases mortality among patients with severe acute respiratory infections, by 75%. 181 Beyond SARS-CoV, CP therapy has been successful in the treatment of MERS-CoV-2 and H1N1 viral infections. 182 Despite the effectiveness of the CP therapy against previous viral infections, this type of therapy was unable to improve the survival rate during an Ebola virus outbreak (141). Because SARS-CoV-2 shares similar genomic sequences with SARS-CoV and

MERS-CoV, CP therapy can be considered a promising treatment strategy with the potential to help the infected patients develop immediate short-term passive immunization before the availability of a new generation of COVID-19 vaccines. In addition, owing to the rise of different strains of the COVID-19 pathogen arising from genetic modifications with consequences in mutability, CP therapy offers an advantage in providing artificially acquired, passive immunity against the local infections in communities. 173 Recent studies on CP therapy for COVID-19 have reported that CP with high levels of NAb titers from recovered patients with COVID-19 ameliorates clinical symptoms by decreasing viral load, while posing low risk of treatment 183 and no significant transfusion-associated morbidity or mortality. 184 A recent study has found that 15% of 2341 enrolled men and women were less likely to die from COVID-19 within 1 month after receiving an injection of CP than patients who received an inactive saline placebo or did not receive the CP therapy. 185

Moreover, a meta-analysis of international randomized clinical trials (RCTs) among hospitalized, non-critically ill patients with COVID-19 has been conducted to evaluate the effects of CP on patients with COVID-19. The findings were derived from eight recently completed studies conducted in Belgium, the United States, India, Brazil, Spain and the Netherlands. The COVID-19 CP was not consistently associated with either benefits or harms in therapy. 186 In addition, CP was not found to have an effect on clinical outcomes for typical patients. 186 However, patients with type A and AB blood groups, and patients with preexisting diseases such as diabetes or cardiovascular disease were most likely to benefit from the CP therapy. 186 However, another study has reported the benefits of CP therapy for older outpatients within the first 72 h of disease onset. 187 Monoclonal antibody-based therapy has been shown to be helpful, but only for outpatients who have recently been diagnosed with SARS-CoV-2 infection. 188,189 consequence, the lack of a clear benefit of CP even in patients with recent symptom onset has been rather unexpected, thus implying that the window of opportunity for antibody-based therapy may be limited and associated with the stage of illness rather than with symptom onset time. 186

Furthermore, the Continuous Monitoring of Pooled International Trials of Convalescent Plasma (COMPILE) for COVID-19 Hospitalized Patients study has established and validated a CP treatment benefit index. 190 COMPILE included a total of 2369 hospitalized adult patients (18 years of age or older) who had COVID-19 and who might or might not have required oxygen supplementation, but did not require mechanical ventilation. The treatment benefit index score represents the estimated relative advantage that a patient is expected to gain from CP treatment compared with a treatment without CP. 190 Another recent CT study has been conducted on 941 patients who were hospitalized with COVID-19 for 3 or fewer days and required noninvasive oxygen supplementation.¹⁹¹ In this trial, patients receiving large doses of CP who were not taking any other drugs, such as remdesivir or corticosteroids, were found to be likely to benefit from CP treatment. 191 These preliminary findings have reinforced the idea that CP may serve as a viable therapeutic option, particularly when alternative medicines are not yet available. However, the lack of an overall advantage from CP therapy in this trial, ¹⁹¹ as well as other trials on CP^{192–194} in hospitalized patients with severe to life-threatening disease, suggests that patients with less severe disease and older patients may benefit most from CP treatment. Moreover, CP obtained from donors who had been previously infected and vaccinated afterward has been found to possess antibodies with sufficient quantities and diversity to provide further protection against new COVID-19 strains. 191 Genetic mutations are typical of viral evolution during the course of the COVID-19 pandemic. Consequently CP has the potential to provide a successful treatment option that keeps pace with the mutations better than synthetic treatment options, such as monoclonal antibody therapies, which tend to become less effective over time because of mutations and must undergo redevelopment to address the new variants. 185

Several key challenges in developing CP therapy that must be considered. First, plasma transfusions are associated with potential risks of adverse effects. Previously, a patient with Ebola virus disease had developed acute respiratory distress syndrome after plasma transfusion from the recovered patients, possibly because of transfusion-associated acute lung injury. A recent case study on CP for COVID-19 has also reported that a patient developed erythema, hypoxia and hypotensivity with increased dyspnea within 10 min of plasma transfusion. Increased dyspnea within 10 min of plasma transfusion-transmitted diseases. Table 2 shows the characteristics of recent RCTs evaluating CP therapy for SARS-CoV-2 infection.

Herd immunity and COVID-19

When a population is immune to an infectious disease, either through vaccination or immunity acquired through earlier infection, this is referred to as herd immunity or population immunity.⁶ Herd immunity can offer indirect protection against the contraction of infectious disease, particularly for the most vulnerable people, such as those with comorbid conditions, as well as infants and older people. Before vaccination, herd immunity could be achieved only by exposing a very large number of people in the community to the disease in hopes that they would develop protective antibodies. 226 The phenomenon of herd immunity, which occurs naturally, was first observed in the 1930s, after many adolescents had developed immunity to measles.²²⁷ Herd immunity is an important concept for understanding the short- and long-term effects of vaccination programs, and many existing examples illustrate this concept. In one of the most prominent examples, epidemics of common pediatric diseases, such as mumps, pertussis, polio, rubella, measles and chickenpox, occur at regular intervals. ^{228,229} In addition, an indirect herd protection has been documented after the introduction of conjugate vaccines against pneumococcal and Haemophilus infections.²³⁰ Herd immunity is supported by existing examples and historical findings, and has again become the most hotly debated topic in science in the 21st century, in the midst of the SARS-CoV-2 pandemic. Concerns have been raised regarding the effectiveness of herd immunity in minimizing SARS-CoV-2 infection rates among susceptible populations.²³¹

The basic reproduction number, or R_0 (pronounced "R naught"), is the main parameter of the herd immunity threshold. It describes the average number of secondary infections acquired by the introduction of a single infectious individual into a fully susceptible population. ²³² If the population is not immune to the infection, then an infected person can theoretically infect as many as four more people throughout the infectious period if the R_0 is 4. For example, if R_0 is 4, the herd immunity threshold is 0.75, which can be calculated with the formula $1-1/R_0$. ²³³ The interaction between the host and pathogen determines R_0 in a given population, and this value may vary among pathogens. Variables such as population characteristics and the dynamics of disease transmission may affect R_0 . ²³⁴

 R_0 is widely used in the research of infectious disease dynamics, as one of the most fundamental metrics. $^{235-237}$ In most cases, the R_0 for an infectious disease event is given as a single number or low-to-high range, and its interpretation is as follows: if $R_0 > 1$, the outbreak is predicted to continue, and if $R_0 < 1$, it is expected to cease. 232 R_0 can be used to estimate the percentage of a population that must be vaccinated to eliminate an infection; hence, it is commonly used to estimate the potential size of an outbreak or epidemic. 234 Many communicable diseases have reported R_0 values, including Ebola virus disease, measles, influenza, polio, acquired immunodeficiency syndrome (AIDS) and a variety of vector borne infectious diseases. $^{238-240}$

The threshold of herd immunity to alleviate or slow the COVID-19 pandemic depends on the R_0 for SARS-CoV-2. This threshold is described as the proportion of a population immune to an infectious disease, be it as the result of vaccination, natural infection or innate immunity, such that serial transmission of the infectious agent of the communicable disease is precluded or substantially decreased. However, because COVID-19 remains actively spreading, the precise R_0 number is difficult to determine. Therefore, over time, the value of R_0 is inconstant, varying over a wide range of values at different areas in the interconnected world, depending on the outbreak stages and policymakers' strategies to control the infection, which differ across countries and often across regions within a single country.

Initial R₀ predictions for COVID-19 differed among different populations. Most values have ranged from 1.66 to $3.58.^{1,241-243}$ For instance, the first R_0 value was reported in Wuhan, China, to be 2.2.244 More recently, however, the team from the University of Oxford has estimated the R₀ to be 2.63.²⁴⁵ Variants including Alpha, Gamma and Beta have reproductive numbers estimated at 4.7–4.9, 246 whereas the Delta variant has a reproductive number of approximately 5.^{247,248} Preliminary estimates of reproductive numbers of the Omicron variant are 4.2 times greater than those of the Delta variant. 249 Thus, as R₀ values increase, estimates of the herd immunity threshold also increase. Ro is influenced by demographic and social factors, including individual behavior, public health indicators, population density and cultural attitudes, thus leading to wide variations in herd immunity threshold estimations. In addition, the herd immunity threshold formula is dependent on the notion that COVID-19 has the ability to mutate to escape the immune system, and also on the nature of immune responses toward non-systemic respiratory viruses in general, which appear to be transient and only partially effective. 250,251 Thus, the central

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Trial registration number	Study phase	Year	Location	Status	Number of enrolled patients	Patient setting	Primary outcome measure	Mortality rate, planned mortality time point	Plasma titer, assay	References
NCT04385199	3	2021	Italy	Completed	182	Inpatients with supplemental oxygen	Proportion of patients without progression in severity of pulmonary disease, defined as worsening of 2 points in the ordinal scale of WHO within 14 days	16.67%, 28 days	Low titer: ≥1:200 S-RBD IgG	198
CTRI/2020/ 04/024775	2	2020	India	Completed	464	Inpatients with supplemental oxygen	1. Progression to severe ARDS (P/F ratio 100) or 2. All-cause mortality at 28 days	14.01%, 28 days	No minimum cut- off described	199
NCT04356534	N/A	2021	Bahrain	Completed	40	Inpatients with supplemental oxygen	Requirement for invasive ventilation	7.5%, 28 days	No minimum cut- off described	200
NCT04345523	2	2021	Spain	Completed	350	Inpatients with or without supplemental oxygen	Category changes in the "7-Ordinal Scale" ranging from 1 (not hospitalized, no limitations on activities) to 7 (death)	4.9%, 29 days	Confirmed High-titer: >1:80, neutralizing assay	201
NCT04346446	2	2020	India	Completed	29	13.7%, 28 days	Proportion of patients remaining free of mechanical ventilation	Inpatients with supplemental oxygen	No minimum cut- off described	202
CTRI/2020/05/ 025209	2	2021	India	Completed	80	Inpatients with supplemental oxygen	All-cause mortality on day 30 after enrollment and identification of immunological correlates of response to CP		No minimum cut- off described	203
NCT04381858	3	2020	Mexico	Completed	196	Inpatients with supplemental oxygen	1. Mean hospitalization time 2. Mean oxygenation index evolution 3. Rate of severe acute respiratory distress syndrome (ARDS) 4. Rate and time to death 5. Mean time with invasive mechanical ventilation	45.2%, 28 days	No minimum cut- off described	204
NCT04433910	2	2021	Germany	Completed	106	ICU	Composite endpoint of survival and no longer meeting criteria for severe COVID-19	14.3%, 21.0%, and 26.7%, 21, 35 and 60 days, respectively	Confirmed high-titer: ≥1:20 neutralizing antibodies	205
NCT04359810	2	2020	USA and Brazil	Completed	223	Inpatients with or without supplemental oxygen	Day 28 severity outcome	28 days	Low titer: ≥1:400 S-RBD IgG, quantitative ELISA	206

Trial registration number	Study phase	Year	Location	Status	Number of enrolled patients	Patient setting	Primary outcome measure	Mortality rate, planned mortality	Plasma titer, assay	References
								time point	_	
NCT04441424	N/A	2020	Iraq	Completed	49	Inpatients with supplemental oxygen	Death versus survival of treated patients	6.1%, Up to 8 weeks	No minimum cut- off described	207
NCT04383535	N/A	2020	Argentina	Completed	333	Inpatients with or without supplemental oxygen	Ordinal outcome with six mutually exclusive categories describing the patient's clinical status during follow-up (ranging from 1 (death) to 6 (discharge))	11.1%, 30 days	Confirmed high-titer: ≥1:800 S-RBD IgG, COV-IDAR IgG test	208
NCT04392414	2	2020	Russia	Completed	60	Inpatients with or without supplemental oxygen	Number and proportion of patients with normal body temperature (≤37.2 °C) at day 1, 2, 3, 4, 5, 6, 7 after the start of therapy	6.06%, 30 days	Confirmed high-titer: ≥1:1000 S-RBD IgG, in-house assay	209,210
NCT04332835	2-3	2020	Colombia	Completed	92	Inpatients with supplemental oxygen	Change in viral load Change in immunoglobulin G COVID-19 titers	8.0%, 7, 14 and 28 days	Confirmed high-titer: IgG $\geq 1/3200$ and IgA $\geq 1/800$ by EUROIMMUN; all transfused plasma presented neutralizing antibodies $\geq 1/256$	
NCT04366245	2	2020	Spain	Completed	72	Inpatients with supplemental oxygen	 Incidence of adverse events and serious adverse events, grade 3 and 4, associated with the product under investigation or the administration procedure, graduated according to CTCAE. Death from any cause Need for mechanical ventilation SOFA scale ≥3 at 72 h after randomization or an increase in 2 points or more from the basal level 	2.44%, 14 and 28 days	Low titer: Vircell SL (Spain) test, correlates to ≥1:320 S-RBD IgG	209,212

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NCT04362176	3	2021	USA	Completed	1000	Inpatients with or without supplemental oxygen	COVID-19 7-point Ordinal Clinical Progression Outcomes Scale (ranging from 1 (not hospitalized with resumption of normal activities) to 7 (death))	Not defined, 14 and 28 days	50% neutralization titer >1:50 (neutralizing antibodies) are selected for transfusion in the trial, Abbott™ ARCHITECT™ platform and the RBD Luminex assay	213
NCT04323800	2	2021	USA	Completed	180	Outpatients	 Efficacy of treatment at day 28 Safety of treatment with high-titer anti- SARS-CoV-2 plasma versus control-1 and control-2 	0%, 28 days	assay Confirmed hightiter: ≥1:320 neutralizing antibodies, Euroimmun ELISA	214
NCT04364737	2	2021	USA	Active, not recruiting	941	Inpatients with supplemental oxygen	The participant scores on the 11-point WHO Ordinal Scale for Clinical Improvement on day 14 after randomization (range from 0 to 10, with 0 indicating uninfected and no viral RNA detected and 10 indicating death)	4.0%, 14 and 28 days	Confirmed high- titer: ≥1:100 neutralizing antibodies, ELISA	215
NCT04373460	2	2020	USA	Active, not recruiting	1225	Inpatients with or without supplemental oxygen	1. Cumulative incidence of hospitalization or death before hospitalization 2. Cumulative incidence of treatment-associated serious adverse events 3. Cumulative incidence of treatment-associated grade 3 or higher adverse events	0.2%, 28 days	Confirmed high- titer: 1:320 SARS-CoV- 2 spike protein titers, Euroimmun ELISA	16,216
IRCT202003 10046736N1	2-3	2020	Iran	Not yet recruiting	30	Outpatients	Improvement in the levels of cytokine storm indices	26.7%, 7–14 days	No minimum cut- off described	217
NCT04338360	N/A	2020	USA	Approved for marketing	3082	Inpatients with or without supplemental oxygen	Death within 30 days after plasma transfusion	26.9%, 30 days	Confirmed high- titer: ≥1:160 neutralizing antibodies	192
NCT04381936	3	2021	United Kingdom	Recruiting	11558	Inpatients with or without supplemental oxygen	All-cause mortality	24.3%, 28 days	Confirmed high- titer: ≥1:100 neutralizing antibodies	218
									(continued or	next page)

Trial registration number	Study phase	Year	Location	Status	Number of enrolled patients	Patient setting	Primary outcome measure	Mortality rate, planned mortality time point	Plasma titer, assay	References
NCT02735707	3	2021	Worldwide	Recruiting	2011	ICU	All-cause mortality Primary endpoint for patients with suspected or demonstrated SARS-CoV-2 infection	37.2%, 90 days	Confirmed high- titer: ≥1:100 neutralizing or equivalent	219,220
NCT04873414	3	2021	Indonesia	Recruiting	364	Inpatients with supplemental oxygen	Mortality of patients with COVID-19 treated with CP (number of deaths from the initiation of CP treatment until hospital discharge or death)	Not defined, 82 days	No minimum cut- off described	221
NCT04344535	2	2021	USA	Terminated	82	Inpatients with and without supplemental oxygen	1. Number of days of invasive mechanical ventilation through 28 days post randomization 2. Patients who died during this time period were assigned 0 ventilator free days	16.1%, 90 days	Confirmed high- titer: >1: 320, neutralizing assay	222
NCT04342182	3	2020	Netherlands	Terminated	86	Inpatients with or without supplemental oxygen	Comparison of mortality in the 300 ml CP group and the control arm	19.8%, 60 days	Confirmed high- titer: >1:80, neutralizing assay	223
ChiCTR 2000029757	N/A	2020	China	Terminated	103	Inpatients with supplemental oxygen	Patient discharged alive or decrease of 2 points on a 6-point disease severity scale (ranging from 1 (discharge) to 6 (death))	19.4%, 28 days	Confirmed high- titer: ≥1:640 S-RBD IgG, in-house assay	224
NCT04479163	N/A	2020	Argentina	Terminated	160	Outpatients	Development of severe respiratory disease defined as a respiratory rate $>$ 30 and/or an O_2 saturation $<$ 93%	3.8%, 25 days	Confirmed high- titer: >1:1000 against SARS- CoV-2 spike protein (COVIDAR IgG)	225
NCT04348656	3	2021	USA	Terminated	940	Inpatients with supplemental oxygen	Endpoint of the need for intubation or patient death	30.3%, 30 days	Confirmed high- titer: ≥1:100 neutralizing antibodies	178

