

Perspective: Human Milk Composition and Related Data for National Health and Nutrition Monitoring and Related Research

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ABSTRACT

National health and nutrition monitoring is an important federal effort in the United States and Canada, and the basis for many of their nutrition and health policies. Understanding of child exposures through human milk (HM) remains out of reach due to lack of current and representative data on HM's composition and intake volume. This article provides an overview of the current national health and nutrition monitoring activities for HM-fed children, HM composition (HMC) and volume data used for exposure assessment, categories of potential measures in HM, and associated variability factors. In this Perspective, we advocate for a framework for collection and reporting of HMC data for national health and nutrition monitoring and programmatic needs, including a shared vision for a publicly available Human Milk Composition Data Repository (HMCD-R) to include essential metadata associated with HMC. HMCD-R can provide a central, integrated platform for researchers and public health officials for compiling, evaluating, and sharing HMC data. The compiled compositional and metadata in HMCD-R would provide pertinent measures of central tendency and variability and allow use of modeling techniques to approximate compositional profiles for subgroups, providing more accurate exposure assessments for purposes of monitoring and surveillance. HMC and related metadata could facilitate understanding the complexity and variability of HM composition, provide crucial data for assessment of infant and maternal nutritional needs, and inform public health policies, food and nutrition programs, and clinical practice guidelines. Adv Nutr 2022;13:2098–2114.

Statement of Significance: Our understanding of exposures through human milk (HM) in the United States and Canada remains out of reach due to lack of current and representative data on the composition and intake volume of HM. This article provides an overview of the current national monitoring activities for HM-fed children, including HM composition (HMC) data. In this Perspective, the authors advocate for a framework for collection and reporting of HM composition data for national monitoring and programmatic needs, including a shared vision for a publicly available Human Milk Composition Data Repository (HMCD-R) to include essential metadata associated with HMC.

Keywords: human milk, intake volume, breast milk, B-24 population, variability in human milk, national monitoring, environmental chemicals in human milk, bioactives in human milk, nutrients in human milk, human milk composition data repository (HMCD-R)

Introduction

Human milk (HM) is a complex biological fluid made up of many structurally and functionally diverse constituents. The impact of HM on infant growth and development goes beyond its nutritional value (1–5); however, its complexity, composition, and variability are not fully understood (5, 6). Christian et al. (7) have recommended studying HM as a biological system to better understand the interactions of factors that impact HM composition (HMC), intake volume, and the functionality of its diverse constituents for optimizing maternal and infant health. Accurate estimates of dietary exposures for HM-fed children require up-todate HMC and intake volume data; however, these estimates remain out of reach due to lack of current and nationally representative data for both HMC and intake volume in the United States and Canada (8).

A joint United States-Canada federal government undertaking, the Human Milk Composition Initiative, was initiated in 2018 to articulate how HMC-related data are relevant to both countries' federal programs, policies, and regulations (9). This article provides an overview of the current national monitoring activities for HM-fed children, potential measures in HM, food composition databases used for exposure assessments, and variability factors associated with HMC. In this Perspective, we advocate for a framework for collection and reporting of HMC data for public health and nutrition monitoring, research, and programmatic needs including a shared vision for a Human Milk Composition Data-Repository (HMCD-R), to include essential metadata associated with HMC.

Monitoring Health, Nutrition, and Environmental Chemicals in the United States and Canada

National health and nutrition monitoring programs by the federal governments in the United States and Canada are essential surveillance efforts of their populations' health. They provide nutrient intake estimates from foods, beverages, and dietary supplements and prevalence and trends of selected

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diseases, risk factors, behaviors, and environmental exposures (10). Linking these data to anthropometric, laboratory, and clinical evaluation data and health outcomes, if available, allows for examination of cross-sectional associations at the national and large subgroup population levels (11). These data are used to inform, guide, and monitor government programs and policies, including the DRIs (12), Dietary Guidelines for Americans (13, 14), Canada's Food Guide (15), food-fortification policies (e.g., iron, vitamin D, and iodine), and food labels (16), among others.

Table 1 summarizes the major federal health and nutrition monitoring efforts in the 2 countries. They vary in their objectives, degree of details, and target populations. NHANES (17–19) monitors the health and nutritional status of the US population of all ages, through interviews and physical examinations. For children aged birth to 24 mo (B-24), it collects frequency of HM consumption through 24-h dietary recalls and questions on infant feeding practices, including breastfeeding, timing of introduction/stoppage of infant formula or complementary foods and beverages, and mode of feeding HM. The Canadian Community Health Survey (CCHS) is an annual general health survey of \sim 65,000 Canadians aged >12 y (17, 20) with periodic focused surveys on topics of interest. The B-24 population is excluded from CCHS-Annual Component; however, a nutrition-focused survey conducted in 2015 (21), provided data on number of times HM is consumed for children aged 12–24 mo (n = 404) (22). Similarly, the Canadian Health Measures Survey does not include children aged <3 y (23, 24). HM samples, its composition, or volume are not collected in any of the above national surveys. However, these surveys can be potentially important vehicles in future, given their representative sampling methodologies and detailed collection of demographic and health-related data.

Environmental chemicals

Monitoring exposures of environmental chemicals during infancy is important, because it can be the period of largest exposure. Many of these chemicals accumulate within the mothers' tissues and have the potential to pass directly into HM, which can function as an excretion pathway (25, 26), as in polychlorinated biphenyls (27). HM is not included in the 2 US national surveys that monitor exposure to environmental chemicals—the FDA's Total Diet Study (28) and NHANES (18, 29).

Two programs in United States/Canada—Environmental influences on Child Health Outcomes (ECHO) and Maternal-Infant Research on Environmental Chemicals (MIREC)—were established with a focus on environmental chemicals, including in HM (Table 1). The ECHO cohort is made up of >70 individual cohorts of mothers and children from ongoing research projects, forming a massive "cohort of cohorts" of >50,000 children across the United States. HM is an ECHO "recommended data element" (i.e., ECHO strongly encourages, but does not require cohorts to collect), and ~225 samples have been collected since 2016. For the cohorts that do collect, HM is collected and

The authors reported no funding received for this study.

Author disclosures: The authors report no conflicts of interest. NA (Associate Editor) played no role in the Journal's evaluation of the manuscript.

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Supplemental Figure 1 is available from the "Supplementary data" link in the online posting of the article and from the same link in the online table of contents at

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Abbreviations used: CANLINE, Canadian Laboratory Information Network; CCHS, Canadian Community Health Survey; ECHO, Environmental influences on Child Health Outcomes; HM, human milk; HMC, human milk composition; HMCD-R, Human Milk Composition Data Repository; MIREC, Maternal-Infant Research on Environmental Chemicals; NASEM, National Academies of Sciences, Engineering, and Medicine; SR, standard reference; TEQ, toxicity equivalent factor.

TABLE 1 Major efforts for monitoring health, nutrition, and environmental chemicals in the United States and Canada¹

			Samula siza	cize	
				Birth to 24 mo	Salactad R-24 relevant
	Objectives	Target population	Total	(B-24)	beletied D-24 Televalit measurements
NHANES, an ongoing nationally representative survey in the USA, since 1971; continuous since 1999 (17, 18)	Continuous monitoring and data collection on health and nutritional status	Noninstitutionalized, civilian US population of all ages	~10,000/2-y cycle	~600-900/2-y cycle	Food, beverage, and nutrient intake, dietary supplements, anthropometric measurements, and laboratory tests. Includes data on human milk (HM) consumption (frequency) collected using 24-h dietary recalls and questions on breastfeeding history and infant feoding corrects
The Canadian Community Health Survey (CCHS), a cross-sectional survey conducted since 2000 (196) the phrase	Gathers health-related data (information on determinants of health, health status, and use of the health system) at community level	Population aged ≥12 y	~ 65,000	Not applicable	Questions on prenatal exposures (e.g., smoking, alcohol consumption, folic acid supplementation) and infant feeding behaviors (e.g., breastfeeding initiation and related behaviors/beliefs, introduction of complementaty food and beverages, vitamin D supplementation, and gestational weight gain) are administered to femen respondents (ages 15–55 y, who can object in provision 5 ty
Canadian Health Measures Survey (CHMS)—ongoing (23)	Gathers health-related information to help improve the prevention, diagnosis, and treatment of illnesses and to promote health and wellness	3–79 y	> 5000 including ~1000 women aged 15-49 y every 2 y	Not applicable	wiru gave sint in previous 5 y/ Includes anthropometric, sociodemographic, nutrition, environmental exposure, and biomarker information, among other clara
Environmental influences on Child Health Outcomes (ECHO) 2016–2022 (197)	Understanding the effects of a broad array of early environmental exposures on children's development and health outcomes	> 70 individual cohorts of mothers and children from ongoing research projects	> 50,000 children	~ 10,000	Standardized data elements such as sociodemographics, pregnancy and family history, maternal diet, caregiver environment, child's physical and neurodevelopmental health, among others. Questions related to breastfeeding include initiation of breastfeeding, duration, and whether the HM consumed was from the breast, pumped, or both. About 225 HM samples have been collected so far; none have been analyzed

			Sa	Sample size	
	Objectives	Target population	Total	Birth to 24 mo (B-24)	Selected B-24 relevant measurements
Maternal-Infant Research on Environmental Chemicals (MIREC), a longitudinal Canadian pregnancy cohort study (2008–2011) (198)	To examine potential adverse health associations on maternal and infant health with exposure to priority environmental chemicals, including those in HM	Pregnant women from 10 cities across Canada	2001	525	Maternal biospecimens were collected throughout pregnancy, along with questionnaire and clinical data, and HM after delivery. HM samples (n = 1017) in MIREC were analyzed for 188 unique environmental chemicals, in addition to several nutritional constituents

TABLE 1 (Continued)

¹B-24, birth to 24 mo; HM, human milk

stored under standardized conditions and is diverse in terms of factors, for example, urban and rural settings, ethnicity, gestational age at delivery (30). MIREC is a longitudinal Canadian pregnancy cohort study conducted between 2008 and 2011. Over 1000 HM samples were collected, along with information on the date and time, hand/pump expression, right/left breast, and timing of sampling within a feed (4).

To summarize, there is no nationwide monitoring of HM in the United States or Canada. HM samples were collected for study of environmental chemicals over a decade ago in MIREC, and limited HM samples are being collected in the ECHO program, to be useful for current national estimates. However, these 2 programs can provide insights for developing a framework for collection of data on HMC for public health nutrition research and informing policies.

