INTERNATIONAL JOURNAL OF MEDICAL BIOCHEMISTRY

DOI: 10.14744/ijmb.2024.30075 Int J Med Biochem 2025;8(1):45-49





Investigation of a number of rare deletional mutations in the alpha globin gene cluster

💿 Majid Arash¹, 💿 Mehdi Gholami Bahnemiri²

¹Faculty of Medicine, Kermanshah University of Medical Sciences, Kermanshah, Iran ²Department of Laboratory Sciences, Sari Branch, Islamic Azad University, Sari, Iran

Abstract

Alpha thalassemia is one of the most common genetic diseases in the world. This disease is prevalent in various parts of the world, such as India, the Middle East, Africa, and many other countries. Several clinical conditions can result from mutations. In the condition where only one of the alpha globin genes is expressed, hemoglobin H disease (Hb H) occurs. Alpha thalassemia trait and silent carrier are milder forms of the disease, caused by the deletion of one and two alpha globin genes, respectively. Several mutations result in the deletion of alpha-globin. Seven common deletional mutations include $-\alpha^{4.2}, -\alpha^{3.7}, -(\alpha)^{20.5}, --^{MED}, --^{SEA}, --^{Fil}$, and $-^{THAI}$. The deletional mutations $-\alpha^{4.2}$ and $-\alpha^{3.7}$ remove only one of the alpha globin genes, while others remove both α^{1} and α^{2} globin genes from the gene cluster. Nowadays, laboratories identify these mutations using the Gap PCR method and other advanced methods. In addition to these mutations, some deletional mutations are found only in certain families or certain regions.

Keywords: Alpha thalassemia, deletional mutation, globin, hydrops fetalis

How to cite this article: Arash M, Bahnemiri MG. Investigation of a number of rare deletional mutations in the alpha globin gene cluster. Int J Med Biochem 2025;8(1):45–49.

Ipha thalassemia is a genetic disease with autosomal recessive inheritance, characterized by a reduction or lack of production of alpha globin chains. These alpha globin chains are components of fetal and adult hemoglobin. Unlike beta thalassemia, which is caused by non-deletion mutations, most known types of alpha thalassemia involve the deletion of one or two alpha globin genes on chromosome 13.3p16 [1, 2]. Both non-deletional and deletional mutations cause alpha thalassemia. Many non-deletional mutations of the alpha globin gene result from nucleotide substitutions. The most common deletional mutations are monogenic deletions of $-\alpha^{3.7}$ and $-\alpha^{4.2}$ in Asia and bigenic deletions $--^{SEA}$ and $--^{FIL}$ in Southeast Asia, $-(\alpha)^{20.5}$ and $--^{MED}$ in Mediterranean regions, and --THAI in Taiwan [3, 4]. Several other deletional mutations have been identified in recent years. In this article, we review eight of these deletional mutations. These deletions have a high prevalence and are seen in different proportions in different parts of the world. However, some deletional mutations are seen only in cases or in much smaller numbers.

Alpha Thalassemia

The alpha globin cluster at its locus includes duplicated alpha genes, an embryonic α -like gene (ζ 2), three pseudogenes ($\Psi\zeta$ 1, $\Psi\alpha$ 2, $\Psi\alpha$ 1), and a gene with an undetermined function (θ 1). These genes and pseudogenes are arranged in the order 1 ζ 2- $\Psi\zeta$ 1- $\Psi\alpha$ 2- $\Psi\alpha$ 1- α 2- α 1- θ 1 [5–7]. Alpha globin is an essential subunit of human hemoglobin from the sixth fetal week to adulthood. Fetal hemoglobin (α 2 γ 2) and adult hemoglobin (α 2 β 2) are produced by the combination of two chains of alpha globin with two chains of gamma and beta globin, respectively [8, 9]. Based on how many of the alpha globin chain genes are deleted, the clinical symptoms of alpha thalassemia vary from mild anemia to severe symptoms. If one, two, three,

Address for correspondence: Majid Arash, MD. Faculty of Medicine, Kermanshah University of Medical Sciences, Kermanshah, Iran Phone: 989900994024 E-mail: majidarashbioc@gmail.com ORCID: 0009-0006-9590-437X

Submitted: August 27, 2024 Revised: October 08, 2024 Accepted: October 09, 2024 Available Online: November 12, 2024 OPEN ACCESS This is an open access article under the CC BY-NC license (http://creativecommons.org/licenses/by-nc/4.0/).