ELISA, enzyme linked immunosorbent assay; N/A, not applicable; SOFA, sequential organ failure assessment; CTCAE, common toxicity criteria scale.

epidemiological question is what proportion of the population should be infected for herd immunity to be achieved and the community protected from contracting and/or spreading infectious SARS-CoV-2. However, given the relatively high number of people who should be infected and might potentially die from SARS-CoV-2 infection for herd immunity to be achieved, strategies to achieve the acquired immunity via natural COVID-19 infection would be difficult to accomplish. Therefore, the WHO recommends that herd immunity against COVID-19 be obtained through vaccination rather than by exposing people to the virus that causes the disease, to avoid unwanted new cases and increased mortality.⁶

In 2016, the annual number of deaths due to measles was less than 100,000, a number significantly lower than the more than 2 million deaths due to COVID-19 reported during the pre-vaccination period.²⁵² To achieve herd immunity, the WHO has set the limit of 95% of the population that should be immunized, because of the high infectivity of measles.²⁵³ Accordingly, despite the lower mortality rate of COVID-19, its high infectivity requires a high proportion of the population to develop immunity to stop the spread of the infection, thereby reflecting the challenges in creating herd immunity. Deciding on priority groups to receive COVID-19 vaccination is challenging not only because of the limited supply of vaccines with early approval for HCWs, older people, and the middle-aged community, 254 but also because of direct effects on herd immunity. Another issue is the duration of immunity offered by the vaccination program. Because SARS-CoV-2 remains relatively new, the duration of the immunity memory, particularly in terms of epitope stability due to viral mutation, remains a large unknown.²⁵⁵

Both vaccinated people and convalescent patients with COVID-19 have shown lower binding of NAbs, 256-258 and vaccine effectiveness has been lower in regions where the Gamma and Beta strains are more widespread.^{259,260} In addition, Omicron has 32 S protein mutations that decrease NAb titers by 27-127 times relative to that of wild-type COVID-19. 261 The Omicron variant of the coronavirus, which is spreading more quickly than previous variants, is unlikely to help countries achieve herd immunity against COVID-19, wherein individuals would become immune to the virus with decreased spreading rates.²⁶² The Omicron variant's transmissibility is enhanced because it is more effective than its predecessors at infecting people who have been vaccinated or have previously been infected. Growing evidence indicates that the coronavirus will continue to develop new ways to overcome immune responses.²⁶² According to recently published studies, vaccination effectiveness against Omicron-associated symptoms is lower than that for other variants, and it decreases over time. Consequently, more people who have been vaccinated against Omicron are at risk of developing severe disease. 263-265

However, these studies also suggest that vaccination continues to provide substantial protection against severe disease and hospitalization, owing to the variety of diseases caused by the Omicron variant. People who have received a booster dose may be more protected than those who have received only the initial course, as suggested by the most recent evidence, including real-world effectiveness data. In South Africa, individuals who have received two doses of the

Pfizer—BioNTech vaccine, for example, have been found to have as much as 70% protection against hospitalization. ²⁶⁴ Additionally, during the proxy Omicron period, the BNT162b2 vaccine maintained its effectiveness, albeit at a lower level, against hospital admission due to COVID-19 thought to have been caused by the Omicron variant than the Delta variant. The UK surveillance report has also revealed that after a booster dose of either the Pfizer or Moderna vaccine, the vaccination effectiveness against symptomatic disease among Omicron-infected patients was 60%—75% in the first 2–4 weeks, after which it decreased to 25%—40% in the next 15+ weeks. ²⁶⁵

Despite the high infection rates, the two-dose COVID-19 vaccine strategy has significantly decreased hospitalizations and deaths. ^{266,267} Similarly to other vaccines, the second dose loses some of its protective efficacy within several months. ^{268,269} Previous studies have shown that, as any with other vaccine, COVID-19 vaccines show a decline in the levels of binding and NAbs with time. 268-270 A preprint has indicated that at least 20 weeks after the second dose of the SARS-CoV-2 vaccine, the efficiency of ChAdOx1 nCoV-19 (Oxford-AstraZeneca) was 44.1%, and that of BNT162b2 (Pfizer-BioNtech) was 62.5%. 271,272 In Qatar, researchers have observed similar findings, with the exception that the effectiveness against hospitalization and death remained strong even after 6 months.^{273–275} Similarly, a large study conducted in the USA has found that the risk of infection significantly increases 6 months after vaccination, and mRNA-based vaccines have a substantially lower increase in risk than the viral vector-based vaccine Ad.26.COV2.S (Janssen).²⁷⁶ Booster doses of BNT162b2 after vaccination have been shown in two Israeli studies to increase protection against symptomatic infection by 93.1%. 277,278 Both humoral and cellular responses to SARS-CoV-2 have been reported to be enhanced by booster doses in the COV-BOOST randomized controlled trial in the UK, with adverse effects similar to those seen after initial vaccination.²⁷⁹ Moreover, the effectiveness against infection for both mRNA (mRNA-1273 [Moderna] and BNT162b2) and viral vector (ChAdOx1 nCov-19) COVID-19 vaccines has been shown to dramatically decline over 5-8 months, with respect to that 1 month after the second dose. 280 Moreover, a booster dose administered 6 months after the second vaccination has been found to restore the vaccine's efficacy to levels greater than those reported 1 month after the second vaccination. Those who received a heterologous booster schedule experienced a higher rate of systemic reactogenicity after vaccination than those who received a homologous booster, although both groups experienced very mild adverse effects after the booster vaccination.²⁸⁰ Moreover, according to a study conducted in Canada, the effectiveness of vaccines in vaccinated individuals (compared with unvaccinated individuals) increased with each additional dose, and provided 49% (95% confidence interval 43%-54%) protection against infection, 69% (61%-76%) protection against symptomatic infection and 86% (81%-90%) protection against severe outcomes after the fourth dose. 281 During the Omicron dominant era, results have suggested that a fourth dose of the mRNA COVID-19 vaccine (mRNA-1273) provides better protection against infection, symptomatic infection and severe outcomes for people living in long-term care centers.²⁸¹

According to the available findings, vaccine boosters are safe for the general public and effective at restoring immunity after a period of waning protection. Homologous booster schedules have more favorable safety profiles than heterologous booster schedules.

The efficacy of COVID-19 vaccines against asymptomatic infection may still be associated with transmission and is generally less efficient and may be less adequately estimated than that against symptomatic infection. 282,283 In a retrospective study conducted on HCWs who received the Pfizer vaccine, the vaccine efficiency for avoiding symptomatic infection was 97%, but was only 86% for avoiding asymptomatic disease.²⁸⁴ The efficacy of the Moderna vaccine against asymptomatic disease was only 63%, as compared with 93.2% against symptomatic infection, whereas the efficacy of the AstraZeneca vaccine with the original dose regimen was only 3.8%. 260,285 Therefore, "universal vaccinations" as vaccine regimens that generate longer-lasting immunity and cover a wider range of potential future mutations may help decrease some of the problems associated with herd immunity against respiratory viruses. However, this type of vaccine remains under investigation.²⁸⁶

Herd immunity against COVID-19 can be safely achieved only by vaccinating large numbers of people, thereby decreasing overall viral transmission in the entire population. On the basis of the opinion of many experts, increasing evidence suggests that 80%-90% of the population would be required to have COVID-19 immunity for herd immunity to be effective. ²⁸⁷ Vaccinations and prior infection will help enhance herd immunity against COVID-19, thus decreasing the severity of the disease for people who are infected or reinfected.²⁶² However, to determine vaccine effectiveness against various outcomes (such as severe disease and subsequent transmission), vaccine effectiveness in various population subgroups and against various variants, as well as vaccine duration of protection, real-world vaccine effectiveness studies after implementation are required. 265 Herd immunity threshold estimates must therefore consider not only partial vaccine effectiveness, but also changes in critical parameters associated with mutations and with dynamic and steadily declining immunity. Table 3 shows recent RCTs that have indicated assessment of efficacy of candidate vaccines against SARS-CoV-2 infection.

SARS-CoV-2 reinfection

One of the largest enigmas surrounding COVID-19 is the probability of reinfection and/or reactivation of latent viral hideouts in the body, as well as the potential negative effects of some drugs on the formation of acquired immunity to the virus. These questions can be considered central for biomedical scientists fighting the outbreak of SARS-CoV-2. Until nearly the third month of the COVID-19 pandemic, scientists expected that the disease would be non-relapsing and would be eliminated through immunization, such that all recovered patients would gain sufficient acquired immunity to SARS-CoV-2. However, as the number of patients with COVID-19 increased, some recovered patients reported relapse of COVID-19 signs and symptoms after they were completely cured and discharged from

hospitals after displaying repeatedly positive SARS-CoV-2 RT-PCR tests. Hence, the dilemma of COVID-19 reinfection versus acquired immunity to SARS-CoV-2 has been raised and requires further research.

The first report showing a positive SARS-CoV-2 RT-PCR assay for a convalescent case (after two consecutive negative results) was from China. 289 Another case report described three patients with COVID-19 who were readmitted in March 2020 to a hospital in Wuhan, China, after having recovered with positive SARS-CoV-2 IgG antibody, negative SARS-CoV-2 RT-PCR and negative SARS-CoV-2 IgM antibody tests.²⁹⁰ Those patients were readmitted because of positive SARS-CoV-2 RT-PCR results after convalescence, although they did not display any of the characteristic COVID-19 signs or symptoms again, and the results of their IgG and IgM tests remained stable. 290 Thereafter, in April 2020, a total of 116 COVID-19 survivors from South Korea tested positive with SARS-CoV-2 RT-PCR tests after recovery.²⁹¹ These positive tests were confirmed according to the Korean criteria, wherein patients must have two consecutive SARS-CoV-2 negative samples within 24 h to be considered to have recovered from COVID-19. ²⁹² In these two reports, the possibility of reinfection was excluded because of the absence of evidence that the detected SARS-CoV-2 RNA was from replication-competent viruses that could be contagious.

Moreover, a case report has described 11 patients with COVID-19 from France in April and May 2020, who experienced second separate symptomatic SARS-CoV-2 infections after recovery, as confirmed by RT-PCR. 288 In May 2020, another case of possible COVID-19 reinfection emerged in the USA. In that case, 1 week after confirmed recovery and discharge, the patient was readmitted with both radiologically and virologically (RT-PCR) confirmed reinfection. Although two similar cases have been reported in South Korea, 293 none of these reports have confirmed whether the convalescence was due to viral reactivation from a hideout in the body or to viral reinfection.

In a retrospective study of 55 patients with COVID-19 who had been admitted to a hospital in Wuhan, five were readmitted after recovery and discharge. ²⁹⁴ All five patients returned with confirmed SARS-CoV-2 infection, as diagnosed on the basis of clinical, laboratory and CT examinations after 4–17 days; the authors hypothesized that SARS-CoV-2 reactivation was the reason. ²⁹⁴ The first SARS-CoV-2 reactivation case in Italy was reported 2 weeks after primary infection; the patient was cured, as confirmed by radiology and two consecutive RT-PCR tests. Interestingly, the results indicated the presence of SARS-CoV-2 IgG antibodies in the patient's serum, but not IgM, thus revealing that his immune system was beyond the acute phase of COVID-19. ²⁹⁵

To investigate the scope of acquired immunity to SARS-CoV-2, another early study conducted by researchers from China infected Rhesus macaques with SARS-CoV-2, then reinfected them after confirmed recovery. ²⁹⁶ Viral replication was detected (nasopharyngeal and anal swabs), and COVID-19 symptoms were recorded in the primary infection, and neutralizing activity against SARS-CoV-2 was confirmed after recovery. However, the viral replication and signs of COVID-19 were not detected after reinfection, thus suggesting that acquired immunity to SARS-CoV-2 may protect against reinfection. ²⁹⁶ However, how long the acquired

Vaccine candidate	Developers	Doses	Trial phase	Clinical trial numbers	Efficacy of vaccine against hospitalization for severe and non-severe SARS-CoV-2 infection	Efficacy of vaccine against emergency department visits associated with SARS-CoV-2 infection	Efficacy of vaccine against severe and non-severe SARS-CoV-2 infection	Seroconversion rates (efficacy)	Immunogenicity and safety of a booster dose
CoronaVac;	Sinovac Research	2	Phase 4	NCT04756830	_	-	~	~	~
inactivated	and Development			NCT04789356					
SARS-CoV-2	Co., Ltd			NCT04801888					
vaccine				NCT04894227					
(Vero cells)				NCT04892459	/				
				NCT04953325					
				NCT04962308				/	
				NCT05057169					
				NCT05148949 NCT05112913				✓	✓
				NCT05112915 NCT05165966					
				TCTR20210308003					
nactivated	Sinopharm;	2	Phase 4	NCT05065892			∠	∠	<u></u>
SARS-CoV-2	China National	-	Thuse T	NCT05075044			<u></u>	<u> </u>	<u></u>
vaccine	Biotec Group Co;			NCT05075057				✓	∠
(Vero cells)	Wuhan Institute of			NCT04790851			✓	✓	
WIBP	Biological Products			NCT04863638			✓	✓	∠
COVID-19				NCT05065879					
vaccine				NCT05075083					
ChAdOx1-S	AstraZeneca +	1-2	Phase 4	NCT04760132					
(AZD1222)	University of			NCT04775069					
Covishield	Oxford			EUCTR2021-					
Vaxzevria -				002327-38-NL			_		_
				NCT04914832					
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				RBR-4vmh6xg			∠	∠	~
				EUCTR2021-					
				004419-14-FI					
			phase 3	NCT04864561				✓	∠
			F	NCT04756271					<u></u>
				NCT04536051	✓		/	✓	/
Recombinant	CanSino Biological	1	Phase 4	NCT04892459			/	/	
novel coronavirus vaccine (adenovirus type 5 vector)	Inc./Beijing Institute of Biotechnology			NCT05303584				∠	-
AG HCO V		1	Phase 4	NCT05303584	✓			∠	✓
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Vaccine candidate	Developers	Doses	Trial	Clinical trial	Efficacy of	Efficacy of	Efficacy of	Seroconversion	Immunogenicity
			phase	numbers	vaccine against hospitalization for severe and non-severe SARS-CoV-2 infection	vaccine against emergency department visits associated with SARS-CoV-2 infection	vaccine against severe and non-severe SARS-CoV-2 infection	rates (efficacy)	and safety of a booster dose
Recombinant COVID-19 vaccine (adenovirus type 5 vector) for inhalation (Ad5-nCoV-IH)	CanSino Biological Inc./Beijing Institute of Biotechnology		Phase 3	NCT05169008 NCT05204589	moction	meetion	meedon	<i>V</i>	V V
Gam-COVID-Vac Adeno-based (rAd26-S + rAd5-S); Sputnik V COVID-19 vaccine	Gamaleya Research Institute; Health Ministry of the Russian Federation		Phase 3	NCT04530396 NCT04564716 CTRI/2022/04/041792 NCT04642339 NCT04656613 NCT04741061 PACTR2021046 01572565	~		V		
Ad26.COV2.S	Janssen Pharmaceutical (Johnson & Johnson)	1-2	Phase 4 Phase 3	EUCTR2021- 002327-38-NL NCT05037266 NCT05075538 NCT04614948 NCT05047640 NCT05091307 NCT05142319 PACTR202102 855526180					
SARS-CoV-2 rS/matrix M1- adjuvant; full length recombinant SARS CoV-2 glycoprotein nanoparticle vaccine adjuvanted with matrix M; NVX-CoV2373	Novavax	2	Phase 3	NCT04611802 NCT05463068 EUCTR2020- 004123-16-GB CTRI/2022/04/042017 NCT05556720 NCT05249816 NCT05372588 NCT04583995	∠	1 -			
mRNA-1273; Spikevax	Moderna + National Institute	2	Phase 4	NCT04760132 NCT04885907			~		✓