Human Milk Volume

Quantifying HM intake volumes is crucial to determining exposure estimates. Researchers have mainly relied on estimating volume based on a child's age, exclusive or partially fed HM, and volume of other types of milk or infant formula consumed. These calculations (Table 2) were first developed for the Feeding Infants and Toddlers Study (31) and have since been used for NHANES (17, 32, 33), and DRIs (25). Problematically, these assumptions and calculations are based on limited data from 3 studies from the 1980s and 1990s, with a limited number of subjects (n = 8-46) from the United States and Australia (34-36). Furthermore, the approach of imputing standard volumes does not consider variability in intake volume. Many health outcomes of HM intake could be dose-dependent (37); the range of intake is large and potentially affected by several physiological and lifestyle factors related to the mother and child (discussed later). Hence, there is a demonstrable need for measurement of HM intake volumes of US and Canadian children to determine accurate exposure estimates. Furthermore, many studies estimate HM intake by weighing infants before and after feeding, a method susceptible to human and mechanical errors, and requiring further corrections to account for insensible fluid loss (38). The use of the deuterium oxide $(^{2}H_{2}O)$ technique for quantifying HM intake has also been used and was reviewed by da Costa et al. (39). In pooled data from 12 countries, they reported an average daily HM intake of \sim 600 mL/d during the first month of life, rising to \sim 820 mL/d at 3-4 mo before declining gradually at 8-9 mo-higher than in previous reports, including those used for NHANES and DRIs (39).

Food Composition Databases and Human Milk Composition Data in the United States and Canada

Food composition databases can contain traditional nutritive (macronutrients, vitamins, minerals, fatty acids, and amino acids) and bioactive components (e.g., carotenoids, flavonoids, glucosinolates), among others. They provide the foundation for food and nutrition research, dietary practice, and analysis of dietary studies, essential to the development **TABLE 2** Human milk intake volume estimates for exclusively HM-fed infants (43–48) currently used for exposure assessment in the United States and Canada¹

	Infants (0–6 mo)	Infants (>6 mo)	Young children (12–24 mo)
Daily intake (mean)	780 mL	600 mL	N/A (varies)
Intake per feed	_	—	89 mL (12–17 mo); 59 mL (18–24 mo)

¹For infants partially fed HM, the volume of infant formula plus "other" milk reported consumed on the recall day are subtracted from the age-specific reference volume to estimate HM consumed.

of food and nutrition policies, guidelines, regulations, food and nutrition programs, and risk assessments (29, 40–42).

The USDA Nutrient Database for Standard Reference (SR) (a legacy database since 2018) has served as the primary source of food composition data for most databases in the United States, including for NHANES. The USDA FoodData Central, launched in 2019, is an integrated data system that includes SR and other USDA food composition databases and adds new databases—"Foundation Foods" and "Experimental Foods"—that have nutrient information and extensive underlying metadata that will help users understand nutrient variability (43). However, the limited existing data for HM are from the late 1970s (original data source unknown and no information on sampling, storage, or laboratory analysis). It is recommended by USDA that these data are not used and should be replaced (44).

The Canadian Nutrient File was initiated in 1979 to support nutrition surveys and other needs of Canadian public health agencies, the food industry, hospitals, and universities (45). The public version of the Canadian Nutrient File (2015) reports only 1 nutrient profile for HM, which is primarily sourced from SR with a few *trans*-fatty acids analyzed in Canada. A separate searchable food data repository, the Canadian Laboratory Information Network (CANLINE) (46), contains chemical, nutritional, and microbial surveillance data, food and collection metadata but no data on HM.

In 2017, USDA conducted a review of the existing literature to summarize current knowledge on macro- and micronutrients and update the HMC data in its database (47). They identified 28 studies conducted in the United States/Canada on macro- and micronutrient content of HM over 37 y (1980-2017); most were published before 1990 with relatively small sample sizes and limited generalizability due to different sampling, storage, and analytic methods. The reviewers underlined the need for comprehensive studies to provide current and complete nutrient information on HM in the United States and to fully understand the magnitude of variability in composition. A joint federal workshop sponsored by the NIH in 2017 on "Human Milk Composition - Biological, Environmental, Nutritional, and Methodological Considerations" also emphasized the need to fill these gaps in availability of information on HMC, factors that influence HMC, and their relation to maternal and infant health (9). In 2020, the National Academies of Sciences, Engineering, and Medicine (NASEM), scanned the existing literature and identified \sim 126 studies on HMC and volume to inform DRIs. The expert group noted the lack of data for several nutritive components and HMC after 6 mo postpartum, and inconsistency in sampling, HM collection, and analytical methods (5).

Potential Measures in Human Milk

Potential measures in HM can be categorized into several groups, based on structural similarity (**Figure 1** lists examples, and **Supplemental Figure 1** provides details on sources used). A brief overview of some of the major categories follows.

Carbohydrates

Potential measures of carbohydrate components of HM include several types of saccharides. Lactose is the predominant sugar, followed by bioactive HM oligosaccharides (nondigestible sugars) (48). Over 200 HM oligosaccharides have been elucidated (49).

Protein and nonprotein nitrogen compounds

The major protein-related categories include amino acids, glycoproteins, and other proteins/peptides (excluding glycoproteins) (50–56). HM glycoproteins are the most abundant proteins or important bioactive proteins. Proteomic studies have reported \sim 3000 proteins in HM (57, 58). Immunoglobulins, cytokines, chemokines, and growth factors contribute to the infant's immune protection and development (59), and gastrointestinal regulatory peptides help with regulation of infant appetite and feeding patterns (60). Small nitrogencontaining compounds, for example, nucleotides and melatonin, are associated with infant sleep regulation (60, 61).

Lipids

Triacylglycerides comprise \sim 98% of the lipid fraction (8). About 200 fatty acids were found in HM (62, 63) as were phospholipids, building blocks forming the membrane of the HM fat globules (64, 65). Steroids in HM, especially cholesterol and its precursors, and hormones, can influence the growth and behavior of the infant, but the studies of their presence in HM are limited (66–71). Other lipids of interest include lipid mediators, such as prostaglandins, resolvins, lipoxins, and leukotrienes (72–74).

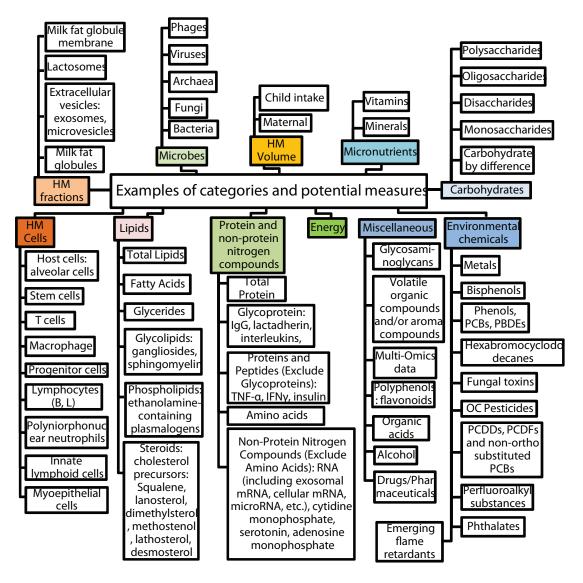


FIGURE 1 Categories of potential components to measure in human milk. HM, human milk; OC, organochlorine; PBDE, Polybrominated diphenyl ethers; PCB, polychlorinated biphenyl; PCDD, polychlorinated dibenzodioxin; PCDF, polychlorinated dibenzofuran.

Minerals and vitamins

HM includes major minerals (e.g., calcium and phosphorus), electrolytes (e.g., sodium and potassium), and trace minerals (e.g., iron, copper, and iodine) (75, 76). Vitamins include water- and fat-soluble vitamins. Dror and Allen (77) summarized the current knowledge of nutrients in HM.

Bioactives

HM is rich in a range of bioactives with diverse functions in infant health including antimicrobial factors, immune proteins, growth factors, enzymes, antioxidants, and endocannabinoids. Their presence can partially explain positive health outcomes of HM-fed infants compared with formulafed counterparts. Many bioactives are proteins (or peptides) with biological activities or carbohydrates like HM oligosaccharides. Bioactives also include living cells like maternal leukocytes and macrophages, or carotenoids, polyphenols, and vitamins (listed under different categories in Figure 1) (53, 62, 78, 79).

Microbes

Bacteria, viruses, fungi, and yeasts are present in HM and associated with the infant's present and future health (80–82). Microbiomes in HM and infant feces are related to each other, but vary by culture/location (83), lactation stage (84), and storage conditions (85). HM transmission of maternal viral infection is well established for cytomegalovirus, HIV-l, and human T-cell lymphotropic viruses (86–89). Transmission of microbes was recently reviewed by Selma-Royo et al. (90).

Environmental chemicals

Women are exposed to a complex mixture of chemicals in their daily lives through their environment, product use, house dust, food, drinking water, air, and workplace exposures; many can partition into HM (25, 26). Figure 1 lists the categories of environmental chemicals analyzed in Methods Advancement for Milk Analysis (US) and MIREC studies (91).

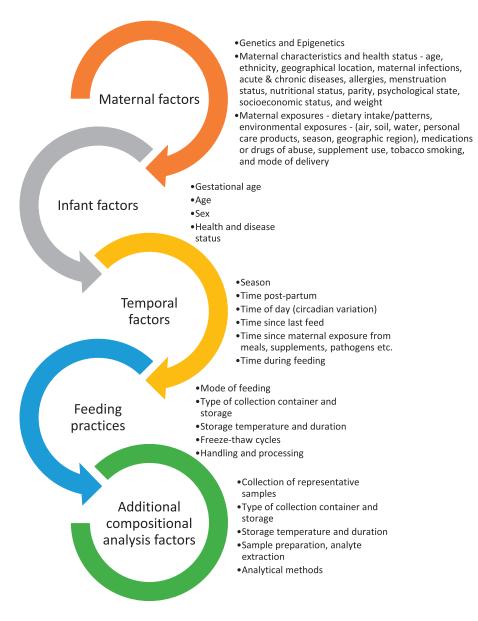


FIGURE 2 Examples of variability factors associated with human milk composition and/or volume.

Prioritizing components to measure

Given the multitude of components in HM, prioritization of the components to analyze is needed. For nutrition monitoring and related research, parameters include:

- Uncertainty or knowledge gap: Data do not exist in the United States/Canada or do exist but with limited generalizability to the US/Canadian population, for example, the HM microbiome.
- Substantial public health concern: Data would address an existing or undercharacterized public health concern and/or are important to monitor in a national sample, for example, per- and polyfluoroalkyl substances (25, 47, 92).
- Potential to inform federal programs: Data are essential to fill gaps in guidance or recommendations to help

promote health and prevent chronic disease in the United States/Canada, including but not limited to DRIs, dietary guidance, clinical practice guidelines, and regulations and standards for food and supplement manufacturers, for example, minerals and vitamins.