) ()()

or four of the alpha globin genes are deleted, alpha thalassemia trait, silent carrier state, hemoglobin H disease, and Hydrops fetalis syndrome occur, respectively. The frequency of alpha thalassemia has been studied in different parts of the world, with varying frequencies in different regions of countries such as Iran, Morocco, China, and Syria [10–13].

Alpha Thalassemia Phenotypes

Alpha thalassemia phenotypes are divided into four categories: Alpha thalassemia silent carrier, Alpha thalassemia trait, Hemoglobin H disease (Hb H), and Hydrops fetalis.

Table 1 shows the characteristics of alpha thalassemia types.

Classification of Alpha Thalassemia Based on the Number of Deleted Genes

Different types of deletional mutations with different frequencies have been investigated in various parts of the world [14–19]. Some of these mutations cause the deletion of one of the two alpha globin genes, while others cause the deletion of both genes [14–21].

Single Gene Deletions

The most common deletional mutations in alpha thalassemia delete one of the genes of the alpha globin chain, which leads to a mild alpha thalassemia phenotype (- $\alpha/\alpha\alpha$). Reciprocal recombination, which occurs between highly similar regions called Z-boxes, leads to the deletion of 3.7 kb, which only includes one alpha gene (- $\alpha^{3.7}$). In contrast, recombination between the mismatched X-boxes results in the deletion of 4.2 kb (- $\alpha^{4.2}$). These recombinations lead to the creation of chromosomes containing three alpha globin genes [22, 23].

Double Gene Deletions

Several $\alpha 0$ defects that delete both alpha globin genes have been observed in different populations and are named according to their geographic region. These deletions include Southeast Asian (--^{SEA}), Mediterranean (--^{MED}), Filipino (--^{FIL}), Thai (--^{THAI}), and 20.5 kb type (-(α)^{20.5}) [24, 25]. Partial or complete deletion of both alpha globin genes in the cis state leads to the absence of alpha globin chain synthesis in the body. Homozygotes with these deletions have hydrops fetalis syndrome. Many deletions also remove ζ and α globin genes. Individuals in the heterozygous state appear to develop normally, but survival is unlikely for individuals in the homozygous state even in the early months or days of pregnancy, as neither $\alpha 2\gamma 2$ nor $\zeta 2\gamma 2$ hemoglobin can be produced. It is worth mentioning that rare deletions causing $\alpha 0$ thalassemia delete the regulatory region 40–50 kb upstream of the alpha globin gene cluster, leaving the alpha globin chain genes intact.

Detailed analysis of the deletional mutation $-(\alpha)^{20.5}$ reveals the importance of providing a more precise description of the 5' end of this deletion from the alpha globin genes. This is especially crucial because different phenotypes are associated with defects in the alpha globin gene cluster. The first cases of alpha-a0 thalassemia deletion, which deleted the entire alpha globin gene set, described a rare condition characterized by Hb H disease and congenital mental retardation [26]. While the --SEA deletional mutation is the most common cis deletion in alpha globin chains in Southeast Asia, several other deletions have also been reported in other scientific papers. --THAI and --FIL deletional mutations are common among some Southeast Asian populations. These common deletional mutations, which were first described by Fischel Ghodsian and colleagues, cover about 30-38 kilobases and delete the zeta2 globin gene as well as alpha globin genes located on chromosome 16 [27].

New Deletional Mutations

-α^{6.9} deletion

This deletion mutation, first observed in China, causes the deletion of the a2 gene but leaves the a1 gene intact (NG-000006.1.29785–36746 del 6962bp). The researchers who found this mutation initially identified alpha thalassemia by examining the phenotype of the subjects. Then, using the Multiplex Ligation-dependent Probe Amplification (MLPA) method, they evaluated the new mutation, and through DNA sequencing and bioinformatics analysis, confirmed the mutation's existence [28]. If this deletional mutation is combined with a single deletion, it causes alpha thalassemia trait, and if it is combined with a double deletional mutation, it causes Hb H disease.

--^{27.2} deletion

In a study, researchers in China discovered this new mutation after examining a young woman in pre-pregnancy screening who had hypochromic and microcytic hematological features [29]. This mutation causes the deletion of both alpha globin genes located on chromosome 16, and if it is combined with another double deletional mutation, it causes hydrops fetalis.

Table 1. Characteristics of alpha thalassemia types			
Name of disease	Number of deleted genes	Genetic condition of the disease	Severity of clinical symptoms
Silent carrier	One	-α/αα	Usually mild or no symptoms
Trait	Тwo	/aa	
-α/-α	Usually mild anemia		
Hb H	Three	/-α	mild to severe
Hydrops fetalis	Four	/	Severe symptoms

$(\alpha \alpha)^{ZRX}$ deletion and --³³⁶ deletion

These deletions, which cause the deletion of both alpha globin chains, were observed in two Chinese families. In the patient with the -336 deletion, a spontaneous mutation was identified, as neither parent had this mutation. Both carriers had symptoms of microcytic hypochromia, a characteristic of people with α^0 -thalassemia [30].