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	(NIAID)			EUCTR2021- 003388-90-NL					
				EUCTR2021-				∠	✓
				001202-30-NL					
				NCT04952402 EUCTR2021-	/	12	<i>1</i> /	✓	
				003618-37-NO					
				NCT05030974					/
				NCT05047718					
				NCT05079633				/	/
				NCT05075538 EUCTR2021-004419-14-				// //	
				FI					
			Phase 3	NCT05387317			~	~	/
BNT162b2	,	2	Phase 4	NCT04760132					
(3 LNP-mRNAs),	BioNTech + Fosun			NCT04780659					/
also known as Comirnaty	Pharma			NCT04775069 EUCTR2021-000893-27-		/	∠		
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				BE					
				RBR-4vmh6xg NCT04852861			<i>1</i> /		<u> </u>
				NCT04878211				∠	
				EUCTR2021-003618-37-	~			~	
				NO					
				NCT04952766 NCT04969250					<u> </u>
				NCT05057169				∠	
				NCT05160766	/			~	/
			Phase 3	NCT05405283					/
Recombinant SARS-CoV-2		2-3	Phase 3	NCT05091411			✓	// //	
vaccine	Longcom Biologic Pharmacy Co., Ltd.			NCT04646590 NCT05128643					
(CHO Cell);									
Zifivax (ZF2001)				7.0m2.1/m1.1-					
CVnCoV vaccine	CureVac AG	2	Phase 3	NCT04674189 NCT04838847				// //	<u> </u>
				NCT04838847 NCT04860258					
SARS-CoV-2	Institute of Medical	2	Phase 3	NCT04659239				1	
vaccine	Biology + Chinese			NCT05163652					
(Vero cells)	Academy of			NCT05033847					
QazCovid-in®;	Medical Sciences Research Institute	2	Phase 3	NCT05164731 NCT04691908				✓	<i>V</i>
COVID-19	for Biological Safety		1 11450 3	110104071700					
inactivated	Problems, Rep of								
vaccine	Kazakhstan								
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Table 3 (continued)									
Vaccine candidate	Developers	Doses	Trial phase	Clinical trial numbers	Efficacy of vaccine against hospitalization for severe and non-severe SARS-CoV-2 infection	Efficacy of vaccine against emergency department visits associated with SARS-CoV-2 infection	Efficacy of vaccine against severe and non-severe SARS-CoV-2 infection	Seroconversion rates (efficacy)	Immunogenicity and safety of a booster dose
INO-4800+ electroporation	Inovio Pharmaceuticals + International Vaccine Institute + Advaccine (Suzhou) Biopharmaceutical Co., Ltd	2	Phase 3	ISRCTN15779782			<i>V</i>	<i>P</i>	~
AG0301-COVID19	AnGes + Takara Bio + Osaka University	2	Phase 2/3	NCT04655625				/	
nCov vaccine	Zydus Cadila	3	Phase 3	CTRI/2020/07/026352				∠	/
BBV152 vaccine; COVAXIN	Bharat Biotech International Limited	2	Phase 3	NCT04641481				~	/
VAT00008:	Sanofi	2	Phase 3	PACTR202011523101903			/		
SARS-CoV-2	Pasteur + GSK			NCT05124171					
S protein with adjuvant				NCT05405283 NCT04904549	∠		∠		✓
Inactivated SARS-CoV-2 vaccine (Vero cells)	Shenzhen Kangtai Biological Products Co., Ltd.	2	Phase 3	NCT04852705			1	1	
GRAd-COV2; replication defective simian adenovirus (GRAd) encoding S	ReiThera + Leukocare + Univercells	1	Phase 2/3	NCT04791423		"	~	1	~
CpG 1018/alum- adjuvanted recombinant SARS- CoV-2 trimeric S- protein subunit vaccine (SCB-2019)	, ,	2	Phase 3	NCT05012787	<i>ν</i>		V	∠	~
MVC-COV1901;	~	2	Phase 4	NCT05079633					
spike-2P	Biologics +		Db 2	NCT05097053					
protein + adjuvant CpG 1018; MVC- COV1901(Beta)	Dynavax + National Institute of Allergy and Infectious Diseases		Phase 3	NCT05198596				1	1
FINLAY-FR-2 anti- SARS-CoV-2 vaccing (RBD chemically conjugated to tetanus toxoid plus adjuvant Soberana 02	s	2	Phase 3	RPCEC00000354			Let	∠	~

the prevention of	Federal Budgetary Research Institution State Research Center of Virology and Biotechnology		Phase 3	NCT04780035	1	1	~	/	▶
production expressed in Sf9 cells); recombinant SARS-	$\begin{aligned} & Hospital + Sichuan \\ & University \end{aligned}$	2	Phase 3	NCT04887207 NCT04904471	"		"	"	- -
UB-612 (Multitope peptide based S1- RBD-protein based vaccine)	Vaxxinity	2	Phase 3	NCT05293665			"		/
DelNS1-2019-nCoV- RBD-OPT1 (Intranasal flu-based- RBD)	University of Hong Kong, Xiamen University and Beijing Wantai Biological Pharmacy	2	Phase 3	ChiCTR2100051391 PACTR202110872285345	5		V		
SARS-CoV-2 mRNA vaccine (ARCoV)	•	2	Phase 3	NCT04847102			~		
Coronavirus-like particle COVID-19 (CoVLP); MT-2766	Medicago Inc.	2	Phase 2/3 phase 3	NCT04636697 NCT05040789			~	1	∠
COVI-VAC	Codagenix/Serum Institute of India	1-2	Phase 3	ISRCTN15779782	~			~	~
CIGB-66 (RBD + aluminum hydroxide)	Center for Genetic Engineering and Biotechnology (CIGB)	3	Phase 3	RPCEC00000359	~		~	~	~
VLA2001	Valneva + National Institute for Health Research, UK	2	Phase 3	NCT04864561 NCT04956224				1	<u> </u>
BECOV2	/	2	Phase 3	CTRI/2021/08/036074				~	~
Recombinant SARS- CoV-2 spike protein, aluminum adjuvanted (Nanocovax)	Nanogen	2	Phase 3	NCT04922788			1	(conti	nued on next page)

Table 3 (continued)									
Vaccine candidate	Developers	Doses	Trial phase	Clinical trial numbers	Efficacy of vaccine against hospitalization for severe and non-severe SARS-CoV-2 infection	Efficacy of vaccine against emergency department visits associated with SARS-CoV-2 infection	Efficacy of vaccine against severe and non-severe SARS-CoV-2 infection	Seroconversion rates (efficacy)	Immunogenicity and safety of a booster dose
Recombinant protein vaccine S-268019 (using baculovirus expression vector system)	Shionogi	2	Phase 3	NCT05212948 JPRN-jRCT2031210613			V		/
TURKOVAC; inactivated virus	Erciyes University and the Health Institutes of Turkey (TUSEB)		Phase 3	NCT04942405 NCT05077176				~	V
GBP510, a recombinant surface protein vaccine with adjuvant AS03 (aluminum hydroxide)	SK Bioscience Co., Ltd. and CEPI	2	Phase 3	NCT05007951				∠	~
Razi Cov Pars, recombinant spike protein	Razi Vaccine and Serum Research Institute	3	Phase 3	IRCT20210206050259N3			/		/
COVID-19 inactivated vaccine, (CovIran- Barkat)	Shifa Pharmed Industrial Co	2	Phase 2/3	IRCT20201202049567N3			~	~	/
ReCOV: recombinant two-component spike and RBD protein COVID-19 vaccine (CHO cells)		2	Phase 3	NCT05398848			~	/	~
ABNCoV2 capsid virus-like particle (cVLP) ± adjuvant MF59	Radboud University	2	Phase 3	NCT05329220				"	V
Recombinant SARS- CoV-2 fusion protein vaccine (V-01)	Livzon Pharmaceutical	2	Phase 3	NCT05096832 NCT05096845	<u></u>		<i>V</i>		₽
Inactivated COVID-19 vaccine	KM Biologics Co., Ltd.	2	Phase 3	JRCT2031210679					

RBD protein recombinant SARS- CoV-2 vaccine (Noora vaccine)	U	3 Phase 3	IRCT202106200516	39N3				V
Recombinant protein RBD fusion dimer adjuvanted vaccine; COVID-19 vaccine Hipra; PHH-1V	Laboratorios Hipra, 2 S.A.	Phase 3	NCT05246137 NCT05303402	100	1		7	1
ARCT-154 mRNA		2 Phase 3	ISRCTN15779782	~			~	
Vaccine SCTV01C; bivalent recombinant trimeric S protein vaccine against SARS-CoV-2 variants		Phase 3	NCT05308576 NCT05323461			le ^a	~	14 14
Inactivated whole virion concentrated purified vaccine (CoviVac)	Chumakov Federal 2 Scientific Center for Research and Development of Immune-and- Biological Products	Phase 3	NCT05407142			V		

NCT, clinicaltrials.gov (https://clinicaltrials.gov/ct2/home); IRCT, Iranian Registry of Clinical Trials (https://en.irct.ir/); ChiCTR, Chinese Clinical Trial Registry (https://www.chictr.org.cn/searchprojen.aspx); EUCTR, European Union Clinical Trials Registre (https://www.clinicaltrialsregister.eu/ctr-search/search); ANZCTR, Australian New Zealand Clinical Trials Registry (https://anzctr.org.au/Default.aspx); PACTR, Pan African Clinical Trials Registry (https://pactr.samrc.ac.za/Search.aspx); ISRCTN, International Standard Randomized Controlled (https://www.isrctn.com); CTRI, Clinical Trials Registry-India (https://ctri.nic.in/Clinicaltrials/login.php); RPCEC, Cuban Public Registry of Clinical Trials (https://rctportal.niph.go.jp/en/link); JRCT, Japan Registry of Clinical Trials (https://jrct.niph.go.jp/); RBR, Brazilian Registry of Clinical Trials (https://ensaiosclinicos.gov.br/).

immunity to SARS-CoV-2 lasts is uncertain. A study on individuals infected with COVID-19, for example, has indicated a consistent decrease in neutralizing SARS-CoV-2 antibody levels over a 2–3 month period following recovery. ²⁹⁷

These early studies have clarified that reinfection with SARS-CoV-2 is a realistic scenario and not a mere possibility. According to a recent study in Tunisia, reinfection has been detected in four women and HCWs 32–46 years of age. Whole-genome sequencing of the viral RNA was performed on the clinical specimens that retested positive for SARS-CoV-2 by RT-PCR after recovery from the first infection and also on samples collected during the initial infection. A reinfection episode was detected in all four patients. Through comparative genome analysis, unique viral lineages were detected among the four patients. The duration between infection episodes ranged between 45 and 141 days, and symptoms were milder in two patients

during the second infection and more severe in the other two patients. Despite the presence of antibodies in three of the patients, all four patients had reinfection.⁸

However, Lumley and colleagues have reported no symptomatic reinfections in HCWs who had anti-spike antibodies, thus indicating that previous SARS-CoV-2 infection may result in anti-spike antibodies that protect against reinfection in most people, at least for the first 6 months. Recently, in contrast to that after two rounds of vaccination, the IgG response in COVID-19 infected people has been demonstrated to be characterized by insufficient avidity maturation. Insufficient avidity maturation may promote SARS-CoV-2 reinfection and consequently impede the development of herd immunity in susceptible individuals. Remarkably, the conjunction of natural infection and one step of vaccination resulted in the same degree of high IgG avidity as two rounds of vaccination, hence increasing the protective immune response. 300

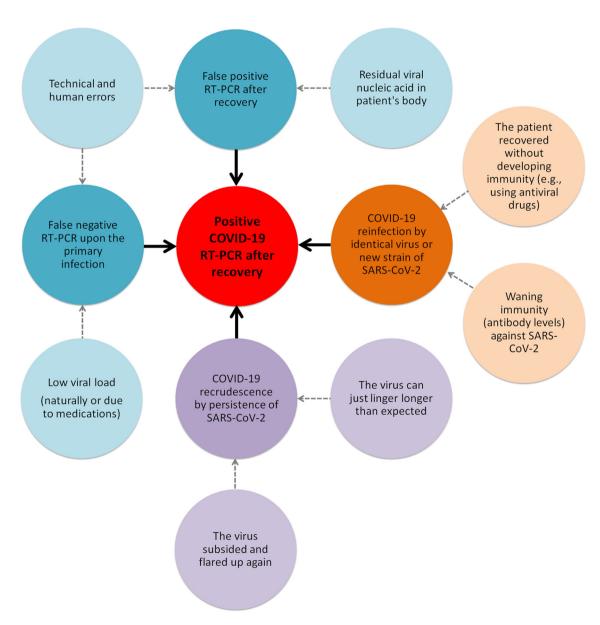


Figure 6: Illustration of potential explanations for positive SARS-CoV-2 RT-PCR tests in recovered patients with COVID-19.

According to these findings, individuals who have been exposed to different COVID-19 strains may not be able to develop sufficiently protective immune responses through natural infection, thus highlighting the importance of vaccination and regular monitoring of their immune status, both qualitatively and quantitatively.

A prospective cohort study has recently been conducted among staff at the International Centre for Diarrheal Disease Research in Bangladesh, who had been registered for COVID-19 testing by real-time RT-PCR. In this study, 750 positive patients were identified to have reinfection, 27.6% of whom required hospitalization and 2.5% of whom died. Among the positive cohort, 731 patients, excluding those who died, who tested positive for COVID-19 (episode 1) were followed up for 0.91 person-years, and 38 (5.7 reinfections/100 person-year) were identified as having reinfection (episode 2). The probability of infection was 80.2% lower in the positive cohort than the negative cohort. Genome sequencing revealed that reinfections were caused by genetically different COVID-19 strains. Naturally infected populations had a lower risk of reinfection with COVID-19 than vaccinated and infection-naive groups. Additionally, the lower mean Ct-value detected among symptomatic patients than asymptomatic patients suggested that the development of COVID-19-like symptoms and seroconversion were due to either a low Ct value or a high viral load. Although reinfected patients did not develop severe disease, a substantial number of naturally infected or vaccinated people were more likely to be reinfected by emerging strains.

Moreover, another prospective cohort study has reported that among 2625 patients who had at least one SARS-CoV-2 infection across the 10-month study period, 5.94% experienced reinfection, and 20.57% experienced recurrence following the first infection.³⁰¹ The average number of days to reinfection was 126.50 days, whereas the average number of days to recurrence was 31.50 days. The incidence rate of reinfection and recurrence was 0.35 cases/ 1000 person-days and 1.47 cases/1000 person-days, respectively, and most recurrences was detected within 30 and 60 days after the first infection. Moreover, people working in clinical units were at greater risk of reinfection than those working in non-clinical facilities. 301 This study contributes to the growing consensus that COVID-19 reinfection, defined as second infection >90 days after initial infection, is uncommon, even among a HCWs who are regularly exposed to the virus. In addition, this study has emphasized the urgent need for widespread vaccination and booster doses to alleviate public health costs.

Although the current vaccines have been demonstrated to be less effective against one or more variants than the original strain, studies have shown that full vaccination is reasonably effective in preventing COVID-19 associated hospitalization and death among vaccinated people compared with non-vaccinated people. ^{7,260,302–306} Studies have demonstrated that Pfizer-BioNTech or Moderna vaccination effectively provides cross-variant neutralization against both the Beta (B.1.351) and Alpha variants following a natural infection by boosting the responses of neutralizing antibody elicited by COVID-19 variants. ^{307,308} According to these findings, complete vaccination against COVID-19 confers additional

protection for people with a previous SARS-CoV-2 infection. In contrast, the molecular evidence for reinfection has indicated that genetically different strains or novel variants can evade the immune response regardless of whether they were acquired from vaccination or natural infection. ³⁰⁹ In a study by Tan et al., all patients, even those with minimal symptoms, exhibited a cellular response to COVID-19 antigens, thus providing cause for optimism in terms of preventing reinfections. ³¹⁰ In contrast, some authors have postulated mechanisms explaining severe second reinfections, including a larger viral load and acquiring a more pathogenic variant. ³¹¹

In addition, several studies have found lower vaccination effectiveness against the Delta (B.1.617.2) variant than against the Alpha (B.1.1.7) or Wuhan (B.1) variants. 41,312,313 One of these studies, conducted in Massachusetts, has reported that more than 74% of 469 patients who were predominantly affected by the Delta variant had been fully vaccinated. Therefore, full understanding of immunity against SARS-CoV-2 infection is urgently needed, to determine the likelihood of second infection and the longevity of vaccine protection. Consequently, assessing the vaccine's effectiveness against newly emerging strains will be critical.

At this point in the discussion, an etymological clarification of the terms "reinfection" and "recrudescence/reactivation" should be made. These two terms have similar outcomes (i.e., viral infection and outbreak) but differ in the origin/cause. In the case of reinfection, recovered patients are susceptible to infection again by an identical or a new strain of the virus. In contrast, in reactivation, although they are immunologically protected, recovered patients are susceptible to the reappearance of disease symptoms due to the persistence of the virus. To further understand the underlying mechanisms in cases of COVID-19 reinfection or reactivation, researchers have highlighted the importance of the assessment of adaptive immunity to SARS-CoV-2, and of the genomic comparison of SARS-CoV-2 strains in both episodes. ²⁸⁸

In general, the occurrence of SARS-CoV-2 reactivation has been suggested to depend on three key risk factors: health status of the host, virological factors (e.g., the viral load and the mutation rate) and whether the host is taking an immunosuppressive medication. Notably, the false-negative RT-PCR results can be obtained due to a low viral load and technical or human errors. False-positive results after recovery can also occur because of the recurrence of SARS-CoV-2 RNA fragments as a result of the detoxification process, or because of human or technical errors during sampling, transport, analysis and recording. The surface of the sample of the sampl

In contrast, because the duration of RT-PCR positivity may vary among patients, ³¹⁷ viral replication may persist for longer periods than expected, thus resulting in late reactivation. Questions regarding whether SARS-CoV-2 can remain active for longer periods than expected, or can subside and reactivate again, lack clear answers, thus increasing the possibility of silent carrier status development. ³¹⁶ Therefore, patients with COVID-19 must crucially be followed during the convalescence period, with a commitment of 14 days quarantine post-discharge. These cases highlight the importance of continuous surveillance of the results of SARS-CoV-2 molecular tests (i.e., quantitative RT-PCR) for the possibility of reinfection and infectivity

assessment in patients at convalescence. In addition, the probability of reinfection by the same strain or another genetically mutated strain should be considered.