Variability in Human Milk Composition

HM is unique to each infant-mother dyad, characterized by variability in its composition. HMC varies both between subjects, associated with maternal and infant factors (e.g., genetics, diet, and gestational age at birth) and within subjects by factors (e.g., time postdelivery and time of day). The variability factors can generally be categorized into 4 types (**Figure 2**), examples of which are briefly discussed below, to emphasize the need for collection of metadata and characterizing factors that can impact HMC and volume to further understand their variability. There is some research on impact of variability factors on HMC of macronutrients. More research is needed to identify factors that contribute to the variation in micronutrients and bioactive components.

Maternal factors

There is a growing body of literature suggesting that several maternal characteristics relate to HMC. Genetic differences can directly impact HMC and alter relations between maternal intake/exposure and HMC (93-98). A systematic review and meta-regression of data from 66 studies reported a positive association (β : 0.56 g/L; 95% CI: 0.034, 1.1) between maternal BMI and HM fat content, but no association with total protein, lactose, or energy (99). Associations have also been reported between individual HM components and various other maternal factors, for example, maternal age, parity, mode of delivery, socioeconomic status, maternal infections, acute and chronic diseases, and allergies, albeit inconsistently (6, 100-102). More research is needed to understand the impact of these factors on HMC. A systematic review found an association between maternal chronic disease prevalence (obesity, cardiovascular disease, and type 2 diabetes mellitus) and HMC in most studies and milk volume in some (103). Hence, documenting medical histories, past and current treatments, and relevant biomarkers of prevalent chronic diseases could provide important metadata for understanding variability.

Maternal nutritional status, dietary intakes, food and behavior choices, food fortification, and supplement use can have varied influences on HMC, especially on micronutrients (53). For example, HMC of vitamin B-12 correlates with maternal intake and status and is lower in women with depleted vitamin B-12 status. In contrast, other than in an overt folate deficiency, HM folate concentration is fairly constant and independent of mother's folate status/intake (6, 77, 104). A systematic review by Bravi et al. (105) of 36 publications found consistent evidence of association only for maternal fish consumption with HM DHA content, and for maternal vitamin C intake with HM vitamin C content. Adhikari et al. (106) reported inconsistent associations between HM macronutrients and energy and maternal dietary intake and methodologies of included studies.

Infant factors

HMC varies according to gestational age at delivery (i.e., preterm compared with term) (53). A systematic review of 26 preterm studies and 30 term studies reported higher protein concentration (+7% to +35%) and lower lactose concentration (-2% to -10%) in preterm compared with term HM in the first weeks of life, with no significant differences after weeks 10–12 of lactation. It also reported significantly higher concentrations of fat, lactose, and energy (+93%, +16%, and +16%, respectively) and lower concentrations of protein (-52%) in term mature milk relative to colostrum (107). Differences in selected vitamins and minerals have been shown in individual studies (108, 109). Documenting

infant-associated factors, for example, sex (110), age (111), and medical and health status (112), could help with better characterization of HMC.

Temporal factors

HMC changes as milk matures over the course of lactation. A systematic review and meta-analysis of HM macronutrient content identified significantly higher concentrations of fat, lactose, and energy (+93%, +16%, and +16%, respectively)and lower concentrations of protein (-52%) in term mature milk relative to colostrum (107). Smaller changes in fat and protein content can continue over the course of extended lactation (113, 114). A systematic review of circadian variation in HM reported diurnal changes in concentrations of amino acids, fat, iron, and some hormones (115), and B-vitamins (116). Timing of sampling within a feed (foremilk compared with hindmilk) and time elapsed since the last feed can affect HMC, particularly fat concentrations (113, 114, 117). The time of day when HM is sampled is also important in relation to the time of maternal (and in some cases infant) exposures, for example, to foods and beverages, supplements, alcohol, medications or drugs of abuse, pathogens, tobacco use, and many environmental chemicals. Feeding frequency and duration-for example, number of feeds per 24 h, number of minutes per feed, and amount of time between feeds-are important. Time of collection within a feed has shown that foremilk and hindmilk composition varies (53, 118).

Feeding practices

This category includes mode of feeding and handling methods that can influence HMC. Breastfeeding, pumping then feeding HM by bottle, and cold storage followed by reheating of HM (freeze-thaw cycles) can lead to additions (e.g., bacterial contamination from a breast pump) or losses (e.g., degradation of nutrients after extended freezing or overheating) (6, 53, 119–125).

Several of these factors have been related to bioactives and microbes too. For example, because most HM bioactives are proteins, these are often sensitive to changes in temperature (e.g., freeze-thaw cycles and pasteurization) that induce denaturation and consequent loss of biological activity (126), or cellular components such as maternal leukocytes are highly susceptible to death/inactivation upon prolonged storage and changes in ambient temperature (127). The diet during pregnancy has a stronger impact on the microbiome compared with the diet during the first month of lactation (128). It is important to understand the factors that influence HM microbiota composition, because that is the most probable way of vertical transfer of bacteria and microbiota establishment in the infant gut.

Environmental chemicals

Maternal age (e.g., older mothers have higher lifetime exposure and bioaccumulation of environmental chemicals, especially those phased out of use/production), parity, lactation history, and duration of breastfeeding (e.g., number of months, exclusive or partial HM feeding) have been associated with HMC of environmental chemicals (25, 47). Maternal exposures to air, soil, water, food, clothes, personal care products (e.g., triclosan in toothpaste), dental amalgam (mercury), electronics, and furniture are part of the maternal environment and potential exposure sources. Geographical location (e.g., living near a point source of environmental chemicals) can lead to differential exposure (47, 129, 130). A recent systematic review based on 20 studies concluded that smoking was inversely associated with HM lipids, energy, and proteins (131). Changes in maternal exposures have the potential to change HMC over time, even daily, especially for chemicals with short halflives; thus, studies that take repeated measurements of HM give a better characterization of exposure. In addition, as a chemical is phased out of commercial use, for example, per- and polyfluoroalkyl substances-perfluorooctanoate and perfluorooctane sulfonate-by major US producers (132), its concentration in environmental media can decrease (133-135). However, replacement biosimilar environmental chemicals (e.g., new polyfluorinated chemicals) often emerge. Nutritional status and dietary supplementation can impact environmental chemical exposure (136-141). HMC following exposure to environmental chemicals can differ in mature and transitional milk (142–144).

The impact of the above factors is potentially complex and current knowledge is generally limited to impact on a single component/factor. A 2020 systematic review of collection techniques for HM research in macronutrient composition recommended that studies report and standardize details of the collection procedure (time of day of collection, collection method, collection breast, time since last expression), stage of lactation, infant gestational age, and mode of feeding (exclusive or partial breast feeding) (117). The authors noted lack of detail in the reviewed studies, limiting their use for public health purposes. Collection of metadata associated with the composition values can provide insights into each HMC data point and enhance our understanding of HM variability.

Additional Considerations for Compositional Analyses of Human Milk

A recent book (145) and several recent reviews (12, 117, 146) have provided details of best practices for collection and analysis of HM for composition research. Broadly, collection, handling, storage, and analytical protocols should be optimized for the components to be measured.

Sample collection

Study design for HMC research should accommodate HMC variability factors (summarized in Figure 2) and seek to control unintended influences on HMC. There is no consensus on the most reliable method for collecting a "representative" HM sample from the lactating mother. Collection and sampling from all HM expressed over 24 h, with multiple samples collected over time from each participant, is considered a "gold standard" method for providing representative HM samples in longitudinal, multianalyte HMC studies (53, 147,

148). This protocol minimizes within-subject variation due to time of day of sampling or time of within-feed sampling, but can be impractical for large population studies (149). In a recent systematic review of HM sampling methodologies, Leghi et al. (117) reported that weighted pooling of preand postfeed samples collected over 24 h provides similar concentrations of total fat, protein, and lactose in HM, and hence an acceptable alternative to the "gold standard." This sampling technique was accepted as an alternative to the 24-h full breast expression method for fat- and fat-soluble components in the 2020 NASEM evidence scan of HMC research (5). The NASEM committee accepted any milk type (fore-, mid-, or hindmilk) for protein and lactose, provided collection conditions were standardized, and any milk type for elements provided trace element-free supplies were used. Afternoon and evening samples are preferred for B-vitamins, because these are reported to best reflect total daily vitamin concentrations (12, 116). In addition, it is recommended that participants in studies of B-vitamins in HM should not be fasting, and that collection should not occur <4 h after supplement use. For large cohort studies, where standardization of protocols is difficult to implement, detailed data on HM collection and storage conditions can be recorded and included in statistical models to facilitate the identification of variables that influence HMC of specific analytes (150).

Handling and storage

Sample storage container type can influence HMC, either by binding analytes of interest or by introducing exogenous substances (151-153), and can differ based on the constituent (154). Fat- and fat-soluble components can adhere to the sides of storage containers, leading to loss of components with aliquoting or container changes (155-157), hence they should be minimized and care should be taken in homogenizing or aliquoting HM samples to limit component loss (158). Trace element-free supplies are recommended for HM mineral analysis to reduce the potential for contamination (12). Many components in HM are light-sensitive, for example, carotenoids, vitamin C, riboflavin, vitamin B-6, folate, and vitamin B-12 (146, 159); opaque or amber storage vials, and processing samples under limited or yellow light can avoid sample degradation (146, 160). Some HM components, for example, vitamin C, long-chain PUFAs, and riboflavin, can also undergo oxidation and degradation during storage, particularly at room temperature or in a standard refrigerator (161); hence ultra-low-temperature freezers $(-70^{\circ}C)$ are recommended for long-term HM storage (146). Storage conditions and length of storage affect the integrity of expressed HM, and are a logistical consideration in planning HMC studies (162, 163). For example, HM intended for vitamin C analysis is recommended to be stored for <1 mo because degradation can occur even at ultra-low temperatures (161, 164). Other components, for example, tocopherols, total protein, lactose, and minerals, are considered more stable over long-term storage (6, 158). Multiple freeze-thaw cycles reduce total lipids in HM, increase concentrations of lipolysis products, and change HMC of lipids (6, 165). HM samples should be aliquoted after collection to avoid unnecessary freeze-thaw cycles, particularly for quantification of lipids. Because HM is nonsterile, incubations at room or body temperature can lead to bacterial metabolism and changes in the HM microbiome, and in substrates used by or metabolites produced by bacteria (166). Protocols for handling and storing donor HM such as pasteurization reduce pathogen load but alter the HMC of multiple constituents (163), curtailing the applicability of compositional analyses of donor milk to the general population. HM collection, handling, and storage are most challenging for large, multianalyte HMC studies, but ultimately logistics must be optimized for priority analytes (or groups).