--^{GB} deletion

This deletion mutation, first observed in Malaysia, results in the deletion of both genes of the alpha globin chain (α^0 -thalassemia). Most individuals with this mutation are of Malay descent. The main risk of this mutation, when accompanied by a single or double deletion, is Hb H disease or hydrops fetalis, respectively [31].

(αα)^{FJ} deletion

This deletion mutation, which caused the deletion of 91.5 kilobases in the alpha globin gene cluster in a family from China, deletes both α 1 and α 2 genes. In areas with a high frequency of alpha thalassemia, this mutation should be investigated [32].

$-\alpha^{6.3}$ deletion and $-\alpha^{27.6}$ deletion

These mutations, identified in two Chinese patients, delete one of the alpha globin chains (α +-thalassemia). Like single deletion mutations such as - $\alpha^{3.7}$ and - $\alpha^{4.2}$, they are accompanied by double deletion mutations, causing Hb H disease [33].

--^{CR} deletion

This deletion mutation, found in Thailand, deletes 44.6 kilobases of the alpha globin gene cluster. There is limited information on the effects of this mutation, especially in combination with other types of thalassemia. Hypochromic and microcytic blood phenotypes have been observed in carriers [34].

--^{14.9} deletion

This mutation, discovered in China, deletes both alpha globin chains. Affected individuals had abnormal blood parameters, and after screening for 23 types of common mutations, this specific mutation was identified [35].

107 kilobase deletion

In a patient suffering from mild anemia, screening for thalassemia was performed by Gap-PCR and PCR-reverse dot blot methods. Following additional investigations, this deletional mutation was identified [36].

Common Diagnostic Methods in Identifying Deletional Mutations in Alpha Thalassemia Disease

For a long time, methods such as loop-mediated isothermal amplification, Multiplex Ligation-dependent Probe Amplification (MLPA), and single-tube multiplex polymerase chain reaction (PCR) have been used worldwide in the diagnosis of alpha thalassemia. However, new methods with greater speed and accuracy are also needed, as they can identify many genetic mutations in a short period of time [37]. In addition to traditional methods that have been used for years by diagnostic centers, new methods have also been introduced in recent years [38–40].

Conclusion

Considering the high prevalence of alpha thalassemia worldwide, as well as the complications of this disease, particularly in cases of Hb H disease and hydrops fetalis, it is advisable to screen individuals suspected of having this condition. Following diagnosis, genetic counseling should be provided to inform patients about the risks in pregnancy and prevent the birth of infants with severe alpha thalassemia. In areas where rare mutations are present, these mutations should also be studied.

Authorship Contributions: Concept – M.A.; Design – M.A., M.G.B.; Supervision – M.A.; Data collection &/or processing – M.A., M.G.B.; Analysis and/or interpretation – M.A.; Literature search – M.A.; Writing – M.A.; Critical review – M.A., M.G.B.

Conflict of Interest: The authors declare that there is no conflict of interest.

Use of Al for Writing Assistance: No Al technologies utilized.

Financial Disclosure: The authors declared that this study has received no financial support.

Peer-review: Externally peer-reviewed.

References

- Higgs DR, Vickers MA, Wilkie AO, Pretorius IM, Jaman AP, Weatherall DJ. A review of the molecular enetics of the human alpha-globin gene cluster. Blood 1989;73(5):1081–104. [Cross-Ref]
- 2. Kazazian HH Jr. The thalassemia syndromes: Molecular basis and prenatal diagnosis in 1990. Semin Hematol 1990;27(3):209–28.
- Ko TM, Chen TA, Hsieh MI, Tseng LH, Hsieh FJ, Chuang SM, et al. Alpha-thalassemia in the four major aboriginal groups in Taiwan. Hum Genet 1993;92(1):79–80. [CrossRef]
- Ko TM, Tseng LH, Kao CH, Lin YW, Hwa HL, Hsu PM, et al. Molecular characterization and PCR diagnosis of Thailand deletion of α-globin gene cluster. Am J Hematol 1998;57(2):124– 30. [CrossRef]
- 5. Lauer J, Shen CK, Maniatis T. The chromosomal arrangement of human alpha-like globin genes: sequence homology and alpha-globin gene deletions. Cell 1980;20(1):119–30. [CrossRef]
- Hardison RC, Sawada I, Cheng JF, Shen CK, Schmid CW. A previously undetected pseudogene in the human alpha globin gene cluster. Nucleic Acids Res 1986;14(4):1903–11. [CrossRef]
- Hsu SL, Marks J, Shaw JP, Tam M, Higgs DR, Shen CC, et al. Structure and expression of the human theta 1 globin gene. Nature 1988;331(6151):94–6. [CrossRef]
- 8. Liebhaber SA, Cash FE, Ballas SK. Human alpha-globin gene expression. The dominant role of the alpha 2-locus in mRNA and protein synthesis. J Biol Chem 1986;261(32):15327–33. [CrossRef]