In addition to the paucity of reports addressing the reinfection dilemma, these reports did not perform crucial comparisons between the genome sequences of SARS-CoV-2 in primary infection and reinfection, and thus could not confirm whether the detected cases of convalescence were due to reinfection or reactivation. Because RT-PCR detects the virus on the basis of a specific small sequence of the viral genome rather than its whole sequence, the test cannot differentiate between reactivation and reinfection, nor can they indicate which strain has caused the reinfection. With this in mind, researchers from the University of Hong Kong have recently followed a case of a confirmed second episode of SARS-CoV-2 infection after more than 4 months of recovery³¹⁸ and performed a genomic comparison of SARS-CoV-2 genome sequences in both episodes. Interestingly, the genome sequence of the SARS-CoV-2 in the second episode of infection significantly differed from that in the first episode.³¹⁸ Therefore, this report has suggested that COVID-19 convalescence is due to reinfection, possibly because SARS-CoV-2 is a positive-sense RNA virus with a comparatively high mutation rate. In the environment, the variability of SARS-CoV-2 is likely to resemble that of the common cold, another RNA virus in constant circulation. Although SARS-CoV-2 may share the high transmissibility and high asymptomatic rates of its relatively benign genetic relative, it is bound to have a far higher fatality rate than the

According to the data available to date, the potential explanations for the positive SARS-CoV-2 RT-PCR results in COVID-19 survivors can be summarized as shown in Figure 6.

Future directions

As SARS-CoV-2 variants have continued to evolve and spread, the risk of COVID-19 reinfection has grown. Hence, caution should be exercised even after confirmed recovery, particularly in older people who are more susceptible to severe COVID-19, and for those with preexisting chronic diseases and/or pneumonia. To ensure a complete cure and prevent recurrence, post-recovery monitoring and stratified management should be implemented whenever possible. Reports on COVID-19 reinfection are limited, owing to the negligible number of these reports and their small sample sizes. Given the global emergency caused by this pandemic, comprehensive studies involving further immunological, virological and epidemiological data are needed to provide further understanding of the underlying pathophysiology of SARS-CoV-2 reinfection.

Owing to the dramatic increase in the number of both active and recovered cases globally, the issue of COVID-19 reinfection has been brought to the public's attention as a necessary factor to consider to properly predict the spread of SARS-CoV-2 and its expected pandemic trajectory. On the basis of the literature discussed above, both herd immunity and vaccines may not be able to eliminate COVID-19 and establish permanent immunity against SARS-CoV-2. Given the cost and time required to develop herd immunity and

vaccines, scientists must seek further valuable and sustainable solutions requiring less time and cost rather than being satisfied with the current approaches.

Conclusions

The current approval of several vaccines for COVID-19 has given the global population hope that the pandemic will be overcome. The urgency of the pandemic, the technological advances and existing vaccine candidates have contributed to the rapid development of vaccines for COVID-19. Nevertheless, much about COVID-19 and its causative agent, SARS-CoV-2, remains undiscovered, including the human response to vaccines and the emergence of reinfection or reactivation of infection, thus further complicating understanding of COVID-19. Hence, healthcare practices and the scientific community should be made aware of these issues for devising guidelines and future R&D plans targeted to the eradication of the COVID-19 pandemic.

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Conflict of interest

The authors have no conflict of interest to declare.

Ethical approval

Not applicable.

Authors contributions

MAIA-H and RM conceived and designed the study, and conducted the preliminary review of articles. MAIA-H, MAHA, MHM-Z and ENSEAR determined the research scope, and collected and organized the extracted data. MAIA-H and MAHA analyzed and interpreted the data. MAIA-H, MAHA, MHM-Z, ENSEAR, MMH, WA, SA, CYY, IZA, VU and RM wrote the initial and final drafts of the article, and provided logistic support. All authors have critically reviewed and approved the final draft and are responsible for the content and similarity index of the manuscript.

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References

 Guan W-J, Ni Z-Y, Hu Y, Liang W-H, Ou C-Q, He J-X, et al. Clinical characteristics of coronavirus disease 2019 in China.

- N Engl J Med 2020; 382(18): 1708—1720. https://doi.org/10.1056/NEJMoa2002032.
- Andersen KG, Rambaut A, Lipkin WI, Holmes EC, Garry RF. The proximal origin of SARS-CoV-2. Nat Med 2020; 26(4): 450-452. https://doi.org/10.1038/s41591-020-0820-9.
- Hatmal MM, Alshaer W, Al-Hatamleh MAI, Hatmal M, Smadi O, Taha MO, et al. Comprehensive structural and molecular comparison of spike proteins of SARS-CoV-2, SARS-CoV and MERS-CoV, and their interactions with ACE2. Cells 2020; 9(12): 2638. https://doi.org/10.3390/cells9122638.
- Niknam Z, Jafari A, Golchin A, Danesh Pouya F, Nemati M, Rezaei-Tavirani M, et al. Potential therapeutic options for COVID-19: an update on current evidence. Eur J Med Res 2022; 27(1): 6. https://doi.org/10.1186/s40001-021-00626-3.
- Klassen SA, Senefeld JW, Senese KA, Johnson PW, Wiggins CC, Baker SE, et al. Convalescent plasma therapy for COVID-19: a graphical mosaic of the worldwide evidence.
 Front Med 2021; 8:684151. https://doi.org/10.3389/fmed.2021.684151.
- World Health Organization. "Immunity passports" in the context of COVID-19. World Health Organization; 2020. Updated 24 April 2020. Available from: https://www.who.int/news-room/commentaries/detail/immunity-passports-in-the-context-of-covid-19.
- Rahman S, Rahman MM, Miah M, Begum MN, Sarmin M, Mahfuz M, et al. COVID-19 reinfections among naturally infected and vaccinated individuals. Sci Rep 2022; 12: 1438. https://doi.org/10.1038/s41598-022-05325-5.
- Gargouri S, Souissi A, Abid N, Chtourou A, Feki-Berrajah L, Karray R, et al. Evidence of SARS-CoV-2 symptomatic reinfection in four health care professionals from the same hospital despite the presence of antibodies. Int J Infect Dis 2022; 117: 146–154. https://doi.org/10.1016/j.ijid.2022.01.006.
- Duggan NM, Ludy SM, Shannon BC, Reisner AT, Wilcox SR.
 A case report of possible novel coronavirus 2019 reinfection.

 Am J Emerg Med 2020. https://doi.org/10.1016/j.ajem.2020.06.079.
- 10. O'Toole Aine, Scher Emily, Rambaut Andrew. *Pango lineages: latest epidemiological lineages of SARS-CoV-2*. Pango; 2022. Available from: https://cov-lineages.org/lineage_list.html.
- Nextstrain. Genomic epidemiology of SARS-CoV-2 with global subsampling. Nextstrain; 2022. Available from: https://nextstrain.org/ncov/gisaid/global.
- GISAID Fv. hCoV-19 data sharing via GISAID. GISAID;
 2022. Available from: https://www.gisaid.org/.
- 13. National Library of Medicine. SARS-CoV-2 data hub. NCBI Virus; 2022. Available from: https://www.ncbi.nlm.nih.gov/labs/virus/vssi/#/virus?SeqType_s=Nucleotide&VirusLineage_ss=Severe%20acute%20respiratory%20syndrome%20coronavirus%202,%20taxid:2697049.
- Rambaut A, Holmes EC, O'Toole Á, Hill V, McCrone JT, Ruis C, et al. A dynamic nomenclature proposal for SARS-CoV-2 lineages to assist genomic epidemiology. Nat Microbiol 2020; 5(11): 1403–1407. https://doi.org/10.1038/s41564-020-0770-5.
- Turoňová B, Sikora M, Schürmann C, Hagen WJH, Welsch S, Blanc FEC, et al. In situ structural analysis of SARS-CoV-2 spike reveals flexibility mediated by three hinges. Science 2020; 370(6513): 203–208. https://doi.org/10.1126/science.abd5223.
- Sullivan DJ, Gebo KA, Shoham S, Bloch EM, Lau B, Shenoy AG, et al. Randomized controlled trial of early outpatient COVID-19 treatment with high-titer convalescent plasma. medRxiv 2021. https://doi.org/10.1101/2021.12.10.21267485.

- Tang X, Wu C, Li X, Song Y, Yao X, Wu X, et al. On the origin and continuing evolution of SARS-CoV-2. Natl Sci Rev 2020; 7(6): 1012–1023. https://doi.org/10.1093/nsr/nwaa036.
- Winger A, Caspari T. The spike of concern-the novel variants of SARS-CoV-2. Viruses 2021; 13(6). https://doi.org/10.3390/v13061002.
- World Health Organization. Tracking SARS-CoV-2 variants.
 World Health Organization; 2022. Available from: https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/.
- European Centre for Disease Prevention and Control (ECDC). SARS-CoV-2 variants of concern as of 7 April 2022. European Centre for Disease Prevention and Control; 2022. Available from: https://www.ecdc.europa.eu/en/covid-19/variants-concern.
- Centers for Disease Control and Prevention (CDC). SARS-CoV-2 variant classifications and definitions: centers for disease control and prevention. Available from: https://www.cdc.gov/coronavirus/2019-ncov/variants/variant-classifications.
 html 2021; 2021.
- Perez-Gomez R. The development of SARS-CoV-2 variants: the gene makes the disease. J Dev Biol 2021; 9(4): 58. https://doi.org/10.3390/jdb9040058.
- Aleem A, Samad ABA, Slenker AK. Emerging variants of SARS-CoV-2 and novel therapeutics against coronavirus (COVID-19); 2021.
- McCallum M, De Marco A, Lempp FA, Tortorici MA, Pinto D, Walls AC, et al. N-terminal domain antigenic mapping reveals a site of vulnerability for SARS-CoV-2. Cell 2021; 184(9): 2332–2347.e2316. https://doi.org/10.1016/j.cell.2021.03.028.
- Suryadevara N, Shrihari S, Gilchuk P, VanBlargan LA, Binshtein E, Zost SJ, et al. Neutralizing and protective human monoclonal antibodies recognizing the N-terminal domain of the SARS-CoV-2 spike protein. Cell 2021; 184(9): 2316— 2331.e2315. https://doi.org/10.1016/j.cell.2021.03.029.
- Lopez-Rincon A, Perez-Romero CA, Tonda A, Mendoza-Maldonado L, Claassen E, Garssen J, et al. Design of specific primer set for detection of B.1.1.7 SARS-CoV-2 variant using deep learning. bioRxiv 2020. https://doi.org/10.1101/2020.12.29.424715.
- [Internet] Genomic characterisation of an emergent SARS-CoV-2 lineage in Manaus: preliminary findings; 2021 [cited 21 January 2021]. Available from:, https://virological.org/t/genomic-characterisation-of-an-emergent-sars-cov-2-lineage-in-manaus-preliminary-findings/586.
- Wibmer CK, Ayres F, Hermanus T, Madzivhandila M, Kgagudi P, Lambson BE, et al. SARS-CoV-2 501Y. V2 escapes neutralization by South African COVID-19 donor plasma. bioRxiv 2021. https://doi.org/10.1101/2021.01.18.427166.
- Harvey WT, Carabelli AM, Jackson B, Gupta RK, Thomson EC, Harrison EM, et al. SARS-CoV-2 variants, spike mutations and immune escape. Nat Rev Microbiol 2021; 19(7): 409–424. https://doi.org/10.1038/s41579-021-00573-0.
- 30. UK Health Security Agency. *COVID-19 variants identified in the UK*. UK Health Security Agency; 2022. Available from: https://www.gov.uk/government/news/covid-19-variants-identified-in-the-uk.
- 31. Greaney AJ, Loes AN, Crawford KHD, Starr TN, Malone KD, Chu HY, et al. Comprehensive mapping of mutations to the SARS-CoV-2 receptor-binding domain that affect recognition by polyclonal human serum antibodies. bioRxiv 2021. https://doi.org/10.1101/2020.12.31.425021.
- 32. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage B.1.617.2.* cov-lineages; 2021. Available from: https://cov-lineages.org/lineage.html?lineage=B.1.617.2.
- 33. Public Health England. SARS-CoV-2 variants of concern and variants under investigation in England, Technical briefing 16.

- Public Health England; 2021. Available from: https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/1001359/Variants_of_Concern_VOC Technical Briefing 16.pdf.
- 34. Stowe Julia, Andrews Nick, Gower Charlotte, Gallagher Eileen, Utsi Lara, Simmons Ruth, et al. Effectiveness of COVID-19 vaccines against hospital admission with the Delta (B.1.617.2) variant. Available from: <a href="https://khub.net/documents/135939561/479607266/Effectiveness+of+COVID-19+vaccines+against+hospital+admission+with+the+Delta+%28B.1.617.2%29+variant.pdf/1c213463-3997-ed16-2a6f-14e5deb0b997?t=1623689315431 2021; 2021.
- Sheikh A, McMenamin J, Taylor B, Robertson C. SARS-CoV-2 Delta VOC in Scotland: demographics, risk of hospital admission, and vaccine effectiveness. Lancet 2021; 397(10293): 2461–2462.
- 36. Public Health England. SARS-CoV-2 variants of concern and variants under investigation in England, Technical briefing 12. Public Health England; 2021. Available from: https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/988619/Variants_of_Concern_VOC_Technical_Briefing_12_England.pdf.
- 37. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage B.1.1.7: cov-lineages*; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=B.1.1.7.
- Davies NG, Abbott S, Barnard RC, Jarvis CI, Kucharski AJ, Munday JD, et al. Estimated transmissibility and impact of SARS-CoV-2 lineage B. 1.1. 7 in England. Science 2021; 372(6538):eabg3055.
- Davies NG, Jarvis CI, Edmunds WJ, Jewell NP, Diaz-Ordaz K, Keogh RH. Increased mortality in community-tested cases of SARS-CoV-2 lineage B. 1.1. 7. Nature 2021; 593(7858): 270–274.
- Funk T, Pharris A, Spiteri G, Bundle N, Melidou A, Carr M, et al. Characteristics of SARS-CoV-2 variants of concern B. 1.1. 7, B. 1.351 or P. 1: data from seven EU/EEA countries, weeks 38/2020 to 10/2021. Euro Surveill 2021; 26(16):2100348.
- Madhi SA, Baillie V, Cutland CL, Voysey M, Koen AL, Fairlie L, et al. Efficacy of the ChAdOx1 nCoV-19 Covid-19 vaccine against the B. 1.351 variant. N Engl J Med 2021; 384(20): 1885–1898.
- 42. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage B.1.351: cov-lineages*; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=B.1.351.
- 43. Pearson Carl AB, Russell TW, Davies Nicholas G, Kucharski Adam J, CMMID COVID-19 working group, Edmunds W John, Eggo Rosalind M. Estimates of severity and transmissibility of novel South Africa SARS-CoV-2 variant 501Y.V2; 2021. Available from: https://cmmid.github.io/topics/covid19/reports/sa-novel-variant/2021_01_11
 Transmissibility_and_severity_of_501Y_V2_in_SA.pdf.
- Cele S, Gazy I, Jackson L, Hwa S-H, Tegally H, Lustig G, et al. Escape of SARS-CoV-2 501Y.V2 from neutralization by convalescent plasma. Nature 2021; 593(7857): 142–146.
- Tegally H, Wilkinson E, Giovanetti M, Iranzadeh A, Fonseca V, Giandhari J, et al. Detection of a SARS-CoV-2 variant of concern in South Africa. Nature 2021; 592(7854): 438–443.
- O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage P.1: cov-lineages*; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=P.1.
- Dejnirattisai W, Zhou D, Supasa P, Liu C, Mentzer AJ, Ginn HM, et al. Antibody evasion by the P. 1 strain of SARS-CoV-2. Cell 2021; 184(11): 2939–2954. e2939.
- Faria NR, Mellan TA, Whittaker C, Claro IM, Candido DdS, Mishra S, et al. Genomics and epidemiology of the P. 1 SARS-CoV-2 lineage in Manaus, Brazil. Science 2021; 372(6544): 815-821.