Analytical methods

HM is not a homogeneous mixture, but consists of several compartments, including true solutions, colloids (e.g., casein micelles), membranes, membrane-bound globules, and live cells (167, 168). Therefore, the sample preparation, analyte extraction, and analytical methods must be carefully selected based upon the physicochemical properties of the specific component, how they are present in HM, and their concentrations. Generally, the method that can provide the most accurate measurement of the component in its original form is desired. Wu et al. (8) discussed various analytical methods available for several macro- and micronutrients and their pros and cons, and Hampel et al. (146) listed preferred methods for micronutrient analysis. Both articles emphasized that to achieve optimal results, the entire analytical procedure must be thoroughly validated for HM (8). Furthermore, use of a certified reference material, wellcharacterized, consensus, or in-house material for method validation and quality assurance, allows comparison of analytical data conducted at different time periods, instruments, and laboratories (146, 169).

Analytical methodologies have evolved with the development of new technology and instrumentation. For instance, microbiological methods were used to analyze B-vitamins, but recently chromatographic methods coupled with UV, fluorometric, or MS detection have become the preferred methods (159). There are still many gaps in the availability of analytical methods for certain components, especially bioactives. For example, despite tremendous efforts in recent years to understand HM oligosaccharides, there is still no consensus method for quantifying them (170). Furthermore, structure-function relations exist for HM oligosaccharides, necessitating the need to quantify them separately (171).

Use of "-omics" approaches for assessing HM can add significantly to our knowledge about specific components and their variability (172, 173). Proteomic and metabolomic approaches are used extensively to characterize various HM components, such as proteins found in casein and whey fractions, HM oligosaccharides and lipids (174–177), and the microbiome (178). Similar approaches could be applied in the characterization of hormones, growth factors, cytokines/chemokines, exosomes (179), or other molecules of interest. The various "-omes" present a high degree of interindividual, geographic, and time-dependent variation, and likely interact, affecting interpretation of results (180). This approach provides details on the type of metadata that could underlie our understanding of variability (173).

Microbes

Next-generation sequencing methods provide data on the relative 16S ribosomal RNA sequence composition but selection of primer pairs is important (4). Sequence data can be used to estimate species composition by comparison with known ribosomal RNA operon copy numbers of the nearest related species (181); limitations of the method have been noted (4, 182). Bacterial strain identification is not possible at present. Estimates of total bacterial load can be determined by qPCR using "universal" primers although primer bias and species-related DNA extraction efficiencies need to be considered (4, 183). Labor-intensive culture-based methods are useful for studying environmental sources of milk-culturable bacteria, but not to estimate total bacterial load or species distributions (184).

Environmental chemicals

Because environmental chemicals are ubiquitous, similar strategies for sample collection, handling, storage, and sample analysis are needed. Often for environmental chemicals, metabolites are the biomarker of exposure and not the parent compound. For example, di-2-ethylhexyl phthalate, the parent chemical in plastics, is metabolized in the body to mono-(2-ethylhexyl) phthalate and other monoester phthalates (185). Analytical methods used include HPLC (186) and MS and nontargeted analyses to identify novel environmental chemicals.

Traditionally, some chemicals, for example, dioxins and dioxin-like compounds, have factors associated with them to define their potential toxicity [toxicity equivalent factors (TEQs)]. TEQs are based on the known effects of a parent compound of a defined toxicity, namely the toxicity of the potent dioxin 2,3,7,8-tetrachlorodibenzodioxin. The TEQ can be summed across hundreds of congeners to give a total TEQ from HM exposure (27).

The above body of information underscores the immense variability reported in HMC and the importance of collecting metadata in conjunction with HM samples. Furthermore, improvement and harmonization of methods for sample collection, handling and storage, and chemical analysis is crucial, allowing compilation of data and comparisons across studies and populations over time. Several recent reviews have also emphasized the need for standardization of methodology for assessment of the variability factors (6, 102, 117).

Potential Next Steps for Human Milk Composition Data

Comprehensive and current data on HMC are lacking in the United States and Canada. Given the variability of

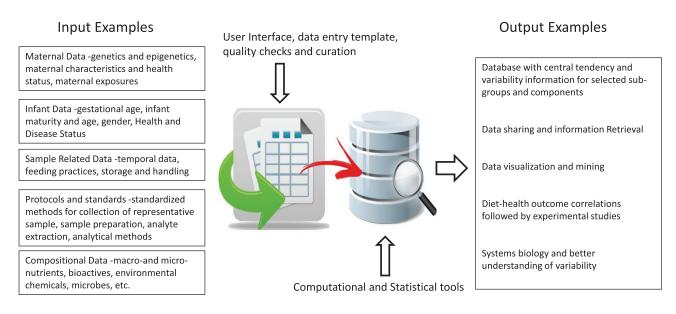


FIGURE 3 Schematic representation of the Human Milk Composition Data Repository (HMCD-R).

HMC, the use of out-of-date, single nutrient values for exposure assessments is inappropriate. FoodData Central and CANLINE in the United States and Canada, respectively, generally have the potential to incorporate metadata for food samples. However, given the uniqueness of HM and its variability factors and potential uses of the data, we articulate a vision for a publicly available HMCD-R. It can provide a central, integrated platform for national and international researchers to submit new or older HMC data that meet established protocols for sampling, collection, storage, analytical methods, and quality control, etc. HMCD-R would enable compiling, evaluating, comparing, tracking, and sharing HMC data, thus facilitating the understanding of the complexity and biological variability of HMC.

Figure 3 provides a schematic representation of a potential HMCD-R (as of now such a data repository does not exist). Some potential features and capabilities envisioned for HMCD-R are:

- Includes known and novel components (Figure 1).
- Contains well-characterized supporting metadata using standardized vocabulary.
- Uses a user-interface and a flexible data-entry template, allowing researchers from heterogeneous research studies to submit data.
- Includes submitted data that have undergone quality control, review, and curation to ensure data quality, integrity, and validity.
- Serves as a portal for standards/protocols for HMC analyses (sample preparation, handling, storage, and analytical methodologies), data sharing, reporting (including minimal metadata reporting standards) and statistical analyses.
- Provides computational, data mining/visualization, and statistical tools for compiling and integrating

data from different studies, comparative analyses, and determining central tendency, variability, and other statistical measures.

- Integrates FAIR (Findability, Accessibility, Interoperability, and Reusability) guiding principles for digital datasets (187).
- A structure that allows for incorporation or linkages to other data systems such as the Canadian Nutrient File (45), CANLINE (46), USDA FoodData Central (43), and NIH Metabolomics Workbench (188).
- Ensures data confidentiality by following Federal Health Insurance Privacy and Portability Act (HIPAA) privacy rules.

The vision and the considerations for the HMCD-R will be shaped by funding availability in the future and determined collaboratively by US and Canadian agencies.

Future Considerations

National health and nutrition monitoring is a cornerstone for many federal and state policies. Robust estimates of exposure assessments through HM are lacking to inform infant and toddler nutrition and health research, policy, and programmatic needs. The assumptions historically used to impute HM volume and the use of out-of-date, single average compositional data for all infants are outdated. Expansion is needed of sampling strategy in population surveys to recruit a larger sample of infants from mothers to capture diverse factors that influence HMC, collection of HM samples, and related metadata, additional focus on the components of public health interest, and statistical approaches to account for HMC variability to provide more robust consumption estimates from HM. More research is needed to understand the impact of variability factors on HMC, particularly from large studies that can adjust for multiple factors to identify independent associations (189– 191). HMC studies should report detailed characteristics that could contribute to variation between and within subjects and be adequately powered to estimate the central tendency and variability of nutrient concentrations.

Federal and nonprofit agencies have increased emphasis on HMC (9, 7, 192). The 2020 Dietary Guidelines Advisory Committee recommended updating HMC data in USDA databases (14). Several efforts, for example, BEGIN (Breastmilk Ecology: Genesis of Infant Nutrition) meeting series to describe in detail the variability factors by NIH (193), and the Mothers, Infants and Lactation Quality (MILQ) (194) and CHILD cohort studies (195) will likely provide crucial understanding to inform the development of HMCD-R. Projects such as MIREC and ECHO could be used as models to develop future North American surveillance and biomonitoring studies on HMC. HMC initiatives will continue to positively impact the pace of research by providing a collaboration platform whereby diverse disciplines related to HMC research in the United States/Canada can collaborate and synchronize their efforts to better understand HM through mechanisms to achieve the vision for HMCD-R.

The compiled compositional and metadata in HMCD-R would provide pertinent measures of central tendency and variability and allow use of statistical modeling techniques to approximate compositional profiles for subgroups, providing more accurate exposure assessments for purposes of monitoring and surveillance and development of recommendations to inform public health policies and food and nutrition programs. This work goes beyond nutrition to clarify potentially detrimental exposures that could occur through HM (e.g., chemicals, alcohol, drugs). Furthermore, it could improve understanding of the complexity and variability of HM (a key mediator of the diet-health-disease outcome correlations) and infant and maternal nutritional needs; allow researchers to study the impact of maternal and environmental factors and HM as a mother-milk-infant triad (192); and guide management of infant feeding, formulation of infant formulas, improved clinical practice guidelines, and personalized interventions/guidance to positively influence exposures and improve health outcomes for both the mother and child.

Acknowledgments

We acknowledge help in manuscript preparation and professional support of Mr Anderson-Villaluz and Ms Cadogan, Ms Ennis, Ms Gibbs, Ms Irrer, Ms McMillan, and Ms Parnel, and review of manuscript by Drs D'Onghia, Flannery, McCrea, Spungen, and Young-Hyman.

The authors' responsibilities were as follows—All authors contributed to writing and read and approved the final manuscript.

References

 World Health Organization. The optimal duration of exclusive breastfeeding: report of the expert consultation. [Internet]. 2001; [cited 2022 Aug 25]. Available from: https://apps.who.int/nutrition/publications/infantfeeding/WHO_NHD_01.09/en/index.html.