- Higgs DR, Weatherall DJ. Alpha-thalassemia. Curr Top Hematol 1983;4:37–97.
- Asghari AM, Pourreza N, Ramezanpour S, Baghersalimi A, Enshaei M, Askari M, et al. An analysis of the distribution and spectrum of alpha thalassemia mutations in Rasht City, North of Iran. Front Pediatr 2023;11:1039148. [CrossRef]
- Laghmich A, Alaoui Ismaili FZ, Barakat A, Ghailani Nourouti N, Khattab M, Bennani Mechita M. Alpha-thalassemia in North Morocco: Prevalence and molecular spectrum. Biomed Res Int 2019;2019:2080352. [CrossRef]
- Yu Y, Yang Z, Yu H, Sun R, Zhu Y, Guo F. High prevalence of thalassemia with a novel α-thalassemia mutationin in Baoan populations in Guangdong province, China. Eur J Obstet Gynecol Reprod Biol 2020;255:263–5. [CrossRef]
- 13. Murad H, Moassas F, Ali B, Katranji E, Mukhalalaty Y. The spectrum of α-thalassemia mutations in Syrian Patients. Hemoglobin 2023;47(6):245–8. [CrossRef]
- Husna N, Handayani NSN. Molecular and haematological characteristics of alpha-thalassemia deletions in Yogyakarta Special Region, Indonesia. Rep Biochem Mol Biol 2021;10(3):346– 53. [CrossRef]
- Rizo-de la Torre LDC, Rentería-López VM, Sánchez-López JY, Magaña-Torres MT, Ibarra-Cortés B, Perea-Díaz FJ. Molecular and hematological analysis of alpha- and beta-thalassemia in a cohort of Mexican Patients. Genet Test Mol Biomark 2021;25(3):247–52. [CrossRef]
- 16. Adekile A, Sukumaran J, Thomas D, D'Souza T, Haider M. Alpha thalassemia genotypes in Kuwait. BMC Med Genet 2020;21(1):170. [CrossRef]
- 17. Zhu Y, Shen N, Wang X, Xiao J, Lu Y. Alpha and beta-thalassemia mutations in Hubei area of China. BMC Med Genet 2020;21(1):6. [CrossRef]
- Wang Z, Sun W, Chen H, Zhang Y, Wang F, Chen H, et al. Prevalence and molecular spectrum of α- and β-globin gene mutations in Hainan, China. Int J Hematol 2021;114(3):307–18. [CrossRef]
- 19. Lam TT, Nguyen DT, Le QT, Nguyen DA, Hoang DT, Nguyen HD, et al. Combined gap-polymerase chain reaction and targeted next-generation sequencing improve α- and β-thalassemia carrier screening in pregnant women in Vietnam. Hemoglobin 2022;46(4):233–9. [CrossRef]
- 20. He S, Li D, Yi S, Huang X, Zhou C, Chen B, et al. Molecular characterization of α - and β -thalassaemia among children from 1 to 10 years of age in Guangxi, a multi-ethnic region in Southern China. Front Pediatr 2021;9:724196. [CrossRef]
- Peng Q, Zhang Z, Li S, Cheng C, Li W, Rao C, et al. Molecular epidemiological and hematological profile of thalassemia in the Dongguan Region of Guangdong Province, Southern China. J Clin Lab Anal 2021;35(2):e23596. [CrossRef]
- 22. Embury SH, Miller JA, Dozy AM, Kan YW, Chan V, Todd D. Two different molecular organizations account for the single alpha-globin gene of the alpha-thalassemia-2 genotype. J Clin Invest 1980;66(6):1319–25. [CrossRef]
- 23. Higgs DR. The molecular basis of α-thalassemia. Cold Spring Harb Perspect Med 2013;3(1):a011718. [CrossRef]