- 49. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage B.1.429*. cov-lineages; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=B.1.429.
- O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage B.1.427*. cov-lineages; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=B.1.427.
- Collier DA, De Marco A, Ferreira IA, Meng B, Datir RP, Walls AC, et al. Sensitivity of SARS-CoV-2 B. 1.1. 7 to mRNA vaccine-elicited antibodies. Nature 2021; 593(7857): 136–141.
- 52. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage B.1.525*. cov-lineages; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=B.1.525.
- 53. Jangra S, Ye C, Rathnasinghe R, Stadlbauer D, Alshammary H, Amoako AA, et al. SARS-CoV-2 spike E484K mutation reduces antibody neutralisation. Lancet Microbe 2021; 2(7): e283–e284.
- Zhao X, Zheng A, Li D, Zhang R, Sun H, Wang Q, et al. Neutralisation of ZF2001-elicited antisera to SARS-CoV-2 variants. Lancet Microbe 2021; 2(10): e494.
- 55. Public Health England. SARS-CoV-2 variants of concern and variants under investigation in England, Technical briefing 11; 2021. Available from: https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/986380/Variants_of_Concern_VOC_Technical_Briefing_11_England.pdf.
- 56. Tut G, Lancaster T, Krutikov M, Sylla P, Bone D, Kaur N, et al. Profile of humoral and cellular immune responses to single doses of BNT162b2 or ChAdOx1 nCoV-19 vaccines in residents and staff within residential care homes (VIVALDI): an observational study. Lancet Healthy Longev 2021; 2(9): e544–e553.
- Yadav PD, Sapkal GN, Abraham P, Ella R, Deshpande G, Patil DY, et al. Neutralization of variant under investigation B. 1.617. 1 with sera of BBV152 vaccinees. Clin Infect Dis 2022; 74(2): 366–368.
- O'Toole Aine, Scher Emily, Rambaut Andrew. Lineage B.1.617.1: cov-lineages; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=B.1.617.1.
- O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage P.2*. cov-lineages; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=P.2.
- Acevedo ML, Gaete-Argel A, Alonso-Palomares L, de Oca MM, Bustamante A, Gaggero A, et al. Differential neutralizing antibody responses elicited by CoronaVac and BNT162b2 against SARS-CoV-2 Lambda in Chile. Nat Microbiol 2022; 7(4): 524–529. https://doi.org/10.1038/s41564-022-01092-1.
- Tada T, Zhou H, Dcosta BM, Samanovic MI, Mulligan MJ, Landau NR. SARS-CoV-2 lambda variant remains susceptible to neutralization by mRNA vaccine-elicited antibodies and convalescent serum. bioRxiv 2021. https://doi.org/10.1101/2021.07.02.450959.
- O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage C.37*.
 cov-lineages; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=C.37.
- O'Toole Aine, Scher Emily, Rambaut Andrew. Lineage B.1.621. cov-lineages; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=B.1.621.
- 64. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage B.1.526*. cov-lineages; 2020. Available from: https://cov-lineages.org/lineage.html?lineage=B.1.526.
- 65. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage XD*. cov-lineages; 2022. Available from: https://cov-lineages.org/lineage.html?lineage=XD.
- O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage XE*. cov-lineages; 2022. Available from: https://cov-lineages.org/lineage.html?lineage=XE.

- O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage XF*. cov-lineages; 2022. Available from: https://cov-lineages.org/lineage.html?lineage=XF.
- 68. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage BA.1*. cov-lineages; 2021. Available from: https://cov-lineages.org/lineage.html?lineage=BA.1.
- 69. Agency UHS. COVID-19 vaccine surveillance report, week 6; 2022. Available from: https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/1054071/vaccine-surveillance-report-week-6.pdf.
- 70. European Centre for Disease Prevention and Control (ECDC). Assessment of the further spread and potential impact of the SARS-CoV-2 Omicron variant of concern in the EU, EU/EEA, 19th update; 2022. Available from, https://www.ecdc.europa.eu/sites/default/files/documents/RRA-19-update-27-jan-2022.pdf.
- Altarawneh HN, Chemaitelly H, Hasan MR, Ayoub HH, Qassim S, AlMukdad S, et al. Protection against the omicron variant from previous SARS-CoV-2 infection. N Engl J Med 2022; 386(13): 1288–1290.
- 72. Hui KP, Ho JC, Cheung M-C, Ng K-C, Ching RH, Lai K-L, et al. SARS-CoV-2 Omicron variant replication in human bronchus and lung ex vivo. Nature 2022; 603(7902): 715–720. https://doi.org/10.1038/s41586-022-04479-6.
- Rössler A, Knabl L, von Laer D, Kimpel J. Neutralization profile after recovery from SARS-CoV-2 Omicron infection. N Engl J Med 2022; 386: 1764–1766. https://doi.org/10.1056/NEJMc2201607.
- 74. UK Health Security Agency (UKHSA) UHSA. SARS-CoV-2 variants of concern and variants under investigation in England Technical briefing: update on hospitalisation and vaccine effectiveness for Omicron VOC-21NOV-01 (B.1.1.529); 2021. Available from, https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/1045619/Technical-Briefing-31-Dec-2021-Omicron_severity_update.pdf.
- 75. Ferguson NGA, Hinsley W, Volz E, team obotICC-r. Report 50: hospitalisation risk for Omicron cases in England; 2021. Available from, https://www.imperial.ac.uk/media/imperial-college/medicine/mrc-gida/2021-12-22-COVID19-Report-50.pdf.
- O'Toole Aine, Scher Emily, Rambaut Andrew. *Lineage BA.2*. cov-lineages; 2021. Available from, https://cov-lineages.org/lineage_html?lineage=BA.2.
- UK Health Security Agency. SARS-CoV-2 variants of concern and variants under investigation in England, Technical briefing 35; 2022. Available from, https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/1050999/Technical-Briefing-35-28January2022.pdf.
- 78. UK Health Security Agency. SARS-CoV-2 variants of concern and variants under investigation in England, Technical briefing 39; 2022. Available from, https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/1063424/Tech-Briefing-39-25March2022_FINAL.pdf.
- Fonager J, Bennedbæk M, Bager P, Wohlfahrt J, Ellegaard KM, Ingham AC, et al. Molecular epidemiology of the SARS-CoV-2 variant omicron BA. 2 sub-lineage in Denmark, 29 November 2021 to 2 January 2022. Euro Surveill 2022; 27(10):2200181.
- 80. Lyngse FP, Kirkeby CT, Denwood M, Christiansen LE, Mølbak K, Møller CH, et al. *Transmission of SARS-CoV-2 Omicron VOC subvariants BA. 1 and BA. 2.* evidence from Danish Households. medRxiv; 2022.
- O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage BA.3*. cov-lineages; 2021. Available from, https://cov-lineages.org/lineage.html?lineage=BA.3.
- 82. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage BA.4*. cov-lineages; 2022. Available from, https://cov-lineages.org/lineage.html?lineage=BA.4.

- 83. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage BA.5*. cov-lineages; 2022. Available from, https://cov-lineages.org/lineage.html?lineage=BA.5.
- 84. European Centre for Disease Prevention and Control (ECDC). SARS-CoV-2 variants of concern as of 15 July 2022. European Centre for Disease Prevention and Control; 2022. Available from, https://www.ecdc.europa.eu/en/covid-19/variants-concern.
- 85. O'Toole Áine, Scher Emily, Rambaut Andrew. *Lineage BA.2.75*. cov-lineages; 2022. Available from, https://cov-lineages.org/lineage.html?lineage=BA.2.75.
- Soler E, Houdebine LM. Preparation of recombinant vaccines. Biotechnol Annu Rev 2007; 13: 65–94. https://doi.org/10.1016/51387-2656(07)13004-0.
- 87. Clem AS. Fundamentals of vaccine immunology. J Glob Infect Dis 2011; 3(1): 73–78. https://doi.org/10.4103/0974-777X.77299.
- Pulendran B, Ahmed R. Immunological mechanisms of vaccination. Nat Immunol 2011; 12(6): 509–517. https://doi.org/10.1038/ni.2039.
- US Government Accountability Office. Vaccine development; capabilities and challenges for addressing infectious diseases. GAO, U.S. Government Accountability Office; 2021. Available from: https://www.gao.gov/assets/720/717636.pdf.
- Howard KL, Wright CN. Operation warp speed: accelerated COVID-19 vaccine development status and efforts to address manufacturing challenges. GAO, U.S. Government Accountability Office; 2021. Available from: https://www.gao.gov/products/gao-21-319.
- Sharma O, Sultan AA, Ding H, Triggle CR. A review of the progress and challenges of developing a vaccine for COVID-19. Front Immunol 2020; 11:585354. https://doi.org/10.3389/fimmu.2020.585354.
- World Health Organization. COVID-19 vaccine tracker and landscape. Available from: https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines 2022;
 2022.
- Robert-Guroff M. Replicating and non-replicating viral vectors for vaccine development. Curr Opin Biotechnol 2007; 18(6): 546–556. https://doi.org/10.1016/j.copbio.2007.10.010.
- 94. Leitner WW, Ying H, Restifo NP. DNA and RNA-based vaccines: principles, progress and prospects. Vaccine 1999; 18(9–10): 765–777. https://doi.org/10.1016/s0264-410x(99)00271-6.
- Dan JM, Mateus J, Kato Y, Hastie KM, Yu ED, Faliti CE, et al. Immunological memory to SARS-CoV-2 assessed for up to 8 months after infection. Science 2021; 371(6529):eabf4063. https://doi.org/10.1126/science.abf4063.
- West R, Kobokovich A, Connell N, Gronvall GK. COVID-19 antibody tests: a valuable public health tool with limited relevance to individuals. Trends Microbiol 2020. https://doi.org/10.1016/j.tim.2020.11.002.
- Shah VK, Firmal P, Alam A, Ganguly D, Chattopadhyay S. Overview of immune response during SARS-CoV-2 infection: lessons from the past. Front Immunol 2020; (1949): 11. https://doi.org/10.3389/fimmu.2020.01949.
- Ali AM, Ali KM, Fatah MH, Tawfeeq HM, Rostam HM. SARS-CoV-2 reinfection in patients negative for immunoglobulin G following recovery from COVID-19. medRxiv 2020. https://doi.org/10.1101/2020.11.20.20234385.
- Mustapha JO, Abdullahi IN, Ajagbe OOR, Emeribe AU, Fasogbon SA, Onoja SO, et al. Understanding the implications of SARS-CoV-2 re-infections on immune response milieu, laboratory tests and control measures against COVID-19. Heliyon 2021; 7(1):e05951. https://doi.org/10.1016/j.heliyon.2021.e05951.
- 100. Lee NH, Lee JA, Park SY, Song CS, Choi IS, Lee JB. A review of vaccine development and research for industry animals in

- Korea. Clin Exp Vaccine Res 2012; 1(1): 18–34. https://doi.org/10.7774/cevr.2012.1.1.18.
- 101. Lurie N, Saville M, Hatchett R, Halton J. Developing covid-19 vaccines at pandemic speed. N Engl J Med 2020; 382(21): 1969—1973. https://doi.org/10.1056/NEJMp2005630.
- 102. Forman R, Shah S, Jeurissen P, Jit M, Mossialos E. COVID-19 vaccine challenges: what have we learned so far and what remains to be done? Health Policy 2021; 125(5): 553-567.
- 103. Ou X, Liu Y, Lei X, Li P, Mi D, Ren L, et al. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. Nat Commun 2020; 11(1): 1620. https://doi.org/10.1038/s41467-020-15562-9.
- 104. Wallis J, Shenton DP, Carlisle RC. Novel approaches for the design, delivery and administration of vaccine technologies. Clin Exp Immunol 2019; 196(2): 189–204. https://doi.org/10.1111/cei.13287.
- Karch CP, Burkhard P. Vaccine technologies: from whole organisms to rationally designed protein assemblies. Biochem Pharmacol 2016; 120: 1–14. https://doi.org/10.1016/j.bcp.2016.05.001.
- Christensen D. Vaccine adjuvants: why and how. Hum Vaccin Immunother 2016; 12(10): 2709–2711. https://doi.org/10.1080/21645515.2016.1219003.
- 107. Riese P, Schulze K, Ebensen T, Prochnow B, Guzman CA. Vaccine adjuvants: key tools for innovative vaccine design. Curr Top Med Chem 2013; 13(20): 2562–2580. https://doi.org/10.2174/15680266113136660183.
- Woodland DL, Hogan RJ, Zhong W. Cellular immunity and memory to respiratory virus infections. Immunol Res 2001; 24(1): 53-67. https://doi.org/10.1385/IR:24:1:53.
- Mullard A. New drugs cost US\$2.6 billion to develop. Nat Rev Drug Discov 2014; 13(12). https://doi.org/10.1038/nrd4507.877-877.
- Uskokovic V. Nanomedicine for the poor: a lost cause or an idea whose time has yet to come? Nanomedicine 2021; 16(14): 1203–1218. https://doi.org/10.2217/nnm-2021-0024.
- 111. Yarlagadda H, Patel MA, Gupta V, Bansal T, Upadhyay S, Shaheen N, et al. COVID-19 vaccine challenges in developing and developed countries. Cureus 2022; 14(4).
- GAVI. The 2021-2025 investment opportunity. Gavi The vaccine Alliance; 2021. Available from: https://www.gavi.org/investing-gavi/resource-mobilisation-process/gavis-3rd-donor-pledging-conference-june-2020.
- 113. Harlan C, Pitrelli S. From Oxford to an Italian lab, one race for coronavirus vaccine is gaining backers. The Washington Post; 2020. Available from: https://www.washingtonpost.com/world/europe/vaccine-coronavirus-oxford-italy-trials/2020/06/20/4b8f5e0a-af1c-11ea-98b5-279a6479a1e4 story.html.
- 114. United Nations. *UN analysis shows link between lack of vaccine equity and widening poverty gap*. UN News; 2022. Available from: https://news.un.org/en/story/2022/03/1114762.
- Maxmen A. The fight to manufacture COVID vaccines in lower-income countries. Nature 2021; 597(7877): 455–457. https://doi.org/10.1038/d41586-021-02383-z.
- UNDP. Vaccine equity and speed: data futures platform; 2022.
 Available from: https://data.undp.org/vaccine-equity-archive/vaccine-equity-and-speed/.
- 117. Peter S, Goodman AM, Robbins Rebecca, Stevis-Gridneff Matina. What would it take to vaccinate the world against covid?. The New York Times; 2021. Available from: https://www.nytimes.com/2021/05/15/world/americas/covid-yaccine-patent-biden.html.
- 118. Kavanagh MM, Gostin LO, Sunder M. Sharing technology and vaccine doses to address global vaccine inequity and end the COVID-19 pandemic. **JAMA 2021**; 326(3): 219–220. https://doi.org/10.1001/jama.2021.10823.
- 119. Center DUGHI. COVID-19 vaccines and treatment: the race for global equity: launch and scale speedometer. Duke

- University; 2021. Available from: https://jamanetwork.com/journals/jama/fullarticle/2781756.
- 120. Global Health Centre at the Graduate Institute G. COVID-19 vaccine access. Geneva: Global Health Centre at the Graduate Institute; 2022. Available from: https://www.knowledgeportalia.org/covid19-vaccine-arrangements.
- 121. Nachega JB, Sam-Agudu NA, Masekela R, van der Zalm MM, Nsanzimana S, Condo J, et al. Addressing challenges to rolling out COVID-19 vaccines in African countries. Lancet Glob Health 2021; 9(6): e746—e748.
- Chakraborty C, Agoramoorthy G. India's cost-effective COVID-19 vaccine development initiatives. Vaccine 2020; 38(50): 7883.
- 123. Finch WH, Hernández Finch ME. Poverty and Covid-19: rates of incidence and deaths in the United States during the first 10 weeks of the pandemic. Front Sociol 2020; 5(47). https://doi.org/10.3389/fsoc.2020.00047.
- Wise J. Covid-19: low skilled men have highest death rate of working age adults. BMJ 2020; 369: m1906. https://doi.org/10.1136/bmj.m1906.
- 125. Butler R, MacDonald NE, Hesitancy SWGoV. Diagnosing the determinants of vaccine hesitancy in specific subgroups: the Guide to Tailoring Immunization Programmes (TIP). Vaccine 2015; 33(34): 4176-4179. https://doi.org/10.1016/j.vaccine.2015.04.038.
- 126. Ten threats to global health in 2019: WHO; [Available from: https://www.who.int/news-room/spotlight/ten-threats-to-global-health-in-2019.
- 127. Norhayati MN, Che Yusof R, Azman YM. Systematic review and meta-analysis of COVID-19 vaccination acceptance. Front Med 2021; 8:783982. https://doi.org/10.3389/fmed.2021.783982.
- 128. Mahmud S, Mohsin M, Hossain S, Islam MM, Muyeed A. The acceptance of COVID-19 vaccine at early stage of development and approval: a global systematic review and meta-analysis. Heliyon 2022; 8(9):e10728. https://doi.org/10.1016/j.heliyon.2022.e10728.
- 129. Leigh JP, Moss SJ, White TM, Picchio CA, Rabin KH, Ratzan SC, et al. Factors affecting COVID-19 vaccine hesitancy among healthcare providers in 23 countries. Vaccine 2022; 40(31): 4081–4089. https://doi.org/10.1016/j.vaccine.2022.04.097.
- 130. Terry E, Cartledge S, Damery S, Greenfield S. Factors associated with COVID-19 vaccine intentions during the COVID-19 pandemic; a systematic review and meta-analysis of cross-sectional studies. BMC Public Health 2022; 22(1): 1667. https://doi.org/10.1186/s12889-022-14029-4.
- 131. Thorsteinsdottir B, Madsen BE. Prioritizing health care workers and first responders for access to the COVID19 vaccine is not unethical, but both fair and effective an ethical analysis. Scand J Trauma Resusc Emerg Med 2021; 29(1): 77. https://doi.org/10.1186/s13049-021-00886-2.
- 132. Lazarus JV, Wyka K, White TM, Picchio CA, Rabin K, Ratzan SC, et al. Revisiting COVID-19 vaccine hesitancy around the world using data from 23 countries in 2021. Nat Commun 2022; 13(1): 3801. https://doi.org/10.1038/s41467-022-31441-x.
- 133. Hatmal MM, Al-Hatamleh MAI, Olaimat AN, Mohamud R, Fawaz M, Kateeb ET, et al. Reported adverse effects and attitudes among Arab populations following COVID-19 vaccination: a large-scale multinational study implementing machine learning tools in predicting post-vaccination adverse effects based on predisposing factors. Vaccines 2022; 10(3). https://doi.org/10.3390/vaccines10030366.
- 134. Riad A, Pokorna A, Antalova N, Krobot M, Zviadadze N, Serdiuk I, et al. Prevalence and drivers of COVID-19 vaccine hesitancy among Czech University students: national cross-