- Pérez-Escamilla R, Buccini GS, Segura-Pérez S, Piwoz E. Perspective: should exclusive breastfeeding still be recommended for 6 months? Adv Nutr 2019;10(6):931–43.
- 3. Eidelman AI, Schanler RJ. Breastfeeding and the use of human milk. Pediatrics 2012;129(3):e827-41.
- Abellan-Schneyder I, Matchado MS, Reitmeier S, Sommer A, Sewald Z, Baumbach J, et al. Primer, pipelines, parameters: issues in 16S rRNA gene sequencing. Msphere 2021;6(1):e01202–20.
- Vorosmarti A, Yaktine AL, Rasmussen K. Scanning for new evidence on the nutrient content of human milk: a process model for determining age-specific nutrient requirements. [Internet]. 2020; [cited 2022 Aug 25]. Available from: https: //nap.nationalacademies.org/catalog/25943/scanning-for-newevidence-on-the-nutrient-content-of-human-milk.
- Samuel TM, Zhou Q, Giuffrida F, Munblit D, Verhasselt V, Thakkar SK. Nutritional and non-nutritional composition of human milk is modulated by maternal, infant, and methodological factors. Front Nutr 2020;7:172.
- Christian P, Smith ER, Lee SE, Vargas AJ, Bremer AA, Raiten DJ. The need to study human milk as a biological system. Am J Clin Nutr 2021;113(5):1063–72.
- 8. Wu X, Jackson RT, Khan SA, Ahuja J, Pehrsson PR. Human milk nutrient composition in the United States: current knowledge, challenges, and research needs. Curr Dev Nutr 2018;2(7):nzy025.
- Casavale KO, Ahuja JKC, Wu X, Li Y, Quam J, Olson R, et al. NIH workshop on human milk composition: summary and visions. Am J Clin Nutr 2019;110(3):769–79.
- Zipf G, Chiappa M, Porter KS, Ostchega Y, Lewis BG, Dostal J. The National Health and Nutrition Examination Survey: plan and operations, 1999–2010. Vital Health Stat 2013;1(56):1–37.
- Hébert JR, Hurley TG, Steck SE, Miller DR, Tabung FK, Peterson KE, et al. Considering the value of dietary assessment data in informing nutrition-related health policy. Adv Nutr 2014;5(4): 447–55.
- 12. National Academies of Sciences, Engineering and Medicine. Dietary Reference Intakes for sodium and potassium. Washington (DC): National Academies Press; 2019.
- 13. Stoody EE, Casavale KO. Making the dietary guidelines for Americans "for Americans": the critical role of data analyses. J Food Compos Anal 2017;64:138–42.
- 14. Dietary Guidelines Advisory Committee. Scientific Report of the 2020 Dietary Guidelines Advisory Committee: Advisory Report to the Secretary of Agriculture and the Secretary of Health and Human Services. [Internet]. 2020; [cited 2021 Sep 7]. Available from: https://www.dietaryguidelines. gov/sites/default/files/2020-07/ScientificReport_of_the_ 2020DietaryGuidelinesAdvisoryCommittee_first-print.pdf.
- Health Canada. Canada's food guide. [Internet]. [cited 2022 Aug 25].
- Available from: https://food-guide.canada.ca/en/.
- 16. Food and Drug Administration. Food labeling: serving sizes of foods that can reasonably be consumed at one eating occasion; dual-column labeling; updating, modifying, and establishing certain reference amounts customarily consumed; serving size for breath mints; and technical amendments: guidance for industry small entity compliance guide. [Internet]. 2018; [cited 2022 Aug 25]. Available from: https: //www.fda.gov/media/111144/download
- Ahluwalia N. Nutrition monitoring of children aged birth to 24 mo (B-24): data collection and findings from the NHANES. Adv Nutr 2020;11(1):113–27.
- Ahluwalia N, Herrick K, Paulose-Ram R, Johnson C. Data needs for B-24 and beyond: NHANES data relevant for nutrition surveillance of infants and young children. Am J Clin Nutr 2014;99(3): 747S–54S.
- 19. National Center for Health Statistics. National Health Interview Survey. [Internet]; [cited 2022 Aug 25]. Available from: https://www.cdc.gov/nchs/nhis/index.htm.

- Statistics Canada. Questionnaires. [Internet]; [cited 2022 Aug 25]. Available from: https://www23.statcan.gc.ca/imdb-bmdi/pub/ indexti-eng.htm.
- 21. Health Canada. Reference guide to understanding and using the data. 2015 Canadian Community Health Survey—Nutrition. [Internet]. 2017; [cited 2022 Aug 25]. Available from: https://www.canada.ca/en/ health-canada/services/food-nutrition/food-nutrition-surveillance/ health-nutrition-survey/canadian-community-health-survey-cchs/ reference-guide-understanding-using-data-2015.html.
- 22. Statistics Canada. Canadian Community Health Survey (CCHS) cycle 2.2 (2004). Nutrition—general health (including vitamin & mineral supplements) & 24-hour dietary recall components user guide. [Internet]; [cited 2022 Aug 25]. Available from: http://www23.statcan.gc.ca/imdb-bmdi/document/5049_D24_T9_V1-eng.pdf.
- Tremblay MS, Gorber SC. Canadian Health Measures Survey. Can J Public Health 2007;98(6):453–6.
- Statistics Canada. Canadian Health Measures Survey (CHMS). [Internet]; [cited 2022 Jul 19]. Available from: https://www.statcan.gc. ca/en/survey/household/5071.
- 25. Lehmann GM, LaKind JS, Davis MH, Hines EP, Marchitti SA, Alcala C, et al. Environmental chemicals in breast milk and formula: exposure and risk assessment implications. Environ Health Perspect 2018;126(9):096001.
- Lorber M, Phillips L. Infant exposure to dioxin-like compounds in breast milk. Environ Health Perspect 2002;110(6):A325–A32.
- 27. van den Berg M, Kypke K, Kotz A, Tritscher A, Lee SY, Magulova K, e tal. WHO/UNEP global surveys of PCDDs, PCDFs, PCBs and DDTs in human milk and benefit–risk evaluation of breastfeeding. Arch Toxicol 2017;91(1):83–96.
- Food and Drug Administration. FDA Total Diet Study. [Internet]; [cited 2022 Aug 25]. Available from: https://www.fda.gov/food/ science-research-food/total-diet-study.
- 29. National Center for Health Statistics. NHANES, National Health and Examination Survey. About the National Health and Examination Survey. [Internet]; [cited 2022 Aug 25]. Available from: https://www.cdc.gov/nchs/nhanes/about_nhanes.htm.
- ECHO. ECHO program materials. [Internet]. Updated August 2022; [cited 2022 Aug 25]. Available from: https://dcricollab.dcri.duke.edu/ sites/echomaterials/SitePages/Home.aspx.
- Nestlé. The Feeding Infants and Toddlers Study. [Internet]; [cited 2022 Aug 25]. Available from: https://www.nestleusa.com/nutrition/fits.
- 32. Ahluwalia N, Herrick KA, Rossen LM, Rhodes D, Kit B, Moshfegh A, et al. Usual nutrient intakes of US infants and toddlers generally meet or exceed Dietary Reference Intakes: findings from NHANES 2009– 2012. Am J Clin Nutr 2016;104(4):1167–74.
- 33. Institute of Medicine. Dietary Reference Intakes Research Synthesis workshop summary. [Internet]. 2007; [cited 2022 Jul 19]. Available from: https://www.nap.edu/catalog/11767/dietary-reference-intakesresearch-synthesis-workshop-summary.
- 34. Dewey KG, Lönnerdal B. Milk and nutrient intake of breast-fed infants from 1 to 6 months: relation to growth and fatness. J Pediatr Gastroenterol Nutr 1983;2(3):497–506.
- Dewey KG, Finley DA, Lönnerdal B. Breast milk volume and composition during late lactation (7–20 months). J Pediatr Gastroenterol Nutr 1984;3(5):713–20.
- Kent JC, Mitoulas L, Cox DB, Owens RA, Hartmann PE. Breast volume and milk production during extended lactation in women. Exp Physiol 1999;84(2):435–47.
- Berube LT, Gross R, Messito MJ, Deierlein A, Katzow M, Woolf K. Concerns about current breast milk intake measurement for population-based studies. J Acad Nutr Diet 2018;118(10):1827–31.
- Heinig MJ, Nommsen LA, Peerson JM, Lonnerdal B, Dewey KG. Intake and growth of breast-fed and formula-fed infants in relation to the timing of introduction of complementary foods: the DARLING study. Acta Paediatr 1993;82(s385):999–1006.
- da Costa TH, Haisma H, Wells JC, Mander AP, Whitehead RG, Bluck LJ. How much human milk do infants consume? Data from

12 countries using a standardized stable isotope methodology. J Nutr 2010;140(12):2227–32.

- U.S. Department of Agriculture Agricultural Research Service. USDA food and nutrient database for dietary studies 2015–2016. [Internet]; [cited 2022 Jul 19]. Available from: http://www.ars.usda.gov/nea/ bhnrc/fsrg.
- 41. U.S. Department of Agriculture, US Department of Health and Human Services. Dietary guidelines for Americans, 2020–2025. 9th edition. [Internet]; [cited 2022 Jul 19]. Available from: https://www. dietaryguidelines.gov/.
- 42. Ahuja JK, Wasswa-Kintu S, Haytowitz DB, Daniel M, Thomas R, Showell B, et al. Sodium content of popular commercially processed and restaurant foods in the United States. Prev Med Rep 2015;2:962–7.
- 43. U.S. Department of Agriculture Agricultural Research Service. FoodData Central. [Internet]; [cited 2022 Jul 19]. Available from: https: //fdc.nal.usda.gov/.
- 44. Grimes C, Szymlek-Gay E, Nicklas T. Beverage consumption among US children aged 0–24 months: National Health and Nutrition Examination Survey (NHANES). Nutrients 2017;9(3):264.
- 45. Health Canada. The Canadian Nutrient File. [Internet]; [cited 2022 Jul 19]. Available from: https://www.canada.ca/en/health-canada/ services/food-nutrition/healthy-eating/nutrient-data/canadiannutrient-file-about-us.html.
- Health Canada. Canadian Laboratory Information Network. [Internet]; [cited 2022 Aug 25]. Available from: https://clin-rcil. hc-sc.gc.ca/clin-rcil/home.do.
- 47. LaKind JS, Lehmann GM, Davis MH, Hines EP, Marchitti SA, Alcala C, et al. Infant dietary exposures to environmental chemicals and infant/child health: a critical assessment of the literature. Environ Health Perspect 2018;126(9):096002.
- Wiciński M, Sawicka E, Gębalski J, Kubiak K, Malinowski B. Human milk oligosaccharides: health benefits, potential applications in infant formulas, and pharmacology. Nutrients 2020;12(1):266.
- Ruhaak LR, Lebrilla CB. Advances in analysis of human milk oligosaccharides. Adv Nutr 2012;3(3):406S–14S.
- 50. Field CJ. The immunological components of human milk and their effect on immune development in infants. J Nutr 2005;135(1):1-4.
- Liu B, Newburg DS. Human milk glycoproteins protect infants against human pathogens. Breastfeed Med 2013;8(4):354–62.
- 52. Meurant G. Handbook of milk composition. Elsevier; 1995.
- 53. Ballard O, Morrow AL. Human milk composition: nutrients and bioactive factors. Pediatr Clin North Am 2013;60(1):49–74.
- Schueler J, Alexander B, Hart AM, Austin K, Enette Larson-Meyer D. Presence and dynamics of leptin, GLP-1, and PYY in human breast milk at early postpartum. Obesity 2013;21(7):1451–8.
- 55. Das M, Padhy L, Koshy R, Sirsat SM, Rich MA. Human milk samples from different ethnic groups contain RNase that inhibits, and plasma membrane that stimulates, reverse transcription. Nature 1976;262(5571):802–5.
- Dalaly B, Eitenmiller R, Friend B, Shahani K. Human milk ribonuclease. Biochim Biophys Acta Enzymol 1980;615(2):381– 91.
- 57. van Herwijnen MJ, Zonneveld MI, Goerdayal S, Nolte EN, Garssen J, Stahl B, et al. Comprehensive proteomic analysis of human milk-derived extracellular vesicles unveils a novel functional proteome distinct from other milk components. Mol Cell Proteomics 2016;15(11):3412–23.
- Dayon L, Macron C, Lahrichi S, Nu'nez Galindo A, Affolter M. Proteomics of human milk: definition of a discovery workflow for clinical research studies. J Proteome Res 2021;20(5): 2283–90.
- Ruiz L, Fernández L, Rodríguez JM. Immune factors in human milk. In: McGuire MK, O'Connor DL, editors. Human milk. Elsevier; 2021. p. 275–98.
- 60. Perrella S, Gridneva Z, Lai CT, Stinson L, George A, Bilston-John S, et al. Human milk composition promotes optimal infant growth, development and health. Semin Perinatol 2021;45(2):151380.