- 24. Weatherall DJ, Clegg JB. The thalassaemia syndromes. 4th ed. Oxford: Blackwell Science;2001. [CrossRef]
- 25. Chui DH, Waye JS. Hydrops fetalis caused by α-thalassemia: An emerging health care problem. Blood 1998;91:2213–22. [Cross-Ref]
- Higgs DR. The molecular basis of alpha -thalassemia. In Steinberg MH, Forget BG, Higgs DR, Nagel RL, editors. Disorders of Hemoglobin, 2nd ed. Cambridge: Cambridge University Press; 2009.
- Fischel-Ghodsian N, Vickers MA, Seip M, Winichagoon P, Higgs DR. Characterization of two deletions that remove the entire human zeta-alpha globin gene complex (- -THAI and - -FIL). Br J Haematol 1988;70(2):233–8. [CrossRef]
- 28. Zhuang J, Tian J, Wei J, Zheng Y, Zhuang Q, Wang Y, et al. Molecular analysis of a large novel deletion causing α+-thalassemia. BMC Med Genet 2019;20(1):74. [CrossRef]
- 29. Wang G, Zou S, Li J, Wang X, Wu H, Tao Z, et al. The diagnosis and molecular analysis of a novel 27.2 kb deletion causing α0-thalassemia. Clin Biochem 2023;116:20–23. [CrossRef]
- 30. Jiwu L, Manna S, Ying Z, Youqing F, Haiyang C, Wanfang X, et al. Two large novel alpha-globin gene cluster deletions causing alpha(0)-thalassemia in two Chinese families. Gene 2022;840:146767. [CrossRef]
- 31. Yasin NM, Abdul Hamid FS, Hassan S, Mat Yusoff Y, Mohd Sahid EN, Esa E. Characterization of new alpha zero (α0) thalassaemia deletion (--GB) among Malays in Malaysian population. Diagnostics Basel 2023;13(20):3286. [CrossRef]
- 32. Xu L, Chen M, Zheng J, Zhang S, Zhang M, Chen L, et al. Identification of a novel 91.5 kb-deletion (αα)FJ in the α-globin gene cluster using single-molecule real-time (SMRT) sequencing. J Matern Fetal Neonatal Med 2023;36(2):2254890. [CrossRef]
- 33. Ke Wang, Ting Yi, Wen-Tao Wu, Juan Lu, Li-Na He, Hong-Ping Zhou, et al. (2022) Investigation of the distribution of thalassemia in children in Jiangxi Province, the People's Republic of China. Hemoglobin 2022;46(5):272–6. [CrossRef]
- 34. Ruengdit C, Khamphikham P, Jinorose N, Pornprasert S. Hb Bart's hydrops fetalis syndrome and Hb H disease caused by deletional Chiang Rai (- -CR) α0-Thalassemia in two unrelated Thai families. Hemoglobin 2021;45(2):75–9. [CrossRef]
- 35. Yuan Y, Zhou X, Deng J, Zhu Q, Peng Z, Chen L, et al. Case report: Long-read sequencing identified a novel 14.9-kb deletion of the α-globin gene locus in a family with α-thalassemia in China. Front Genet 2023;14:1156071. [CrossRef]
- 36. Li Y, Liang L, Guo W, Wu X, Qin T, Tian M. Identification of a novel 107 kb deletion in the alpha-globin gene cluster using third-generation sequencing. Clin Biochem 2023;113:36–9. [CrossRef]
- 37. Vijian D, Wan Ab Rahman WS, Ponnuraj KT, Zulkafli Z, Mohd Noor NH. Molecular detection of alpha thalassemia: A review of prevalent techniques. Medeni Med J 2021;36(3):257–69. [CrossRef]
- Ha Vuong VV, Nguyen PD, Thi NN, Le Thi P, Minh Nguyet DT, Nguyen MH, et al. Application of short tandem repeats (STRs)

in the preimplantation genetic diagnosis (PGD) of α -thalassemia. Taiwan J Obstet Gynecol 2024;63(3):375–80. [CrossRef]

39. Sani A, Tian Y, Zha G, Xiao X, Shah S, Idrees Khan M, et al. A high stable sample loading for analysis of adult alpha-thalassemia via the improved microarray isoelectric focusing of Hb species. J Chromatogr B Analyt Technol Biomed Life Sci 2024;1244:124238. [CrossRef]

40. Li Y, Ye J, Liang L, Tan X, Zheng L, Qin T, et al. Detection of α-thalassemia South-East Asian deletion based on a fully integrated digital polymerase chain reaction system DropXpert Hematol 2024;29(1):2365596. [CrossRef]