- sectional study. **Vaccines 2021**; 9(9): 948. https://doi.org/10.3390/vaccines9090948.
- 135. Al-Qerem WA, Jarab AS. COVID-19 vaccination acceptance and its associated factors among a middle Eastern population. Front Public Health 2021; 9:632914. https://doi.org/10.3389/fpubh.2021.632914.
- 136. Riad A, Jouzova A, Ustun B, Lagova E, Hruban L, Janku P, et al. COVID-19 vaccine acceptance of pregnant and lactating women (PLW) in Czechia: an analytical cross-sectional study. Int J Environ Res Public Health 2021; 18(24):13373. https://doi.org/10.3390/ijerph182413373.
- 137. Gupta S, Watanabe S, Laurent SM. Psychological predictors of vaccination intentions among U.S. undergraduates and online panel workers during the 2020 COVID-19 pandemic. PLoS One 2021; 16(11):e0260380. https://doi.org/10.1371/journal.pone.0260380.
- Riad A, Abdulqader H, Morgado M, Domnori S, Koscik M, Mendes JJ, et al. Global prevalence and drivers of dental students' COVID-19 vaccine hesitancy. Vaccines 2021; 9(6): 566. https://doi.org/10.3390/vaccines9060566.
- 139. Jennings W, Stoker G, Bunting H, Valgarethsson VO, Gaskell J, Devine D, et al. Lack of trust, conspiracy beliefs, and social media use predict COVID-19 vaccine hesitancy. Vaccines 2021; 9(6). https://doi.org/10.3390/vaccines9060593.
- 140. Lounis M, Bencherit D, Rais MA, Riad A. COVID-19 vaccine booster hesitancy (VBH) and its drivers in Algeria: national cross-sectional survey-based study. Vaccines 2022; 10(4): 621. https://doi.org/10.3390/vaccines10040621.
- 141. Klugar M, Riad A, Mohanan L, Pokorna A. COVID-19 vaccine booster hesitancy (VBH) of healthcare workers in Czechia: national cross-sectional study. Vaccines 2021; 9(12): 1437. https://doi.org/10.3390/vaccines9121437.
- 142. Dziedzic A, Issa J, Hussain S, Tanasiewicz M, Wojtyczka R, Kubina R, et al. COVID-19 vaccine booster hesitancy (VBH) of healthcare professionals and students in Poland: cross-sectional survey-based study. Front Public Health 2022; 10: 938067. https://doi.org/10.3389/fpubh.2022.938067.
- 143. Attia S, Mausbach K, Klugar M, Howaldt HP, Riad A. Prevalence and drivers of COVID-19 vaccine booster hesitancy among German University students and employees. Front Public Health 2022; 10:846861. https://doi.org/10.3389/fpubh.2022.846861.
- 144. Hatmal MM, Al-Hatamleh MAI, Olaimat AN, Hatmal M, Alhaj-Qasem DM, Olaimat TM, et al. Side effects and perceptions following COVID-19 vaccination in Jordan: a randomized, cross-sectional study implementing machine learning for predicting severity of side effects. Vaccines 2021; 9(6). https://doi.org/10.3390/vaccines9060556.
- 145. Klein NP, Lewis N, Goddard K, Fireman B, Zerbo O, Hanson KE, et al. Surveillance for adverse events after COVID-19 mRNA vaccination. JAMA 2021; 326(14): 1390–1399. https://doi.org/10.1001/jama.2021.15072.
- 146. Riad A, Schunemann H, Attia S, Pericic TP, Zuljevic MF, Jurisson M, et al. COVID-19 vaccines safety tracking (CoVaST): protocol of a multi-center prospective cohort study for active surveillance of COVID-19 vaccines' side effects. Int J Environ Res Public Health 2021; 18(15): 7859. https://doi.org/10.3390/ijerph18157859.
- 147. Wu Q, Dudley MZ, Chen X, Bai X, Dong K, Zhuang T, et al. Evaluation of the safety profile of COVID-19 vaccines: a rapid review. BMC Med 2021; 19(1): 173. https://doi.org/10.1186/s12916-021-02059-5.
- 148. Riad A, Sagiroglu D, Ustun B, Pokorna A, Klugarova J, Attia S, et al. Prevalence and risk factors of CoronaVac side effects: an independent cross-sectional study among healthcare workers in Turkey. J Clin Med 2021; 10(12): 2629. https://doi.org/10.3390/jcm10122629.

- 149. Riad A, Pokorna A, Attia S, Klugarova J, Koscik M, Klugar M. Prevalence of COVID-19 vaccine side effects among healthcare workers in the Czech Republic. J Clin Med 2021; 10(7): 1428. https://doi.org/10.3390/jcm10071428.
- 150. Riad A, Hockova B, Kantorova L, Slavik R, Spurna L, Stebel A, et al. Side effects of mRNA-based COVID-19 vaccine: nationwide phase IV study among healthcare workers in Slovakia. Pharmaceuticals 2021; 14(9): 873. https://doi.org/10.3390/ph14090873.
- 151. Riad A, Pokorna A, Klugarova J, Antalova N, Kantorova L, Koscik M, et al. Side effects of mRNA-based COVID-19 vaccines among young adults (18-30 Years old): an independent post-marketing study. Pharmaceuticals 2021; 14(10): 1049. https://doi.org/10.3390/ph14101049.
- 152. Yesuf EA, Riad A, Sofi-Mahmudi A, Sudhakar M, Mekonnen A, Endalkachew S, et al. Self-reported side effects of the Oxford AstraZeneca COVID-19 vaccine among healthcare workers in Ethiopia, Africa: a cross-sectional study. Front Public Health 2022; 10:937794. https://doi.org/10.3389/fpubh.2022.937794.
- 153. Orebi HA, Emara HE, Alhindi AA, Shahin MR, Hegazy AH, Kabbash IA, et al. Perceptions and experiences of COVID-19 vaccines' side effects among healthcare workers at an Egyptian University Hospital: a cross-sectional study. Trop Med Health 2022; 50(1): 37. https://doi.org/10.1186/s41182-022-00427-2.
- 154. Ganesan S, Al Ketbi LMB, Al Kaabi N, Al Mansoori M, Al Maskari NN, Al Shamsi MS, et al. Vaccine side effects following COVID-19 vaccination among the residents of the UAE-an observational study. Front Public Health 2022; 10: 876336. https://doi.org/10.3389/fpubh.2022.876336.
- 155. Milken Institute's COVID-19 treatment and vaccine tracker. Milken Institute; 2020. Available from: https://covid-19tracker.milkeninstitute.org/#vaccines intro.
- 156. Khan Z, Karataş Y, Ceylan AF, Rahman H. COVID-19 and therapeutic drugs repurposing in hand: the need for collaborative efforts. Pharm Hosp Clin 2020. https://doi.org/10.1016/j.phclin.2020.06.003.
- 157. Santos J, Brierley S, Gandhi MJ, Cohen MA, Moschella PC, Declan ABL. Repurposing therapeutics for potential treatment of SARS-CoV-2: a review. Viruses 2020; 12(7). https://doi.org/10.3390/v12070705.
- 158. Rajoli RK, Pertinez H, Arshad U, Box H, Tatham L, Curley P, et al. Dose prediction for repurposing nitazoxanide in SARS-CoV-2 treatment or chemoprophylaxis. medRxiv 2020. https://doi.org/10.1101/2020.05.01.20087130.
- 159. Yamamoto M, Kiso M, Sakai-Tagawa Y, Iwatsuki-Horimoto K, Imai M, Takeda M, et al. The anticoagulant nafamostat potently inhibits SARS-CoV-2 S protein-mediated fusion in a cell fusion assay system and viral infection in vitro in a cell-type-dependent manner. Viruses 2020; 12(6). https://doi.org/10.3390/v12060629.
- 160. Singh AK, Singh A, Shaikh A, Singh R, Misra A. Chloroquine and hydroxychloroquine in the treatment of COVID-19 with or without diabetes: a systematic search and a narrative review with a special reference to India and other developing countries. Diabetes Metab Syndr 2020; 14(3): 241–246. https://doi.org/10.1016/j.dsx.2020.03.011.
- 161. Tang N, Bai H, Chen X, Gong J, Li D, Sun Z. Anticoagulant treatment is associated with decreased mortality in severe coronavirus disease 2019 patients with coagulopathy. J Thromb Haemost 2020; 18(5): 1094–1099. https://doi.org/10.1111/jth.14817.
- 162. Shi C, Wang C, Wang H, Yang C, Cai F, Zeng F, et al. The potential of low molecular weight heparin to mitigate cytokine storm in severe COVID-19 patients: a retrospective clinical study. medRxiv 2020. https://doi.org/10.1101/2020.03.28.20046144.

- 163. Shi R, Shan C, Duan X, Chen Z, Liu P, Song J, et al. A human neutralizing antibody targets the receptor-binding site of SARS-CoV-2. Nature 2020; 584(7819): 120–124. https://doi.org/10.1038/s41586-020-2381-v.
- 164. Liu L, Wang P, Nair MS, Yu J, Rapp M, Wang Q, et al. Potent neutralizing antibodies against multiple epitopes on SARS-CoV-2 spike. Nature 2020; 584(7821): 450–456. https://doi.org/10.1038/s41586-020-2571-7.
- 165. Pinto D, Park YJ, Beltramello M, Walls AC, Tortorici MA, Bianchi S, et al. Structural and functional analysis of a potent sarbecovirus neutralizing antibody. bioRxiv 2020. https://doi.org/10.1101/2020.04.07.023903.
- 166. te Velthuis AJ, van den Worm SH, Sims AC, Baric RS, Snijder EJ, van Hemert MJ. Zn(2+) inhibits coronavirus and arterivirus RNA polymerase activity in vitro and zinc ionophores block the replication of these viruses in cell culture. PLoS Pathog 2010; 6(11):e1001176. https://doi.org/10.1371/journal.ppat.1001176.
- 167. Kumar A, Kubota Y, Chernov M, Kasuya H. Potential role of zinc supplementation in prophylaxis and treatment of COVID-19. Med Hypotheses 2020; 144:109848. https://doi.org/10.1016/j.mehy.2020.109848.
- 168. Derwand R, Scholz M. Does zinc supplementation enhance the clinical efficacy of chloroquine/hydroxychloroquine to win today's battle against COVID-19? Med Hypotheses 2020; 142: 109815. https://doi.org/10.1016/j.mehy.2020.109815.
- 169. Bauer SR, Kapoor A, Rath M, Thomas SA. What is the role of supplementation with ascorbic acid, zinc, vitamin D, or Nacetylcysteine for prevention or treatment of COVID-19? Clevel Clin J Med 2020. https://doi.org/10.3949/ccjm.87a.ccc046.
- 170. Al-Hatamleh MAI, Hatmal MM, Sattar K, Ahmad S, Mustafa MZ, Bittencourt MDC, et al. Antiviral and immunomodulatory effects of phytochemicals from honey against COVID-19: potential mechanisms of action and future directions. Molecules 2020; 25: 5017. https://doi.org/10.3390/molecules25215017.
- 171. Lembo D, Donalisio M, Civra A, Argenziano M, Cavalli R. Nanomedicine formulations for the delivery of antiviral drugs: a promising solution for the treatment of viral infections. Expert Opin Drug Deliv 2018; 15(1): 93–114. https://doi.org/10.1080/17425247.2017.1360863.
- 172. Wu F, Wang A, Liu M, Wang Q, Chen J, Xia S, et al. Neutralizing antibody responses to SARS-CoV-2 in a COVID-19 recovered patient cohort and their implications. medRxiv 2020. https://doi.org/10.1101/2020.03.30.20047365.
- 173. Marano G, Vaglio S, Pupella S, Facco G, Catalano L, Liumbruno GM, et al. Convalescent plasma: new evidence for an old therapeutic tool? **Blood Transfus 2016**; 14(2): 152–157. https://doi.org/10.2450/2015.0131-15.
- 174. Casadevall A, Pirofski L-A. Antibody-mediated regulation of cellular immunity and the inflammatory response. Trends Immunol 2003; 24(9): 474–478.
- 175. Bloch EM, Shoham S, Casadevall A, Sachais BS, Shaz B, Winters JL, et al. Deployment of convalescent plasma for the prevention and treatment of COVID-19. J Clin Investig 2020; 130(6): 2757–2765.
- 176. Casadevall A, Henderson JP, Joyner MJ, Pirofski LA. SARS-CoV-2 variants and convalescent plasma: reality, fallacies, and opportunities. J Clin Investig 2021; 131(7). https://doi.org/10.1172/jci148832.
- 177. Walensky RP, Walke HT, Fauci AS. SARS-CoV-2 variants of concern in the United States-challenges and opportunities. JAMA 2021; 325(11): 1037–1038. https://doi.org/10.1001/jama.2021.2294.
- 178. Bégin P, Callum J, Jamula E, Cook R, Heddle NM, Tinmouth A, et al. Convalescent plasma for hospitalized patients with COVID-19: an open-label, randomized controlled

- trial. **Nat Med 2021**; 27(11): 2012—2024. https://doi.org/10.1038/s41591-021-01488-2.
- Casadevall A, Pirofski LA, Joyner MJ. The principles of antibody therapy for infectious diseases with relevance for COVID-19. mBio 2021; 12(2). https://doi.org/10.1128/mBio.03372-20.
- Lesne E, Brodin P, Saint-Girons F. Plasma therapy in influenza. Presse Med 1919; 27: 181–182.
- 181. Mair-Jenkins J, Saavedra-Campos M, Baillie JK, Cleary P, Khaw FM, Lim WS, et al. The effectiveness of convalescent plasma and hyperimmune immunoglobulin for the treatment of severe acute respiratory infections of viral etiology: a systematic review and exploratory meta-analysis. J Infect Dis 2015; 211(1): 80–90. https://doi.org/10.1093/infdis/jiu396.
- 182. Goodarzi P, Mahdavi F, Mirzaei R, Hasanvand H, Sholeh M, Zamani F, et al. Coronavirus disease 2019 (COVID-19): immunological approaches and emerging pharmacologic treatments. Int Immunopharm 2020; 88:106885. https://doi.org/10.1016/j.intimp.2020.106885.
- 183. Duan K, Liu B, Li C, Zhang H, Yu T, Qu J, et al. Effectiveness of convalescent plasma therapy in severe COVID-19 patients. Proc Natl Acad Sci U S A 2020; 117(17): 9490–9496. https://doi.org/10.1073/pnas.2004168117.
- 184. Liu STH, Lin H-M, Baine I, Wajnberg A, Gumprecht JP, Rahman F, et al. Convalescent plasma treatment of severe COVID-19: a propensity score—matched control study. Nat Med 2020; 26(11): 1708–1713. https://doi.org/10.1038/s41591-020-1088-9.
- 185. March D. New study validates benefits of convalescent plasma for some patients with COVID-19. NYU Langone Health; 2022. Available from: https://nyulangone.org/news/new-study-validates-benefits-convalescent-plasma-some-patients-covid-19.
- 186. Troxel AB, Petkova E, Goldfeld K, Liu M, Tarpey T, Wu Y, et al. Association of convalescent plasma treatment with clinical status in patients hospitalized with COVID-19: a meta-analysis. JAMA Netw Open 2022; 5(1). e2147331—e2147331.
- 187. Simonovich VA, Burgos Pratx LD, Scibona P, Beruto MV, Vallone MG, Vázquez C, et al. A randomized trial of convalescent plasma in Covid-19 severe pneumonia. N Engl J Med 2021; 384(7): 619–629.
- 188. The Science Advisory Board. FDA issues EUA for GSK-Vir COVID-19 mAb treatment. The Science Advisory Board; 2021. Available from: https://www.scienceboard.net/index.aspx?sec=ser&sub=def&pag=dis&ItemID=2761.
- 189. Dougan M, Nirula A, Gottlieb R, Azizad M, Mocherla B, Chen P, et al. Bamlanivimab + etesevimab for treatment of COVID-19 in high-risk ambulatory patients. Top Antivir Med 2021. 33–33.
- 190. Park Hyung, Tarpey Thaddeus, Li Yi, Petkova Eva, Antman Elliott, Troxel Andrea. *Is treatment with convalescent plasma right for your patient*. COMPILE; 2021. Available from: https://covid-convalescentplasma-tbi-calc.org/.
- 191. Ortigoza MB, Yoon H, Goldfeld KS, Troxel AB, Daily JP, Wu Y, et al. Efficacy and safety of COVID-19 convalescent plasma in hospitalized patients: a randomized clinical trial. JAMA Intern Med 2021; 182(2): 115–126. https://doi.org/10.1001/jamainternmed.2021.6850.
- 192. Joyner MJ, Carter RE, Senefeld JW, Klassen SA, Mills JR, Johnson PW, et al. Convalescent plasma antibody levels and the risk of death from Covid-19. N Engl J Med 2021; 384(11): 1015–1027.
- 193. Luke TC, Kilbane EM, Jackson JL, Hoffman SL. Metaanalysis: convalescent blood products for Spanish influenza pneumonia: a future H5N1 treatment? Ann Intern Med 2006; 145(8): 599-609.
- 194. Horby PW, Estcourt L, Peto L, Emberson JR, Staplin N, Spata E, et al. Convalescent plasma in patients admitted to