- 61. Sánchez CL, Cubero J, Sánchez J, Chanclón B, Rivero M, Rodríguez AB, et al. The possible role of human milk nucleotides as sleep inducers. Nutr Neurosci 2009;12(1):2–8.
- Andreas NJ, Kampmann B, Le-Doare KM. Human breast milk: a review on its composition and bioactivity. Early Hum Dev 2015;91(11):629–35.
- Demmelmair H, Ahmed TB, Koletzko B. Content, variability, and regulation of fatty acids in human milk. In: McGuire MK, O'Connor DL, editors. Human milk. Elsevier; 2021. p. 103–43.
- 64. Demmelmair H, Koletzko B. Lipids in human milk. Best Pract Clin Endocrinol Metab 2018;32(1):57–68.
- Keenan TW, Patton S. The structure of milk: implications for sampling and storage: a. The milk lipid globule membrane. In: Jensen RG, editor. Handbook of milk composition. San Diego: Academic Press; 1995. p. 5–50.
- 66. Fox FW, Gardner JA. The cholesterol content of human milk. Biochem J 1924;18(1):127.
- Wolford S, Argoudelis C. Measurement of estrogens in cow's milk, human milk, and dairy products. J Dairy Sci 1979;62(9):1458–63.
- 68. Woollett L, Heubi JE. Fetal and neonatal cholesterol metabolism: MDText.com, Inc. South Dartmouth (MA), 2002.
- Pundir S, Wall CR, Mitchell CJ, Thorstensen EB, Lai CT, Geddes DT, et al. Variation of human milk glucocorticoids over 24 hour period. J Mammary Gland Biol Neoplasia 2017;22(1):85–92.
- Kallio M, Siimes MA, Perheentupa J, Salmenperä L, Miettinen TA. Cholesterol and its precursors in human milk during prolonged exclusive breast-feeding. Am J Clin Nutr 1989;50(4):782–5.
- Picciano MF, Guthrie HA, Sheehe DM. The cholesterol content of human milk: a variable constituent among women and within the same woman. Clin Pediatr (Phila) 1978;17(4):359–62.
- 72. Laiho K, Lampi A-M, Hämäläinen M, Moilanen E, Piironen V, Arvola T, et al. Breast milk fatty acids, eicosanoids, and cytokines in mothers with and without allergic disease. Pediatr Res 2003;53(4):642–7.
- 73. Weiss GA, Troxler H, Klinke G, Rogler D, Braegger C, Hersberger M. High levels of anti-inflammatory and pro-resolving lipid mediators lipoxins and resolvins and declining docosahexaenoic acid levels in human milk during the first month of lactation. Lipids Health Dis 2013;12(1):1–12.
- Arnardottir H, Orr SK, Dalli J, Serhan CN. Human milk proresolving mediators stimulate resolution of acute inflammation. Mucosal Immunol 2016;9(3):757–66.
- 75. Dahlqvist A. Method for assay of intestinal disaccharaidases. Anal Biochem 1964;7(1):18-25.
- Dror DK, Allen LH. Iodine in human milk: a systematic review. Adv Nutr 2018;9(Suppl 1):347S–57S.
- Dror DK, Allen LH. Overview of nutrients in human milk. Adv Nutr 2018;9(Suppl 1):278S–94S.
- Lönnerdal B. Bioactive proteins in breast milk. J Paediatr Child Health 2013;49:1–7.
- Bode L, McGuire M, Rodriguez JM, Geddes DT, Hassiotou F, Hartmann PE, et al. It's alive: microbes and cells in human milk and their potential benefits to mother and infant. Oxford University Press; 2014.
- Martín R, Langa S, Reviriego C, Jimínez E, Marín ML, Xaus J, et al. Human milk is a source of lactic acid bacteria for the infant gut. J Pediatr 2003;143(6):754–8.
- Consales A, Cerasani J, Sorrentino G, Morniroli D, Colombo L, Mosca F, et al. The hidden universe of human milk microbiome: origin, composition, determinants, role, and future perspectives. Eur J Pediatr 2022;181:1811–20.
- Shenhav L, Azad MB. Using community ecology theory and computational microbiome methods to study human milk as a biological system. Msystems 2022;7(1):e01132–21.
- 83. Lackey KA, Williams JE, Meehan CL, Zachek JA, Benda ED, Price WJ, et al. What's normal? Microbiomes in human milk and infant feces are related to each other but vary geographically: the INSPIRE study. Front Nutr 2019;6:45.

- 84. Lyons KE, Shea C-AO, Grimaud G, Ryan CA, Dempsey E, Kelly AL, et al. The human milk microbiome aligns with lactation stage and not birth mode. Sci Rep 2022;12(1):5598.
- Stinson LF, Trevenen ML, Geddes DT. Effect of cold storage on the viable and total bacterial populations in human milk. Nutrients 2022;14(9):1875.
- Stiehm ER, Keller MA. Breast milk transmission of viral disease. Adv Nutr Res 2001;10:105–22.
- Oxtoby MJ. Human immunodeficiency virus and other viruses in human milk: placing the issues in broader perspective. Pediatr Infect Dis J 1988;7(12):825–35.
- Kreiss J. Breastfeeding and vertical transmission of HIV-1. Acta Paediatr 1997;86(S421):113–7.
- Jim W-T, Chiu N-C, Ho C-S, Shu C-H, Chang J-H, Hung H-Y, et al. Outcome of preterm infants with postnatal cytomegalovirus infection via breast milk: a two-year prospective follow-up study. Medicine (Baltimore) 2015;94(43):e1835.
- 90. Selma-Royo M, Calvo-Lerma J, Bäuerl C, Esteban-Torres M, Cabrera-Rubio R, Collado MC. Human milk microbiota: what did we learn in the last 20 years? Microbiome Res Rep 2022;1(3):19.
- 91. Marchitti SA, Fenton SE, Mendola P, Kenneke JF, Hines EP. Polybrominated diphenyl ethers in human milk and serum from the US EPA MAMA study: modeled predictions of infant exposure and considerations for risk assessment. Environ Health Perspect 2017;125(4):706–13.
- 92. VanNoy BN, Lam J, Zota AR. Breastfeeding as a predictor of serum concentrations of per-and polyfluorinated alkyl substances in reproductive-aged women and young children: a rapid systematic review. Curr Environ Health Rep 2018;5(2):213–24.
- 93. Xie L, Innis SM. Genetic variants of the FADS1 FADS2 gene cluster are associated with altered (n-6) and (n-3) essential fatty acids in plasma and erythrocyte phospholipids in women during pregnancy and in breast milk during lactation. J Nutr 2008;138(11): 2222-8.
- 94. Morales E, Bustamante M, Gonzalez JR, Guxens M, Torrent M, Mendez M, et al. Genetic variants of the FADS gene cluster and ELOVL gene family, colostrums LC-PUFA levels, breastfeeding, and child cognition. PLoS One 2011;6(2):e17181.
- 95. Lattka E, Rzehak P, Szabo E, Jakobik V, Weck M, Weyermann M, et al. Genetic variants in the FADS gene cluster are associated with arachidonic acid concentrations of human breast milk at 1.5 and 6 mo postpartum and influence the course of milk dodecanoic, tetracosenoic, and trans-9-octadecenoic acid concentrations over the duration of lactation. Am J Clin Nutr 2011;93(2):382–91.
- Bezerra FF, Cabello GM, Mendonca LM, Donangelo CM. Bone mass and breast milk calcium concentration are associated with vitamin D receptor gene polymorphisms in adolescent mothers. J Nutr 2008;138(2):277–81.
- Alam S, Hennigar SR, Gallagher C, Soybel DI, Kelleher SL. Exome sequencing of SLC30A2 identifies novel loss-and gain-of-function variants associated with breast cell dysfunction. J Mammary Gland Biol Neoplasia 2015;20(3-4):159–72.
- Page R, Wong A, Arbuckle TE, MacFarlane AJ. The MTHFR 677C>T polymorphism is associated with unmetabolized folic acid in breast milk in a cohort of Canadian women. Am J Clin Nutr 2019;110(2):401– 9.
- 99. Daniel AI, Shama S, Ismail S, Bourdon C, Kiss A, Mwangome M,et al. Maternal BMI is positively associated with human milk fat: a systematic review and meta-regression analysis. Am J Clin Nutr 2021;113(4):1009–22.
- Pereira KB, de Azeredo VB, da Sileira CB, Pedruzzi LM. Composition of breast milk of lactating adolescents in function of time of lactation. Nutr Hosp 2013;28(6):1971–6.
- 101. Campanhon IB, da Silva MRS, de Magalhães MTQ, Zingali RB, Bezerra FF, Torres AG. Protective factors in mature human milk: a look into the proteome and peptidome of adolescent mothers' breast milk. Br J Nutr 2019;122(12):1377–85.