- hospital with COVID-19 (RECOVERY): a randomised, controlled, open-label, platform trial. medRxiv; 2021.
- 195. Mora-Rillo M, Arsuaga M, Ramirez-Olivencia G, de la Calle F, Borobia AM, Sanchez-Seco P, et al. Acute respiratory distress syndrome after convalescent plasma use: treatment of a patient with Ebola virus disease contracted in Madrid, Spain. Lancet Respir Med 2015; 3(7): 554–562. https://doi.org/10.1016/S2213-2600(15)00180-0.
- 196. Wang KY, Shah P, Pierce M. Convalescent plasma for COVID-19 complicated by ARDS due to TRALI. BMJ Case Rep 2021; 14(1). https://doi.org/10.1136/bcr-2020-239762.
- Roback JD, Guarner J. Convalescent plasma to treat COVID-19: possibilities and challenges. JAMA 2020; 323(16): 1561– 1562. https://doi.org/10.1001/jama.2020.4940.
- 198. Teofili L, Landolfi R, Cingolani A, Antinori A, Vecchiet J, Sanguinetti M, et al. Early transfusion of convalescent plasma in older patients with COVID-19 to prevent disease progression: a structured summary of a study protocol for a randomised controlled trial. Trials 2020; 21(1): 875. https://doi.org/10.1186/s13063-020-04821-1.
- 199. Mukherjee A, Agarwal A. Study to assess the efficacy and safety of convalescent plasma in moderate COVID-19 disease. Clin Trials Regist India 2020: 1–4.
- 200. AlQahtani M, Abdulrahman A, Almadani A, Alali SY, Al Zamrooni AM, Hejab AH, et al. Randomized controlled trial of convalescent plasma therapy against standard therapy in patients with severe COVID-19 disease. Sci Rep 2021; 11(1). https://doi.org/10.1038/s41598-021-89444-5. 9927—9927.
- 201. Avendaño-Solá C, Ramos-Martínez A, Muñez-Rubio E, Ruiz-Antorán B, Malo de Molina R, Torres F, et al. A multicenter randomized open-label clinical trial for convalescent plasma in patients hospitalized with COVID-19 pneumonia. J Clin Investig 2021; 131(20):e152740. https://doi.org/10.1172/JCI152740.
- 202. Bajpai M, Kumar S, Maheshwari A, Chhabra K, kale P, Gupta A, et al. Efficacy of convalescent plasma therapy compared to fresh frozen plasma in severely ill COVID-19 patients: a pilot randomized controlled trial. medRxiv 2020. https://doi.org/10.1101/2020.10.25.20219337.
- 203. Ray Y, Paul SR, Bandopadhyay P, D'Rozario R, Sarif J, Raychaudhuri D, et al. A phase 2 single center open label randomised control trial for convalescent plasma therapy in patients with severe COVID-19. Nat Commun 2022; 13(1): 1–9.
- 204. Gonzalez JLB, González Gámez M, Mendoza Enciso EA, Esparza Maldonado RJ, Palacios DH, Campos SD, et al. Efficacy and safety of convalescent plasma and intravenous immunoglobulin in critically ill COVID-19 patients. A controlled clinical trial. medRxiv 2021. https://doi.org/10.1101/2021.03.28.21254507.
- Körper S, Weiss M, Zickler D, Wiesmann T, Zacharowski K, Corman VM, et al. Results of the CAPSID randomized trial for high-dose convalescent plasma in patients with severe COVID-19. J Clin Investig 2021; (20): 131. https://doi.org/10.1172/jci152264.
- 206. O'Donnell MR, Grinsztejn B, Cummings MJ, Justman JE, Lamb MR, Eckhardt CM, et al. A randomized double-blind controlled trial of convalescent plasma in adults with severe COVID-19. J Clin Investig 2021; 131(13).
- 207. Rasheed AM, Fatak DF, Hashim HA, Maulood MF, Kabah KK, Almusawi YA, et al. The therapeutic effectiveness of Convalescent plasma therapy on treating COVID-19 patients residing in respiratory care units in Baghdad, Iraq. medRxiv 2020. https://doi.org/10.1101/2020.06.24.20121905.
- 208. Simonovich VA, Burgos Pratx LD, Scibona P, Beruto MV, Vallone MG, Vázquez C, et al. A randomized trial of convalescent plasma in covid-19 severe pneumonia. N Engl J Med 2020; 384(7): 619–629. https://doi.org/10.1056/NEJMoa2031304.

- 209. Axfors C, Janiaud P, Schmitt AM, van't Hooft J, Smith ER, Haber NA, et al. Association between convalescent plasma treatment and mortality in COVID-19: a collaborative systematic review and meta-analysis of randomized clinical trials. BMC Infect Dis 2021; 21(1): 1–23.
- 210. Averyanov Alexander V, Baklaushev Vladimir P. Hyperimmune convalescent plasma in moderate and severe COVID-19 disease. ClinicalTrials; 2020. Available from, https://www.clinicaltrials.gov/ct2/show/record/NCT04392414? term=NCT04392414&draw=2&rank=1.
- 211. Cabrera JMA. Convalescent plasma for patients with COVID-19: a randomized, single blinded, parallel, controlled clinical study (CP-COVID-19). ClinicalTrials; 2020. Available from, https://www.clinicaltrials.gov/ct2/show/record/ NCT04332835?term=NCT04332835&draw=2&rank=1.
- 212. Andalusian Network for Design and Translation of Advanced Therapies. Clinical trial to evaluate the efficacy of treatment with hyperimmune plasma obtained from convalescent antibodies of COVID-19 infection. ClinicalTrials; 2020. Available from, https://www.clinicaltrials.gov/ct2/show/study/NCT04366245? term=NCT04366245&draw=2&rank=1.
- 213. Self WH, Stewart TG, Wheeler AP, El Atrouni W, Bistran-Hall AJ, Casey JD, et al. Passive Immunity Trial for Our Nation (PassITON): study protocol for a randomized placebo-control clinical trial evaluating COVID-19 convalescent plasma in hospitalized adults. Res Sq 2021. https://doi.org/10.21203/rs.3.rs-227796/v1.
- 214. Shoham S, Bloch EM, Casadevall A, Hanley D, Lau B, Gebo K, et al. Randomized controlled trial transfusing convalescent plasma as post-exposure prophylaxis against SARS-CoV-2 infection. medRxiv 2021. https://doi.org/10.1101/2021.12.13.21267611.
- Ortigoza MB, Yoon H, Goldfeld KS, Troxel AB, Daily JP, Wu Y, et al. Efficacy and safety of COVID-19 convalescent plasma in hospitalized patients: a randomized clinical trial. JAMA Intern Med 2022; 182(2): 115–126. https://doi.org/10.1001/jamainternmed.2021.6850.
- Sullivan DJ, Gebo KA, Shoham S, Bloch EM, Lau B, Shenoy AG, et al. Early outpatient treatment for covid-19 with convalescent plasma. N Engl J Med 2022. https://doi.org/10.1056/NEJMoa2119657.
- 217. Pouladzadeh M, Safdarian M, Eshghi P, Abolghasemi H, Bavani AG, Sheibani B, et al. A randomized clinical trial evaluating the immunomodulatory effect of convalescent plasma on COVID-19-related cytokine storm. Intern Emerg Med 2021; 16(8): 2181–2191. https://doi.org/10.1007/s11739-021-02734-8.
- 218. RECOVERY Collaborative Group. Convalescent plasma in patients admitted to hospital with COVID-19 (RECOVERY): a randomised controlled, open-label, platform trial. Lancet 2021; 397(10289): 2049–2059. https://doi.org/10.1016/s0140-6736(21)00897-7.
- 219. Writing Committee for the REMAP-CAP Investigators, Estcourt LJ, Turgeon AF, McQuilten ZK, McVerry BJ, Al-Beidh F, et al. Effect of convalescent plasma on organ support-free days in critically ill patients with COVID-19: a randomized clinical trial. JAMA 2021; 326(17): 1690-1702. https://doi.org/10.1001/jama.2021.18178.
- Investigators TR-C, Estcourt LJ. Convalescent plasma in critically ill patients with covid-19. medRxiv 2021. https://doi.org/10.1101/2021.06.11.21258760.
- 221. David H, Muljono I, Idaian Sri, Fajarwati Tetra. *Convalescent plasma as adjunct therapy for COVID-19 (PlaSenTer)*. ClinicalTrials; 2021. Available from, https://clinicaltrials.gov/ct2/show/record/NCT04873414?
 term=NCT04873414&draw=2&rank=1.
- 222. Bennett-Guerrero E. Convalescent plasma vs. standard plasma for COVID-19. ClinicalTrials; 2021. Available from, https://

- www.clinicaltrials.gov/ct2/show/NCT04344535? term=NCT04344535&draw=2&rank=1.
- 223. Gharbharan A, Jordans CCE, Geurtsvankessel C, den Hollander JG, Karim F, Mollema FPN, et al. Convalescent Plasma for COVID-19. A randomized clinical trial. medRxiv 2020. https://doi.org/10.1101/2020.07.01.20139857.
- 224. Li L, Zhang W, Hu Y, Tong X, Zheng S, Yang J, et al. Effect of convalescent plasma therapy on time to clinical improvement in patients with severe and life-threatening COVID-19: a randomized clinical trial. **JAMA 2020**; 324(5): 460–470. https://doi.org/10.1001/jama.2020.10044.
- 225. Libster R, Pérez Marc G, Wappner D, Coviello S, Bianchi A, Braem V, et al. Early high-titer plasma therapy to prevent severe covid-19 in older adults. N Engl J Med 2021; 384(7): 610–618. https://doi.org/10.1056/NEJMoa2033700.
- Giurgea LT, Morens DM. Great expectations of COVID-19 herd immunity. mBio 2022; 13(1). e03495-03421.
- Hinman AR, Orenstein WA, Papania MJ. Evolution of measles elimination strategies in the United States. J Infect Dis 2004; 189(Suppl. 1): S17—S22. https://doi.org/10.1086/377694.
- 228. Hedrich AW. Monthly estimates of the child population "susceptible' to measles, 1900–1931, Baltimore, MD. Am J Epidemiol 1933; 17(3): 613–636. https://doi.org/10.1093/oxfordjournals.aje.a117929.
- Fine P, Eames K, Heymann DL. "Herd immunity": a rough guide. Clin Infect Dis 2011; 52(7): 911–916. https://doi.org/10.1093/cid/cir007.
- Heymann DL. Control of communicable diseases manual. American Public Health Association; 2008.
- 231. Prajapati S, Narasimha Kumar GV. Assumption of herd immunity against COVID-19: a plausibility and hope or a terrible thought in modern-day to save the life. J Infect Dis 2020; 6: 147. https://doi.org/10.23937/2474-3658/1510147.
- 232. Foppa IM, Diekmann O, Heesterbeek J, and Metz JA (1991) and Van den Driessche P and Watmough J (2002). The spread of infectious diseases in heterogeneous populations. In: Foppa IM, editor. A historical introduction to mathematical modeling of infectious diseases. Boston: Academic Press; 2017. pp. 157–194.
- 233. Achaiah NC, Subbarajasetty SB, Shetty RM. R(0) and R(e) of COVID-19: can we predict when the pandemic outbreak will be contained? **Indian J Crit Care Med 2020**; 24(11): 1125—1127. https://doi.org/10.5005/jp-journals-10071-23649.
- 234. Delamater P, Street E, Leslie T, Yang YT, Jacobsen K. Complexity of the basic reproduction number (R₀). Emerg Infect Dis J 2019; 25(1): 1. https://doi.org/10.3201/eid2501.171901.
- 235. Pellis L, Ball F, Trapman P. Reproduction numbers for epidemic models with households and other social structures. I. Definition and calculation of R0. Math Biosci 2012; 235(1): 85–97. https://doi.org/10.1016/j.mbs.2011.10.009.
- Heffernan JM, Smith RJ, Wahl LM. Perspectives on the basic reproductive ratio. J R Soc Interface 2005; 2(4): 281–293. https://doi.org/10.1098/rsif.2005.0042.
- Roberts MG. The pluses and minuses of R0. J R Soc Interface
 2007; 4(16): 949–961. https://doi.org/10.1098/rsif.2007.1031.
- 238. Chowell G, Nishiura H. Transmission dynamics and control of Ebola virus disease (EVD): a review. **BMC Med 2014**; 12: 196. https://doi.org/10.1186/s12916-014-0196-0.
- 239. Fine PE. Herd immunity: history, theory, practice. **Epidemiol Rev 1993**; 15(2): 265–302. https://doi.org/10.1093/oxfordjournals.epirev.a036121.
- Van Seventer JM, Hochberg NS. Principles of infectious diseases: transmission, diagnosis, prevention, and control. International encyclopedia of public health; 2017. p. 22.
- 241. Sy KTL, White LF, Nichols BE. Population density and basic reproductive number of COVID-19 across United States counties. PLoS One 2021; 16(4):e0249271.

- Locatelli I, Trächsel B, Rousson V. Estimating the basic reproduction number for COVID-19 in Western Europe. PLoS One 2021; 16(3):e0248731.
- 243. Zhao S, Lin Q, Ran J, Musa SS, Yang G, Wang W, et al. Preliminary estimation of the basic reproduction number of novel coronavirus (2019-nCoV) in China, from 2019 to 2020: a data-driven analysis in the early phase of the outbreak. Int J Infect Dis 2020: 92: 214–217.
- 244. Li Q, Guan X, Wu P, Wang X, Zhou L, Tong Y, et al. Early transmission dynamics in Wuhan, China, of novel coronavirus-infected pneumonia. N Engl J Med 2020; 382(13): 1199–1207. https://doi.org/10.1056/NEJMoa2001316.
- Mahase E. Covid-19: what is the R number? BMJ 2020; 369: m1891. https://doi.org/10.1136/bmj.m1891.
- 246. Lippi G, Henry BM. How will emerging SARS-CoV-2 variants impact herd immunity? **Ann Transl Med 2021**; 9(7): 585. https://doi.org/10.21037/atm-21-893.
- 247. Liu Y, Rocklöv J. The reproductive number of the delta variant of SARS-CoV-2 is far higher compared to the ancestral SARS-CoV-2 virus. J Travel Med 2021; 28(7): taab124. https://doi.org/10.1093/jtm/taab124.
- 248. Del Rio C, Malani PN, Omer SB. Confronting the delta variant of SARS-CoV-2, summer 2021. JAMA 2021; 326(11): 1001–1002. https://doi.org/10.1001/jama.2021.14811.
- 249. Nishiura H, Ito K, Anzai A, Kobayashi T, Piantham C, Rodríguez-Morales AJ. Relative reproduction number of SARS-CoV-2 Omicron (B. 1.1. 529) compared with Delta variant in South Africa. J Clin Med 2021; 11(1): 30. https://doi.org/10.3390/jcm11010030.
- 250. Petrie JG, Bazzi LA, McDermott AB, Follmann D, Esposito D, Hatcher C, et al. Coronavirus occurrence in the household influenza vaccine evaluation (HIVE) cohort of Michigan households: reinfection frequency and serologic responses to seasonal and severe acute respiratory syndrome coronaviruses. J Infect Dis 2021; 224(1): 49-59. https://doi.org/10.1093/infdis/jiab161.
- 251. Memoli MJ, Han A, Walters K-A, Czajkowski L, Reed S, Athota R, et al. Influenza A reinfection in sequential human challenge: implications for protective immunity and "universal" vaccine development. Clin Infect Dis 2020; 70(5): 748–753. https://doi.org/10.1093/cid/ciz281.
- 252. Hayman DTS. Measles vaccination in an increasingly immunized and developed world. **Hum Vaccin Immunother 2019**; 15(1): 28–33. https://doi.org/10.1080/21645515.2018.1517074.
- 253. Global measles and rubella strategic plan: 2012-2020 Geneva, Switzerland. World Health Organization; 2012. Available from: https://apps.who.int/iris/rest/bitstreams/53400/retrieve.
- 254. Haque A, Pant AB. Efforts at COVID-19 vaccine development: challenges and successes. Vaccines 2020; 8(4): 739.
- 255. Neagu M. The bumpy road to achieve herd immunity in COVID-19. J Immunoassay Immunochem 2020; 41(6): 928– 945. https://doi.org/10.1080/15321819.2020.1833919.
- 256. Khoury DS, Cromer D, Reynaldi A, Schlub TE, Wheatley AK, Juno JA, et al. Neutralizing antibody levels are highly predictive of immune protection from symptomatic SARS-CoV-2 infection. Nat Med 2021; 27(7): 1205–1211.
- 257. Liu Y, Liu J, Xia H, Zhang X, Fontes-Garfias CR, Swanson KA, et al. Neutralizing activity of BNT162b2-elicited serum. N Engl J Med 2021; 384(15): 1466–1468.
- 258. Planas D, Veyer D, Baidaliuk A, Staropoli I, Guivel-Benhassine F, Rajah MM, et al. Reduced sensitivity of SARS-CoV-2 variant Delta to antibody neutralization. Nature 2021; 596(7871): 276–280.
- 259. Sadoff J, Gray G, Vandebosch A, Cárdenas V, Shukarev G, Grinsztejn B, et al. Safety and efficacy of single-dose Ad26. COV2. S vaccine against Covid-19. N Engl J Med 2021; 384(23): 2187–2201.