- 102. Verduci E, Gianni ML, Vizzari G, Vizzuso S, Cerasani J, Mosca F, et al. The triad mother-breast milk-infant as predictor of future health: a narrative review. Nutrients 2021;13(2):486.
- 103. do Amaral YNdV, Rocha DM, da Silva LML, Soares FVM, Moreira MEL. Do maternal morbidities change the nutritional composition of human milk? A systematic review. Cien Saude Colet 2019;24: 2491–8.
- 104. Allen LH. B vitamins in breast milk: relative importance of maternal status and intake, and effects on infant status and function. Adv Nutr 2012;3(3):362–9.
- 105. Bravi F, Wiens F, Decarli A, Dal Pont A, Agostoni C, Ferraroni M. Impact of maternal nutrition on breast-milk composition: a systematic review. Am J Clin Nutr 2016;104(3):646–62.
- 106. Adhikari S, Kudla U, Nyakayiru J, Brouwer-Brolsma EM. Maternal dietary intake, nutritional status, and macronutrient composition of human breast milk: systematic review. [Internet]. Br J Nutr 2021. doi:10.1017/S0007114521002786.
- 107. Gidrewicz DA, Fenton TR. A systematic review and meta-analysis of the nutrient content of preterm and term breast milk. BMC Pediatr 2014;14(1):1–14.
- 108. Gidrewicz DA, Fenton TR. A systematic review and meta-analysis of the nutrient content of preterm and term breast milk. BMC Pediatr 2014;14(1):216.
- 109. Fischer Fumeaux CJ, Garcia-Rodenas CL, De Castro CA, Courtet-Compondu M-C, Thakkar SK, Beauport L, et al. Longitudinal analysis of macronutrient composition in preterm and term human milk: a prospective cohort study. Nutrients 2019;11(7):1525.
- 110. Galante L, Milan AM, Reynolds CM, Cameron-Smith D, Vickers MH, Pundir S. Sex-specific human milk composition: the role of infant sex in determining early life nutrition. Nutrients 2018;10(9):1194.
- 111. Phattraprayoon N, Kraisonsin N, Kanjanapattanakul W. Comparison of breast milk compositions among mothers delivering smallfor-gestational age, appropriate-for-gestational age, and large-forgestational age infants. Breastfeed Med 2018;13(9):627–30.
- 112. Gardner AS, Rahman IA, Lai CT, Hepworth A, Trengove N, Hartmann PE, et al. Changes in fatty acid composition of human milk in response to cold-like symptoms in the lactating mother and infant. Nutrients 2017;9(9):1034.
- Karra MV, Udipi S, Kirksey A, Roepke J. Changes in specific nutrients in breast milk during extended lactation. Am J Clin Nutr 1986;43(4):495–503.
- 114. Czosnykowska-Łukacka M, Królak-Olejnik B, Orczyk-Pawiłowicz M. Breast milk macronutrient components in prolonged lactation. Nutrients 2018;10(12):1893.
- 115. Italianer MF, Naninck EF, Roelants JA, van der Horst GT, Reiss IK, Goudoever JBv, et al. Circadian variation in human milk composition, a systematic review. Nutrients 2020;12(8):2328.
- 116. Hampel D, Shahab-Ferdows S, Islam MM, Peerson JM, Allen LH. Vitamin concentrations in human milk vary with time within feed, circadian rhythm, and single-dose supplementation. J Nutr 2017;147(4):603–11.
- 117. Leghi GE, Middleton PF, Netting MJ, Wlodek ME, Geddes DT, Muhlhausler BS. A systematic review of collection and analysis of human milk for macronutrient composition. J Nutr 2020;150(6):1652– 70.
- 118. við Streym S, Højskov C, Møller U, Heickendorff L, Vestergaard P, Mosekilde L, et al. Vitamin D content in human breast milk: a 9-mo follow-up study1. Am J Clin Nutr 2016;103(1):107–14.
- 119. Fehr K, Moossavi S, Sbihi H, Boutin RC, Bode L, Robertson B, et al. Breastmilk feeding practices are associated with the co-occurrence of bacteria in mothers' milk and the infant gut: the CHILD cohort study. Cell Host Microbe 2020;28(2):285–97.e4.
- 120. Mense L, Rößler S, Hanusch R, Roßberg C, Rüdiger M. Bacterial contamination of mechanically extracted breast milk. Am J Perinatol 2014;31(04):293–8.
- 121. Flaherman VJ, Lee HC. "Breastfeeding" by feeding expressed mother's milk. Pediatr Clin North Am 2013;60(1):227–46.

- 122. Pittard WB, Geddes KM, Brown S, Mintz S, Hulsey TC. Bacterial contamination of human milk: container type and method of expression. Am J Perinatol 1991;8(01):25–7.
- 123. Ahrabi AF, Handa D, Codipilly CN, Shah S, Williams JE, McGuire MA, et al. Effects of extended freezer storage on the integrity of human milk. J Pediatr 2016;177:140–3.
- 124. Pandya SP, Doshi H, Codipilly CN, Fireizen Y, Potak D, Schanler RJ. Bacterial stability with freezer storage of human milk. J Perinat Med 2021;49(2):225–8.
- 125. Păduraru L, Zonda GI, Avasiloaiei A-L, Moscalu M, Dimitriu DC, Stamatin M. Influence of refrigeration or freezing on human milk macronutrients and energy content in early lactation: results from a tertiary centre survey. Paediatr Child Health 2019;24(4):250–7.
- 126. Lönnerdal B. Bioactive proteins in human milk: health, nutrition, and implications for infant formulas. J Pediatr 2016;173:S4–S9.
- 127. Ewaschuk JB, Unger S, Harvey S, O'Connor DL, Field CJ. Effect of pasteurization on immune components of milk: implications for feeding preterm infants. Appl Physiol Nutr Metab 2011;36(2):175–82.
- 128. Padilha M, Danneskiold-Samsøe NB, Brejnrod A, Hoffmann C, Cabral VP, de Melo Iaucci J, et al. The human milk microbiota is modulated by maternal diet. Microorganisms 2019;7(11):502.
- 129. Allmyr M, Adolfsson-Erici M, McLachlan MS, Sandborgh-Englund G. Triclosan in plasma and milk from Swedish nursing mothers and their exposure via personal care products. Sci Total Environ 2006;372(1):87–93.
- 130. Allmyr M, McLachlan MS, Sandborgh-Englund G, Adolfsson-Erici M. Determination of triclosan as its pentafluorobenzoyl ester in human plasma and milk using electron capture negative ionization mass spectrometry. Anal Chem 2006;78(18):6542–6.
- 131. Macchi M, Bambini L, Franceschini S, ID Alexa, Agostoni C. The effect of tobacco smoking during pregnancy and breastfeeding on human milk composition—a systematic review. Eur J Clin Nutr 2021;75(5):736–47.
- 132. US Environmental Protectin Agency. Technical fact sheet perfluorooctane sulfonate (PFOS) and perfluorooctanoic acid (PFOA). [Internet]. November 2017; [cited 2022 Jul 19]. Available from: https://19january2021snapshot.epa.gov/sites/static/files/2017-12/documents/ffrrofactsheet_contaminants_pfos_pfoa_11-20-17_508_0.pdf.
- 133. Nyberg E, Awad R, Bignert A, Ek C, Sallsten G, Benskin JP. Interindividual, inter-city, and temporal trends of per-and polyfluoroalkyl substances in human milk from Swedish mothers between 1972 and 2016. Environ Sci Process Impacts 2018;20(8):1136–47.
- 134. Sundström M, Ehresman DJ, Bignert A, Butenhoff JL, Olsen GW, Chang S-C, et al. A temporal trend study (1972–2008) of perfluorooctanesulfonate, perfluorohexanesulfonate, and perfluorooctanoate in pooled human milk samples from Stockholm, Sweden. Environ Int 2011;37(1):178–83.
- 135. Calafat AM, Wong L-Y, Kuklenyik Z, Reidy JA, Needham LL. Polyfluoroalkyl chemicals in the US population: data from the National Health and Nutrition Examination Survey (NHANES) 2003–2004 and comparisons with NHANES 1999–2000. Environ Health Perspect 2007;115(11):1596–602.
- 136. Hernandez-Avila M, Gonzalez-Cossio T, Hernandez-Avila JE, Romieu I, Peterson KE, Aro A, et al. Dietary calcium supplements to lower blood lead levels in lactating women: a randomized placebo-controlled trial. Epidemiology 2003;14(2):206–12.
- 137. Ettinger AS, Téllez-Rojo MM, Amarasiriwardena C, Peterson KE, Schwartz J, Aro A, et al. Influence of maternal bone lead burden and calcium intake on levels of lead in breast milk over the course of lactation. Am J Epidemiol 2006;163(1):48–56.
- 138. Gulson B. Lead in breast milk. Fact sheet for medical professionals. Lead Action News 2004;6:2.
- 139. Johnson MA. High calcium intake blunts pregnancy-induced increases in maternal blood lead. Nutr Rev 2001;59(5):152.
- 140. Ettinger A, Amarasiriwardena C, Smith D, Mercado-García A, Lamadrid-Figueroa H, Téllez-Rojo M. Milk-to-plasma ratios

indicate that lead concentrates in human breast milk. Epidemiology 2007;18(5):S61.

- 141. Arora M, Ettinger AS, Peterson KE, Schwartz J, Hu H, Hernández-Avila M, et al. Maternal dietary intake of polyunsaturated fatty acids modifies the relationship between lead levels in bone and breast milk. J Nutr 2008;138(1):73–9.
- Oskarsson A, Hallén IP, Sundberg J. Exposure to toxic elements via breast milk. Analyst 1995;120(3):765–70.
- 143. Oskarsson A, Schütz A, Skerfving S, Hallén IP, Ohlin B, Lagerkvist BJ. Total and inorganic mercury in breast milk and blood in relation to fish consumption and amalgam fillings in lactating women. Arch Environ Health 1996;51(3):234–41.
- 144. Drasch G, Aigner S, Roider G, Staiger E, Lipowsky G. Mercury in human colostrum and early breast milk. Its dependence on dental amalgam and other factors. J Trace Elem Med Biol 1998;12(1):23–7.
- 145. McGuire M, O'Connor LD. Human milk: sampling and measurement of energy-yielding nutrients and other macromolecules. Academic Press; 2020.
- 146. Hampel D, Dror DK, Allen LH. Micronutrients in human milk: analytical methods. Adv Nutr 2018;9(Suppl 1):313S-31S.
- 147. Bauer J, Gerss J. Longitudinal analysis of macronutrients and minerals in human milk produced by mothers of preterm infants. Clin Nutr 2011;30(2):215–20.
- 148. Nommsen LA, Lovelady CA, Heinig MJ, Lönnerdal B, Dewey KG. Determinants of energy, protein, lipid, and lactose concentrations in human milk during the first 12 mo of lactation: the DARLING study. Am J Clin Nutr 1991;53(2):457–65.
- 149. Leghi GE, Middleton PF, Muhlhausler BS. A methodological approach to identify the most reliable human milk collection method for compositional analysis: a systematic review protocol. Syst Rev 2018;7(1):122.
- 150. Page R, Robichaud A, Arbuckle TE, Fraser WD, MacFarlane AJ. Total folate and unmetabolized folic acid in the breast milk of a cross-section of Canadian women. Am J Clin Nutr 2017;105(5):1101–9.
- 151. Chang J-C, Chen C-H, Fang L-J, Tsai C-R, Chang Y-C, Wang T-M. Influence of prolonged storage process, pasteurization, and heat treatment on biologically-active human milk proteins. Pediatr Neonatol 2013;54(6):360–6.
- 152. Lawrence RA. Milk banking: the influence of storage procedures and subsequent processing on immunologic components of human milk. Adv Nutr Res 2001;10:389–404.
- Friend LL, Perrin MT. Fat and protein variability in donor human milk and associations with milk banking processes. Breastfeed Med 2020;15(6):370–6.
- 154. Goldblum RM, Garza C, Johnson CA, Harrist R, Nichols BL, Goldman AS. Human milk banking I. Effects of container upon immunologic factors in mature milk. Nutr Res 1981;1(5):449–59.
- 155. Ewaschuk JB, Unger S. Human milk pasteurization: benefits and risks. Curr Opin Clin Nutr Metab Care 2015;18(3):269–75.
- 156. Castro M, Asbury M, Shama S, Stone D, Yoon EW, O'Connor DL, et al. Energy and fat intake for preterm infants fed donor milk is significantly impacted by enteral feeding method. JPEN J Parenter Enteral Nutr 2019;43(1):162–5.
- 157. Gao C, Miller J, Middleton PF, Huang Y-C, McPhee AJ, Gibson RA. Changes to breast milk fatty acid composition during storage, handling and processing: a systematic review. Prostaglandins Leukot Essent Fatty Acids 2019;146:1–10.
- García-Lara NR, Escuder-Vieco D, García-Algar O, De la Cruz J, Lora D, Pallás-Alonso C. Effect of freezing time on macronutrients and energy content of breastmilk. Breastfeed Med 2012;7(4):295–301.
- 159. Hampel D, Allen LH. Analyzing B-vitamins in human milk: methodological approaches. Crit Rev Food Sci Nutr 2016;56(3):494– 511.
- 160. Francis J, Dickton D. Effects of light on riboflavin and ascorbic acid in freshly expressed human milk. J Nutr Health Food Eng 2015;2(6): 221–3.

- 161. Abramovich M, Friel J, Hossain Z. Polyunsaturated fatty acids, riboflavin and vitamin C: effect of different storage conditions of human milk. Vitam Miner 2013;2:110.
- 162. Miller EM, Aiello MO, Fujita M, Hinde K, Milligan L, Quinn E. Field and laboratory methods in human milk research. Am J Hum Biol 2013;25(1):1–11.
- 163. Sousa SG, Delgadillo I, Saraiva JA. Human milk composition and preservation: evaluation of high-pressure processing as a nonthermal pasteurization technology. Crit Rev Food Sci Nutr 2016;56(6):1043– 60.
- 164. Romeu-Nadal M, Morera-Pons S, Castellote A, Lopez-Sabater M. Rapid high-performance liquid chromatographic method for vitamin C determination in human milk versus an enzymatic method. J Chromatogr B 2006;830(1):41–6.
- 165. Garwolin 'ska D, Młynarczyk M, Kot-Wasik A, Hewelt-Belka W. The influence of storage on human milk lipidome stability for lipidomic studies. J Proteome Res 2022;21(2):438–46.
- 166. Hamosh M, Ellis LA, Pollock DR, Henderson TR, Hamosh P. Breastfeeding and the working mother: effect of time and temperature of short-term storage on proteolysis, lipolysis, and bacterial growth in milk. Pediatrics 1996;97(4):492–8.
- Picciano MF. Human milk: nutritional aspects of a dynamic food. Neonatology 1998;74(2):84–93.
- Witkowska-Zimny M, Kaminska-El-Hassan E. Cells of human breast milk. Cell Mol Biol Lett 2017;22(1):1–11.
- 169. Holden JM, Bhagwat SA, Patterson KY. Development of a multinutrient data quality evaluation system. J Food Compos Anal 2002;15(4):339–48.
- 170. Thurl S, Munzert M, Boehm G, Matthews C, Stahl B. Systematic review of the concentrations of oligosaccharides in human milk. Nutr Rev 2017;75(11):920–33.
- 171. van Leeuwen SS. Challenges and pitfalls in human milk oligosaccharide analysis. Nutrients 2019;11(11):2684.
- 172. Bardanzellu F, Fanos V, Reali A. "Omics" in human colostrum and mature milk: looking to old data with new eyes. Nutrients 2017;9(8):843.
- 173. Ten-Doménech I, Ramos-Garcia V, Piñeiro-Ramos JD, Gormaz M, Parra-Llorca A, Vento M, et al. Current practice in untargeted human milk metabolomics. Metabolites 2020;10(2):43.
- 174. Liao Y, Alvarado R, Phinney B, Lo"nnerdal B. Proteomic characterization of specific minor proteins in the human milk casein fraction. J Proteome Res 2011;10(12):5409–15.
- 175. Liao Y, Weber D, Xu W, Durbin-Johnson BP, Phinney BS, Lo"nnerdal B. Absolute quantification of human milk caseins and the whey/casein ratio during the first year of lactation. J Proteome Res 2017;16(11):4113–21.
- 176. Wu LD, Ruhaak LR, Lebrilla CB. Analysis of milk oligosaccharides by mass spectrometry. In: Lauc G, Wuhrer M, editors. High-throughput glycomics and glycoproteomics: methods and protocols. New York (NY): Springer New York; 2017. p. 121–9.
- 177. Garwolin 'ska D, Hewelt-Belka W, Namies 'nik J, Kot-Wasik A. Rapid characterization of the human breast milk lipidome using a solid-phase microextraction and liquid chromatography-mass spectrometrybased approach. J Proteome Res 2017;16(9):3200–8.
- Ruiz L, García-Carral C, Rodriguez JM. Unfolding the human milk microbiome landscape in the omics era. Front Microbiol 2019;10:1378.
- 179. de la Torre Gomez C, Goreham RV, Bech Serra JJ, Nann T, Kussmann M. "Exosomics"—a review of biophysics, biology and biochemistry of exosomes with a focus on human breast milk. Front Genet 2018;9:92.
- 180. Gómez-Gallego C, Morales JM, Monleón D, Du Toit E, Kumar H, Linderborg KM, et al. Human breast milk NMR metabolomic profile across specific geographical locations and its association with the milk microbiota. Nutrients 2018;10(10):1355.
- 181. Klappenbach JA, Saxman PR, Cole JR, Schmidt TM. rrndb: the ribosomal RNA operon copy number database. Nucleic Acids Res 2001;29(1):181–4.

- Louca S, Doebeli M, Parfrey LW. Correcting for 16S rRNA gene copy numbers in microbiome surveys remains an unsolved problem. Microbiome 2018;6(1):1–12.
- 183. Brooks JP, Edwards DJ, Harwich MD, Rivera MC, Fettweis JM, Serrano MG, et al. The truth about metagenomics: quantifying and counteracting bias in 16S rRNA studies. BMC Microbiol 2015;15(1):1– 14.
- 184. Nayfach S, Shi ZJ, Seshadri R, Pollard KS, Kyrpides NC. New insights from uncultivated genomes of the global human gut microbiome. Nature 2019;568(7753):505–10.
- 185. Agency for Toxic Substances and Disease Registry (ATSDR). Public health statement, di(2-ethylhexyl)phthalate (DEHP), CAS#: 117-81-7. [Internet]. September 2002; [cited 2022 Aug 25]. Available from: https: //www.atsdr.cdc.gov/ToxProfiles/tp9-c1-b.pdf.
- 186. CDC. Laboratory procedure manual. PCDDs, PCDFs, cPCBs and ortho-substituted PCBs. [Internet]. 2006; [cited 2022 Aug 25]. Available from: https://wwwn.cdc.gov/nchs/data/nhanes/2003-2004/ labmethods/l28_c_met_dioxins.pdf.
- 187. Fairsharing.org. A curated, informative and educational resource on data and metadata standards, inter-related to databases and data policies. [Internet]; [cited 2022 Jul 19]. Available from: https:// fairsharing.org/.
- 188. Metabolomics Workbench. National Metabolomics Data Repository. [Internet]; [cited 2022 Jul 19]. Available from: https://www.metabolomicsworkbench.org.
- 189. Miliku K, Duan QL, Moraes TJ, Becker AB, Mandhane PJ, Turvey SE, et al. Human milk fatty acid composition is associated with dietary, genetic, sociodemographic, and environmental factors in the CHILD cohort study. Am J Clin Nutr 2019;110(6):1370–83.
- 190. Azad MB, Robertson B, Atakora F, Becker AB, Subbarao P, Moraes TJ, et al. Human milk oligosaccharide concentrations are

associated with multiple fixed and modifiable maternal characteristics, environmental factors, and feeding practices. J Nutr 2018;148(11): 1733–42.

- 191. Hopperton KE, Pitino MA, Chouinard-Watkins R, Shama S, Sammut N, Bando N, et al. Determinants of fatty acid content and composition of human milk fed to infants born weighing <1250 g. Am J Clin Nutr 2021;114(4):1523–34.</p>
- 192. Bode L, Raman AS, Murch SH, Rollins NC, Gordon JI. Understanding the mother-breastmilk-infant "triad." Science 2020;367(6482): 1070–2.
- 193. NIH Eunice Kennedy Shriver National Institute of Child Health and Human Development. Breastmilk Ecology: Genesis of Infant Nutrition (BEGIN) series. [Internet]; [cited 2022 Aug 25]. Available from: https: //www.nichd.nih.gov/about/meetings/2021/011521.
- 194. Allen LH, Hampel D, Shahab-Ferdows S, Andersson M, Barros E, Doel AM, et al. The Mothers, Infants, and Lactation Quality (MILQ) study: a multi-center collaboration. Curr Dev Nutr 2021;5(10): nzab116.
- 195. CHILD Cohort Study. CHILD Cohort Study. 3400+ Canadian kids are helping to predict, prevent and treat chronic diseases. [Internet]; [cited 2022 Aug 25]. Available from: https://childstudy.ca/.
- 196. Statistics Canada. Canadian Community Health Survey (CCHS) Annual Component. User guide. 2015 Microdata files. Available on request from: statcan.hd-ds.statcan@canada.ca.
- 197. Gillman M, Blaisdell C. Environmental influences on child health outcomes, a research program of the National Institutes of Health. Curr Opin Pediatr 2018;30(2):260–2.
- 198. Arbuckle TE, Fraser WD, Fisher M, Davis K, Liang CL, Lupien N, et al. Cohort profile: the maternal-infant research on environmental chemicals research platform. Paediatr Perinat Epidemiol 2013;27(4):415–25.