- 260. Voysey M, Clemens SAC, Madhi SA, Weckx LY, Folegatti PM, Aley PK, et al. Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. Lancet 2021; 397(10269): 99–111.
- 261. Schmidt F, Muecksch F, Weisblum Y, Da Silva J, Bednarski E, Cho A, et al. Plasma neutralization of the SARS-CoV-2 Omicron variant. N Engl J Med 2022; 386(6): 599–601.
- Steenhuysen J. Analysis: how Omicron highlights fading hope of herd immunity from COVID. REUTERS; 2022. Available from: https://www.reuters.com/business/healthcare-pharmaceuticals/how-omicron-highlights-fading-hope-herd-immunity-covid-2022-01-20/.
- 263. Pulliam JRC, van Schalkwyk C, Govender N, von Gottberg A, Cohen C, Groome MJ, et al. Increased risk of SARS-CoV-2 reinfection associated with emergence of the Omicron variant in South Africa. medRxiv 2021. https://doi.org/10.1101/2021.11.11.21266068.
- Collie S, Champion J, Moultrie H, Bekker LG, Gray G. Effectiveness of BNT162b2 vaccine against omicron variant in South Africa. N Engl J Med 2022; 386(5): 494–496. https://doi.org/10.1056/NEJMc2119270.
- UK Health Security Agency. COVID-19 vaccine surveillance report. United Kingdom; 2022. Report No.: GOV-11969 Contract No.: GOV-11969.
- 266. Haas EJ, McLaughlin JM, Khan F, Angulo FJ, Anis E, Lipsitch M, et al. Infections, hospitalisations, and deaths averted via a nationwide vaccination campaign using the Pfizer-BioNTech BNT162b2 mRNA COVID-19 vaccine in Israel: a retrospective surveillance study. Lancet Infect Dis 2022; 22(3): 357–366. https://doi.org/10.1016/s1473-3099(21)00566-1.
- 267. Cabezas C, Coma E, Mora-Fernandez N, Li X, Martinez-Marcos M, Fina F, et al. Associations of BNT162b2 vaccination with SARS-CoV-2 infection and hospital admission and death with covid-19 in nursing homes and healthcare workers in Catalonia: prospective cohort study. Bmj 2021; 374: n1868. https://doi.org/10.1136/bmj.n1868.
- Levin EG, Lustig Y, Cohen C, Fluss R, Indenbaum V, Amit S, et al. Waning immune humoral response to BNT162b2 covid-19 vaccine over 6 months. N Engl J Med 2021; 385(24): e84. https://doi.org/10.1056/NEJMoa2114583.
- Shekhar R, Garg I, Pal S, Kottewar S, Sheikh AB. COVID-19 vaccine booster: to boost or not to boost. Infect Dis Rep 2021; 13(4): 924–929. https://doi.org/10.3390/idr13040084.
- 270. Ali H, Alahmad B, Al-Shammari AA, Alterki A, Hammad M, Cherian P, et al. Previous COVID-19 infection and antibody levels after vaccination. Front Public Health 2021; 9. https://doi.org/10.3389/fpubh.2021.778243.
- 271. Andrews N, Stowe J, Kirsebom F, Gower C, Toffa S, Sachdeva R, et al. Effectiveness of COVID-19 booster vaccines against COVID-19-related symptoms, hospitalization and death in England. **Nat Med 2022**; 28(4): 831–837. https://doi.org/10.1038/s41591-022-01699-1.
- 272. Mahase E. COVID-19: booster vaccine gives "significant increased protection" in over 50s. BMJ 2021; 375:n2814. https://doi.org/10.1136/bmj.n2814.
- 273. Chemaitelly H, Tang P, Hasan MR, AlMukdad S, Yassine HM, Benslimane FM, et al. Waning of BNT162b2 vaccine protection against SARS-CoV-2 infection in Qatar. N Engl J Med 2021; 385(24): e83.
- 274. Chemaitelly H, Yassine HM, Benslimane FM, Al Khatib HA, Tang P, Hasan MR, et al. mRNA-1273 COVID-19 vaccine effectiveness against the B. 1.1. 7 and B. 1.351 variants and severe COVID-19 disease in Qatar. Nat Med 2021; 27(9): 1614–1621.
- 275. Tang P, Hasan MR, Chemaitelly H, Yassine HM, Benslimane FM, Al Khatib HA, et al. BNT162b2 and mRNA-

- 1273 COVID-19 vaccine effectiveness against the SARS-CoV-2 Delta variant in Qatar. **Nat Med 2021**; 27(12): 2136–2143.
- 276. Cohn BA, Cirillo PM, Murphy CC, Krigbaum NY, Wallace AW. SARS-CoV-2 vaccine protection and deaths among US veterans during 2021. Science 2022; 375(6578): 331–336
- 277. Barda N, Dagan N, Cohen C, Hernán MA, Lipsitch M, Kohane IS, et al. Effectiveness of a third dose of the BNT162b2 mRNA COVID-19 vaccine for preventing severe outcomes in Israel: an observational study. Lancet 2021; 398(10316): 2093–2100.
- 278. Bar-On YM, Goldberg Y, Mandel M, Bodenheimer O, Freedman L, Kalkstein N, et al. Protection of BNT162b2 vaccine booster against Covid-19 in Israel. N Engl J Med 2021; 385(15): 1393–1400.
- 279. Munro AP, Janani L, Cornelius V, Aley PK, Babbage G, Baxter D, et al. Safety and immunogenicity of seven COVID-19 vaccines as a third dose (booster) following two doses of ChAdOx1 nCov-19 or BNT162b2 in the UK (COV-BOOST): a blinded, multicentre, randomised, controlled, phase 2 trial. Lancet 2021; 398(10318): 2258–2276.
- 280. Menni C, May A, Polidori L, Louca P, Wolf J, Capdevila J, et al. COVID-19 vaccine waning and effectiveness and side-effects of boosters: a prospective community study from the ZOE COVID Study. **Lancet Infect Dis 2022**; 22(7): 1002—1010. https://doi.org/10.1016/S1473-3099(22)00146-3.
- 281. Grewal R, Kitchen SA, Nguyen L, Buchan SA, Wilson SE, Costa AP, et al. Effectiveness of a fourth dose of covid-19 mRNA vaccine against the omicron variant among long term care residents in Ontario, Canada: test negative design study. BMJ 2022; 378:e071502. https://doi.org/10.1136/bmj-2022-071502.
- 282. Madewell ZJ, Yang Y, Longini IM, Halloran ME, Dean NE. Household transmission of SARS-CoV-2: a systematic review and meta-analysis. JAMA Netw Open 2020; 3(12). e2031756 e2031756.
- 283. Wilmes P, Zimmer J, Schulz J, Glod F, Veiber L, Mombaerts L, et al. SARS-CoV-2 transmission risk from asymptomatic carriers: results from a mass screening programme in Luxembourg. Lancet Reg Health Eur 2021; 4: 100056.
- 284. Angel Y, Spitzer A, Henig O, Saiag E, Sprecher E, Padova H, et al. Association between vaccination with BNT162b2 and incidence of symptomatic and asymptomatic SARS-CoV-2 infections among health care workers. JAMA 2021; 325(24): 2457–2465
- 285. El Sahly HM, Baden LR, Essink B, Doblecki-Lewis S, Martin JM, Anderson EJ, et al. Efficacy of the mRNA-1273 SARS-CoV-2 vaccine at completion of blinded phase. N Engl J Med 2021; 385(19): 1774–1785.
- 286. Giurgea LT, Han A, Memoli MJ. Universal coronavirus vaccines: the time to start is now. Npj Vaccines 2020; 5(1): 1-3.
- 287. MU Health Care. COVID-19 vaccine key to reaching 'herd immunity'. Health Care; 2022. Available from: https://www.muhealth.org/our-stories/covid-19-vaccine-key-reaching-herd-immunity
- 288. Batisse D, Benech N, Botelho-Nevers E, Bouiller K, Collarino R, Conrad A, et al. Clinical recurrences of COVID-19 symptoms after recovery: viral relapse, reinfection or inflammatory rebound? J Infect 2020. https://doi.org/10.1016/j.jinf.2020.06.073.
- 289. Chen D, Xu W, Lei Z, Huang Z, Liu J, Gao Z, et al. Recurrence of positive SARS-CoV-2 RNA in COVID-19: a case report. Int J Infect Dis 2020; 93: 297–299. https://doi.org/10.1016/j.ijid.2020.03.003.
- 290. Fu W, Chen Q, Wang T. Letter to the Editor: three cases of redetectable positive SARS-CoV-2 RNA in recovered

- COVID-19 patients with antibodies. J Med Virol 2020. https://doi.org/10.1002/jmv.25968.
- 291. Smith J. South Korea reports more recovered coronavirus patients testing positive again. Reuters; 2020. Available from: https://www.reuters.com/article/us-health-coronavirus-southkorea/south-korea-reports-more-recovered-coronavirus-patients-testing-positive-again-idUSKCN21V0JQ.
- 292. Kayat S. Doctor's note: can the coronavirus reactivate?; 2020. Available from: https://www.aljazeera.com/indepth/features/doctor-note-coronavirus-reactivate-200412062905537.html.
- 293. Ravioli S, Ochsner H, Lindner G. Reactivation of COVID-19 pneumonia: a report of two cases. J Infect 2020; 81(2): e72–e73. https://doi.org/10.1016/j.jinf.2020.05.008.
- 294. Ye G, Pan Z, Pan Y, Deng Q, Chen L, Li J, et al. Clinical characteristics of severe acute respiratory syndrome coronavirus 2 reactivation. J Infect 2020; 80(5): e14—e17. https://doi.org/10.1016/j.jinf.2020.03.001.
- 295. Loconsole D, Passerini F, Palmieri VO, Centrone F, Sallustio A, Pugliese S, et al. Recurrence of COVID-19 after recovery: a case report from Italy. Infection 2020. https://doi.org/10.1007/s15010-020-01444-1.
- 296. Bao L, Deng W, Gao H, Xiao C, Liu J, Xue J, et al. Lack of reinfection in rhesus macaques infected with SARS-CoV-2. bioRxiv 2020. <u>https://doi.org/10.1101/2020.</u> 03.13.990226.
- 297. Seow J, Graham C, Merrick B, Acors S, Steel KJA, Hemmings O, et al. Longitudinal evaluation and decline of antibody responses in SARS-CoV-2 infection. medRxiv 2020. https://doi.org/10.1101/2020.07.09.20148429.
- 298. Lumley SF, O'Donnell D, Stoesser NE, Matthews PC, Howarth A, Hatch SB, et al. Antibody status and incidence of SARS-CoV-2 infection in health care workers. N Engl J Med 2020; 384(6): 533-540. https://doi.org/10.1056/NEJMoa2034545.
- 299. Bauer G. The potential significance of high avidity immunoglobulin G (IgG) for protective immunity towards SARS-CoV-2. Int J Infect Dis 2021; 106: 61-64.
- 300. Struck F, Schreiner P, Staschik E, Wochinz-Richter K, Schulz S, Soutschek E, et al. Vaccination versus infection with SARS-CoV-2: establishment of a high avidity IgG response versus incomplete avidity maturation. J Med Virol 2021; 93(12): 6765–6777.
- Rivelli A, Fitzpatrick V, Blair C, Copeland K, Richards J. Incidence of COVID-19 reinfection among Midwestern healthcare employees. PLoS One 2022; 17(1):e0262164.
- 302. Rubin D, Eisen M, Collins S, Pennington JW, Wang X, Coffin S. SARS-CoV-2 infection in public school district employees following a district-wide vaccination program—Philadelphia County, Pennsylvania, March 21—April 23, 2021. Morb Mortal Wkly Rep 2021; 70(30): 1040.
- 303. Abu-Raddad LJ, Chemaitelly H, Ayoub HH, Yassine HM, Benslimane FM, Al Khatib HA, et al. Protection afforded by the BNT162b2 and mRNA-1273 COVID-19 vaccines in fully vaccinated cohorts with and without prior infection. medRxiv 2021. https://doi.org/10.1101/2021.07.25.21261093.
- 304. Bergwerk M, Gonen T, Lustig Y, Amit S, Lipsitch M, Cohen C, et al. Covid-19 breakthrough infections in vaccinated health care workers. N Engl J Med 2021; 385(16): 1474—1484.
- 305. CDC COVID-19 Vaccine Breakthrough Case Investigations Team. COVID-19 vaccine breakthrough infections reported to CDC—United States, January 1—April 30, 2021. Morb Mortal Wkly Rep 2021; 70(21): 792.
- 306. Cavanaugh AM, Spicer KB, Thoroughman D, Glick C, Winter K. Reduced risk of reinfection with SARS-CoV-2 after

- COVID-19 vaccination—Kentucky, May—June 2021. Morb Mortal Wkly Rep 2021; 70(32): 1081.
- 307. Reynolds CJ, Pade C, Gibbons JM, Butler DK, Otter AD, Menacho K, et al. Prior SARS-CoV-2 infection rescues B and T cell responses to variants after first vaccine dose. Science 2021; 372(6549): 1418–1423.
- 308. Stamatatos L, Czartoski J, Wan Y-H, Homad LJ, Rubin V, Glantz H, et al. mRNA vaccination boosts cross-variant neutralizing antibodies elicited by SARS-CoV-2 infection. Science 2021; 372(6549): 1413–1418.
- 309. Roberts AT, Piani F, Longo B, Andreini R, Meini S. Reinfection of SARS-CoV-2—analysis of 23 cases from the literature. Infect Dis 2021; 53(7): 479–485.
- 310. Tan Y, Liu F, Xu X, Ling Y, Huang W, Zhu Z, et al. Durability of neutralizing antibodies and T-cell response post SARS-CoV-2 infection. Front Med 2020: 14(6): 746-751.
- Larson D, Brodniak SL, Voegtly LJ, Cer RZ, Glang LA, Malagon FJ, et al. A case of early reinfection with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Clin Infect Dis 2020; 73(9): e2827—e2828. https://doi.org/10.1093/cid/ciaa1436.
- 312. Bernal JL, Andrews N, Gower C, Gallagher E, Simmons R, Thelwall S, et al. Effectiveness of COVID-19 vaccines against the B.1.617.2 variant. medRxiv 2021. https://doi.org/10.1101/2021.05.22.21257658.
- 313. Emary K, Golubchik T, Aley P, Ariani C, Angus B, Bibi S, et al., C.-GU consortium, A. Project, C.-VTG Oxford. Efficacy of ChAdOx1 nCoV-19 (AZD1222) vaccine against SARS-CoV-2 variant of concern 202012/01 (B. 1.1. 7): an exploratory analysis of a randomised controlled trial. Lancet 2021; 397: 1351–1362.
- 314. Brown CM, Vostok J, Johnson H, Burns M, Gharpure R, Sami S, et al. Outbreak of SARS-CoV-2 infections, including COVID-19 vaccine breakthrough infections, associated with large public gatherings—Barnstable County, Massachusetts, July 2021. Morb Mortal Wkly Rep 2021; 70(31): 1059.
- 315. MacIntyre CR, Chughtai AA. Recurrence and reinfection a new paradigm for the management of Ebola virus disease. Int J Infect Dis 2016; 43: 58—61. https://doi.org/10.1016/j.ijid.2015.12.011.
- Roy S. COVID-19 reinfection: myth or truth? SN Compr Clin Med 2020: 1-4. https://doi.org/10.1007/s42399-020-00335-8.
- 317. Transmission of SARS-CoV-2: implications for infection prevention precautions. World Health Organization; 2020. Available from: https://www.who.int/news-room/commentaries/detail/transmission-of-sars-cov-2-implications-for-infection-prevention-precautions.
- 318. To KK-W, Hung IF-N, Ip JD, Chu AW-H, Chan W-M, Tam AR, et al. Coronavirus disease 2019 (COVID-19) reinfection by a phylogenetically distinct severe acute respiratory syndrome coronavirus 2 strain confirmed by whole genome sequencing. Clin Infect Dis 2020. https://doi.org/10.1093/cid/ciaa1275.

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